

Biological activities and potential functional optimization strategies of corosolic acid: a review

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Abstract

Corosolic acid, a naturally occurring pentacyclic triterpenic acid, is widely recognized for its broad spectrum of biological activities, particularly its anti-diabetic properties, making it a popular ingredient in dietary supplements for regulating blood sugar levels. Beyond its anti-diabetic effects, recent studies have revealed its therapeutic potential in areas such as anti-cancer, anti-inflammatory, and antibacterial activities. However, its clinical application is hindered by poor water solubility and low bioavailability due to its molecular structure. This review systematically examines the pharmacological activities of corosolic acid, emphasizing its mechanisms of action in disease intervention. Emerging strategies to overcome its inherent limitations, including chemical modifications, microbial transformations, and advanced delivery systems, are also discussed. Notably, some chemical derivatives exhibit α -glucosidase inhibition with IC₅₀ values half that of corosolic acid. Microbial transformations have been shown to enhance its bioavailability while reducing cancer cell toxicity. Additionally, corosolic acid-based delivery systems have demonstrated significant improvements in solubility, stability, and biological activity. By consolidating current insights into its functional properties and biological activity enhancement methods, this review aims to emphasize the practical application values in food and medicine and the future development of corosolic acid as a versatile bioactive compound.

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Introduction

Corosolic acid, a ursane-type pentacyclic triterpenic acid, is a naturally occurring compound widely distributed in various plants, including *banaba*^[1], loquat leaves^[2], guava^[3], rosemary^[4], allspice, clove^[5], cranberry^[6], sweet osmanthus^[7], and black cherry^[8]. Early studies focused on determining its chemical structure and its biological function in plants. More recently, corosolic acid has garnered significant attention from the scientific community, particularly in the field of diabetes research, due to its widespread availability and diverse biological activities^[9]. Corosolic acid and its structural analogs, including ursolic, oleanolic, maslinic, asiatic, glycyrrhetic, madecassic, moronic, pomolic, boswellic, and betulinic acids, have been reported to exhibit various biological activities, such as anti-diabetic, anti-cancer, anti-inflammatory, and antimicrobial effects^[9–11]. Among these, corosolic acid has demonstrated a stronger potential anti-diabetic effect compared to other pentacyclic triterpenic acids^[12,13].

Many studies have highlighted the therapeutic potential of corosolic acid in various areas. Its anti-diabetic properties are primarily attributed to the inhibition of enzymes such as α -amylase^[13], α -glucosidase^[12], and diabetes-related non-receptor protein tyrosine phosphatases^[14], as well as its ability to enhance insulin signaling pathways^[15]. As for the application in the food field, corosolic acid has been used as a key ingredient in dietary supplements to support glucose metabolism and maintain healthy blood sugar levels^[1].

With the increasing body of research on corosolic acid, its pharmacological effects beyond anti-diabetes have been increasingly

revealed. In addition to its anti-diabetic properties, corosolic acid has demonstrated multiple other biological activities. Its anti-cancer effects have been reported through mechanisms such as inducing ferroptosis^[16,17], promoting apoptosis in cancer cells^[18,19], and inhibiting cancer cell migration and invasion^[20]. Furthermore, corosolic acid exhibits anti-inflammatory effects by suppressing pro-inflammatory cytokines and key inflammatory pathways^[21,22]. Additionally, reports indicate that corosolic acid acts as a biofilm inhibitor^[23], demonstrating antibacterial properties and enhancing the bactericidal activity of antibiotics^[24], thereby further expanding its application potential. These diverse bioactivities suggest that corosolic acid holds significant promise as both a dietary supplement and a pharmaceutical adjunct.

Corosolic acid, as a triterpenoid compound, is biosynthesized from 2,3-oxidosqualene, a common C30 acyclic precursor produced through the mevalonate pathway^[25]. The molecular structure of corosolic acid consists of a side chain containing both an alcohol group and a carboxylic group, as well as a cyclic triterpenoid skeleton. Corosolic acid suffers from limited solubility in water and low bioavailability due to its lack of polar functional groups and its long lipophilic backbone, which restrict its practical applications in food and pharmaceutical fields^[26]. To overcome these limitations, several studies have explored strategies to enhance its functionality and bioavailability, including chemical modifications^[27], microbial transformations^[28], and delivery systems such as liposomes^[29], and nanoemulsion^[30].

This review provides an overview of recent advances in the functional activities of corosolic acid and discusses emerging strategies aimed at enhancing its functional properties and therapeutic

efficacy (Fig. 1). It aims to contribute to the growing body of knowledge on corosolic acid, highlighting its potential as a valuable bioactive compound and guiding future research into its applications in food and pharmaceuticals.

Pharmacological activity

Anti-diabetic properties

Corosolic acid initially attracted attention for its potential therapeutic effects on diabetes, earning it the nickname 'phyto-insulin' or 'botanical insulin'^[31]. Its impact on glucose metabolism and insulin sensitivity has made it a compound of considerable interest in diabetes research. Corosolic acid has shown a promising impact in managing diabetes, particularly type 2 diabetes, through several mechanisms that enhance insulin sensitivity and regulate glucose metabolism^[32]. One of its primary therapeutic actions is the inhibition of α -glucosidase activity, primarily governed by hydrophobic interactions, which delays glucose absorption and helps manage postprandial hyperglycemia^[12,13,33,34] (Fig. 2). Corosolic acid inhibits both α -glucosidase and α -amylase in a non-competitive, reversible manner, with an IC₅₀ value of 1.35×10^{-5} mol/L for α -glucosidase inhibition—less than half the value of the positive control acarbose, demonstrating a higher inhibitory effect on α -glucosidase^[12,13]. Furthermore, when combined with myricetin, corosolic acid exhibits a synergistic effect on α -glucosidase inhibition, further supporting its role in glucose regulation^[12].

In addition to its effects on glucose absorption, corosolic acid has been shown to enhance insulin signaling pathways. It improves insulin sensitivity in high-fat diet-induced diabetic mice by

modulating the phosphorylation of Insulin Receptor Substrate-1 and its downstream target, Protein Kinase B^[15,25]. This modulation enhances the insulin signaling cascade, contributing to improved glucose homeostasis. Corosolic acid also activates AMP-activated protein kinase, a key regulator of energy metabolism, which inhibits inflammation and reduces insulin resistance^[15,35]. Furthermore, studies have demonstrated that corosolic acid reduces the phosphorylation of I κ B kinase β and suppresses the expression of pro-inflammatory cytokines, alleviating adipose tissue inflammation—a key contributor to insulin resistance^[15].

In vitro studies also highlight the role of corosolic acid in glucose uptake. It enhances glucose uptake in L6 myotubes by inhibiting several diabetes-related non-receptor protein tyrosine phosphatases and promoting the translocation of glucose transporter type 4 in CHO/hIR cells, suggesting its potential to improve cellular glucose transport^[14,36], further contributing to the regulation of glucose metabolism. The anti-diabetic effects of corosolic acid extend to animal models of type 2 diabetes, such as the KK-Ay mouse, a model of insulin resistance and hyperglycemia^[37]. In these mice, a single oral dose of corosolic acid (2 mg/kg body weight) significantly reduced blood glucose levels within 4 h, with sustained effects over 2 weeks^[38]. Moreover, corosolic acid treatment lowered plasma insulin levels and improved glucose tolerance in the insulin tolerance test, indicating a reduction in insulin resistance. One study investigated the effects of corosolic acid in a randomized, double-blind, crossover trial involving 14 middle-aged, non-diabetic men with impaired fasting glucose tolerance. The results suggested that corosolic acid may enhance insulin action and improve glucose metabolism in prediabetic individuals^[39].

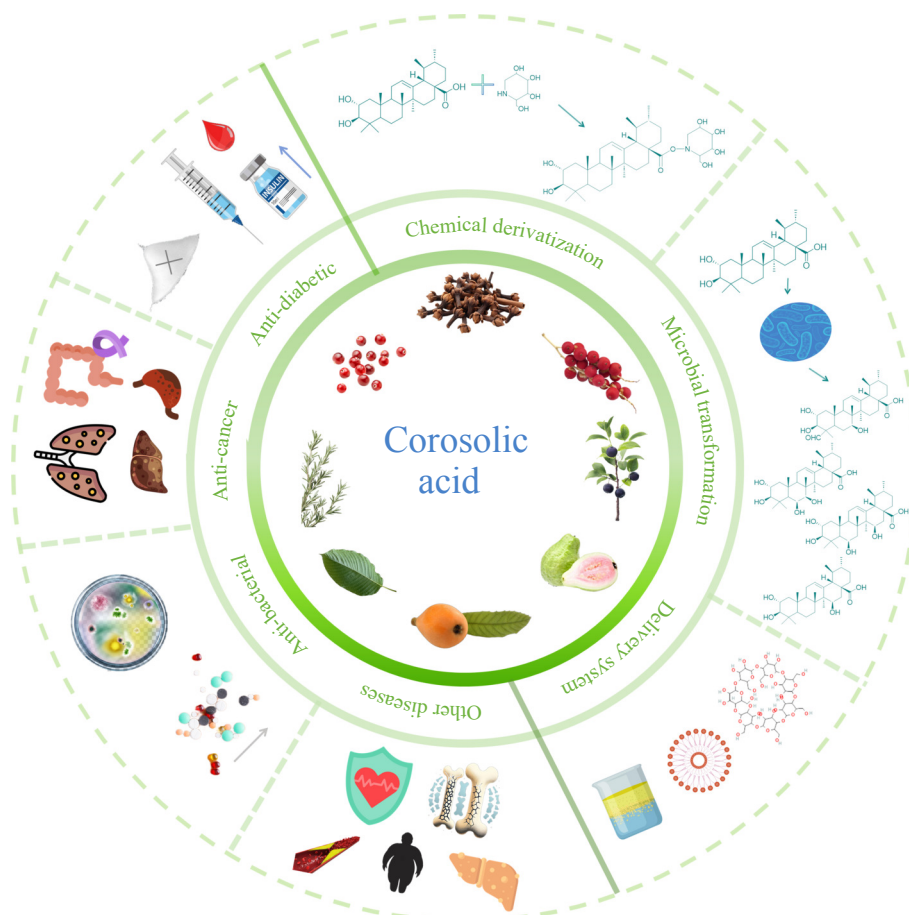


Fig. 1 Biological activities of corosolic acid and strategies for enhancing the potential efficacy.

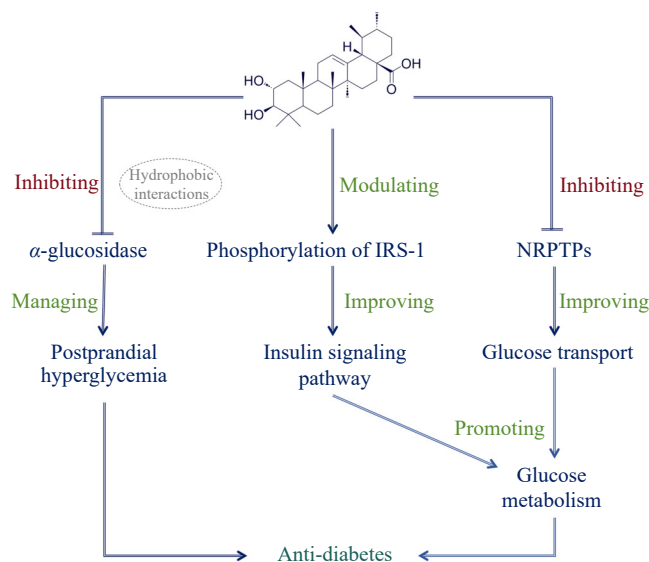


Fig. 2 Proposed mechanisms of corosolic acid in diabetes inhibition. IRS-1: Insulin Receptor Substrate-1; NRPTs: Non-Receptor Protein Tyrosine Phosphatases.

Overall, the ability of corosolic acid to improve glucose metabolism and insulin sensitivity, without inducing anti-insulin antibodies, supports its potential as a therapeutic agent for managing type 2 diabetes.

Anti-cancer effect

Apart from its anti-diabetic properties, corosolic acid has demonstrated significant potential as an anti-cancer agent through various mechanisms, positioning it as a promising compound for cancer treatment, as highlighted by increasing studies on its anti-cancer activity^[40,41]. Table 1 summarizes the proposed inhibitory mechanisms of corosolic acid against different types of cancer. One of its most notable anti-cancer properties is its ability to induce ferroptosis, a form of regulated cell death characterized by iron-dependent lipid peroxidation^[42,43]. Experimental results have further confirmed the role of ferroptosis in the anti-cancer action of corosolic acid, as ferroptosis inhibitors can effectively rescue corosolic acid-induced lung cancer cell death, while corosolic acid itself suppresses epithelial-mesenchymal transition and induces ferroptosis independently of the inhibitors^[16]. Corosolic acid has also been reported to sensitize liver cancer cells to ferroptosis^[17] and reduce the proliferation and invasion of pancreatic cancer cells by inducing SAT1-mediated ferroptosis^[2], marking it as a novel approach in cancer therapy. Additionally, corosolic acid has demonstrated notable cytotoxicity

under hypoxic conditions, commonly found in tumors, suggesting its potential as an adjunctive treatment for cancer^[44]. A concentration of 4 μ M corosolic acid significantly inhibits the growth of A549 lung cancer cells^[44].

Effect on liver cancer

Corosolic acid can target key signaling pathways directly or exert anticancer effects on various cancers through synergistic interactions with other substances. In liver cancer, corosolic acid interacts with the ATP-binding pocket of VEGFR2 kinase, thereby inhibiting its activity and downregulating the VEGFR2/Src/FAK/cdc42 axis, which is essential for liver cancer cell migration and invasion^[45]. Additionally, corosolic acid suppresses the CDK19/YAP/o-GlcNAcylation pathway, a mechanism critical for the proliferation of high-glucose cancer cells, thus inhibiting liver cancer cell growth^[46]. Furthermore, corosolic acid activates the PERK-eIF2 α -ATF4 pathway, inducing endoplasmic reticulum stress-mediated apoptosis in hepatocellular carcinoma cells^[19]. *In vitro* studies have demonstrated that corosolic acid inhibits the proliferation of liver cancer cells, including Bel-7404 and HepG2, in a dose-dependent manner, resulting in G2/M cell cycle arrest and promoting cell death^[58].

Effect on gastric cancer

Molecular docking studies indicate that corosolic acid interacts with key targets, including PIK3CA, PTGS2, AKT1, and CASP3, modulating the PI3K-AKT signaling pathway (hsa04151) to exert anti-cancer effects^[48]. Additionally, corosolic acid enhances the sensitivity of gastric cancer cells to the chemotherapy drug 5-fluorouracil by activating AMP-activated protein kinase (AMPK)^[47]. Through regulation of the AMPK-mTOR signaling pathway, corosolic acid inhibits mammalian target of rapamycin (mTOR) activity, thereby modulating cancer cell growth and apoptosis^[59]. Furthermore, it has been shown to suppress HER2 gene expression, resulting in cell cycle arrest and apoptosis, which provides a theoretical basis for its application in treating HER2-positive gastric cancer^[18]. In SNU-620 gastric cancer cells, corosolic acid further potentiates the anticancer efficacy of 5-fluorouracil by inhibiting mTOR signaling, underscoring its potential as an adjuvant therapy^[60].

Effect on colon cancer

Corosolic acid demonstrates significant potential in combating colon cancer through multiple mechanisms. One study indicates that corosolic acid inhibits the proliferation of APC-mutant colon cancer cells by promoting N-terminal phosphorylation of β -catenin, leading to its subsequent proteasomal degradation^[49]. Additionally, corosolic acid exhibits a strong binding affinity to COX-2 and effectively reduces its expression, highlighting its dual anti-inflammatory and anticancer properties^[50]. *In vitro* studies further reveal that corosolic acid induces apoptosis in CT-26 cells by activating

Table 1. Proposed inhibitory mechanisms of corosolic acid against different types of cancer.

Cancer	Proposed inhibitory mechanisms	Ref.
Lung cancer	Inhibition of epithelial-mesenchymal transition, promoting ferroptosis	[16,44]
Liver cancer	Promoting ferroptosis, downregulating the VEGFR2/Src/FAK/cdc42 axis, suppressing the CDK19/YAP/o-GlcNAcylation pathway, activating the PERK-eIF2 α -ATF4 pathway	[19,45,46]
Gastric cancer	Modulating the PI3K-AKT signaling pathway, influencing AMPK-mTOR signaling pathway, inhibiting HER2 gene expression	[18,47,48]
Colon cancer	Promoting N-terminal phosphorylation of β -catenin, reducing COX-2 expression, activating caspase-3	[49–51]
Pancreatic cancer	Promoting ferroptosis, inhibiting the JAK2/STAT3 signaling pathway	[2,52]
Kidney cancer	Reducing MMP2 expression and stimulates ERK1/2 phosphorylation, stimulating ERK1/2 phosphorylation	[20,42]
Breast cancer	Reducing the phosphorylation levels of JAK2 and STAT3 proteins and activating apoptosis-related caspases	[53]
Bladder cancer	Inhibit DNA synthesis, downregulating cell cycle regulators, triggering autophagy-related pathways	[54]
Ovarian cancer	Inhibiting macrophage-epithelial interactions	[55]
Oral cancer	Inhibits the expression of MMP1	[56]
Bone cancer	Disrupting mitochondrial membrane potential, increasing the Bax/Bcl-2 ratio, releasing cytochrome c, activating caspases-8, -9, and -3	[57]

caspase-3. Furthermore, *in vivo* experiments using a mouse colon cancer model demonstrate that corosolic acid significantly reduces the final tumor volume, as well as the blood and lymphatic vessel density within the tumor^[51]. Similarly, the molecular targets and signaling pathways of ursolic acid contribute to apoptosis in colon cancer cell lines by up-regulating caspase-3, -8, and -9 and activating the phosphoinositide 3-kinase and MAPK/extracellular signal-regulated kinase pathways^[61].

Effect on other cancers

As research on corosolic acid progresses, its effects on various cancers are being increasingly explored. In pancreatic cancer, corosolic acid inhibits the JAK2/STAT3 signaling pathway, suppressing tumor progression^[52]. For renal cell carcinoma, it reduces MMP2 expression and stimulates ERK1/2 phosphorylation in 786-O and Caki-1 cells, thereby preventing cancer cell migration and invasion through interactions with MMP2^[20]. In breast cancer, corosolic acid induces apoptosis in MDA-MB-231 cells by downregulating JAK2 and STAT3 phosphorylation and activating apoptosis-related caspases^[53]. Furthermore, corosolic acid has shown potential in reversing tumor-promoter-induced DNA methylation changes in mouse epidermal JB6 P+ cells^[62].

In bladder cancer, corosolic acid suppresses TOP2A and LIG1 to inhibit DNA synthesis, downregulates cell cycle regulators, and triggers autophagy-related pathways involving proteins such as NBR1, TAXBP1, SQSTM1/P62, and UBB^[54]. Additionally, corosolic acid enhances the efficacy of traditional chemotherapy agents like paclitaxel, cisplatin, and doxorubicin by suppressing macrophage-epithelial interactions that activate epithelial ovarian cancer cells^[55]. In human oral squamous cell carcinoma, it significantly inhibits MMP1 expression and impairs cell migration and invasion, demonstrating synergistic effects when combined with siMMP1^[56]. The isomer of corosolic acid, maslinic acid, has also been reported to effectively inhibit the viability of bladder cancer cell lines (T24, TCCSUP, 253J, and RT4), with IC50 values ranging from 27 to 71 μ M^[63].

Corosolic acid also induces G2/M cell cycle arrest in CaSki cervical cancer cells in a dose-dependent manner^[64]. In MG-63 osteosarcoma cells, it disrupts mitochondrial membrane potential, increases the Bax/Bcl-2 ratio, releases cytochrome c, and activates caspases-8, -9, and -3, leading to apoptosis^[57]. Collectively, these findings highlight the versatility of corosolic acid as an anticancer agent, targeting diverse mechanisms across various cancer types.

Protective effects against some diseases

Corosolic acid has been investigated for its broader biological activities beyond its well-established effects on diabetes and cancer, highlighting its potential as a multifunctional dietary supplement or therapeutic agent. One notable function is its ability to protect against diabetic nephropathy, a common complication of diabetes. Corosolic acid has been shown to inhibit the proliferation of glomerular mesangial cells via the p38 MAPK and NADPH-mediated ERK1/2 signaling pathways, offering protection against diabetic kidney damage^[65].

Additionally, corosolic acid exerts significant cardioprotective effects, particularly in the context of ischemia-reperfusion injury^[66]. It mitigates oxidative stress and preserves myocardial mitochondrial structure and function. Studies have demonstrated that corosolic acid treatment significantly reduces markers of myocardial injury, such as CK-MB and LDH levels, in rats subjected to isoproterenol-induced myocardial damage^[67]. By decreasing lipid peroxidation and boosting endogenous antioxidant levels, corosolic acid improves cardiac function and protects against acute myocardial injury.

Corosolic acid also plays a role in protecting against heart failure and myocardial fibrosis. It attenuates myocardial infarction-induced cardiac fibrosis and dysfunction through modulation of inflammation and oxidative stress pathways, with AMPK α being a key regulator in this process^[68]. Moreover, corosolic acid has been found to protect against doxorubicin-induced cardiac toxicity by activating AMPK-dependent mechanisms, including the activation of TFEB, which enhances autophagy and mitigates cardiac damage^[69].

In vascular health, corosolic acid demonstrates potent vasorelaxant effects via activation of the NO/cGMP and H2S/ATP-sensitive potassium (KATP) channel pathways^[70]. It also alleviates pulmonary arterial hypertension (PAH)-induced vascular remodeling by inhibiting the PDGF-PDGF receptor β -STAT3/NF- κ B signaling axis, which is involved in PAH pathogenesis^[71]. Its structural analog, ursolic acid, has also been reported to maintain cardiovascular health by regulating the expression of vascular injury factors, promoting vascular endothelial cell proliferation, and enhancing angiogenesis^[72].

Beyond its cardiovascular effects, corosolic acid exhibits significant anti-osteoporotic properties. It inhibits the PI3K/AKT/mTOR signaling pathway, enhancing autophagy and preventing IL-1 β -induced degradation of extracellular matrix in cartilage^[73]. Additionally, corosolic acid reduces osteoclastogenesis and oxidative stress, offering protection against lipopolysaccharide-induced bone resorption^[74]. The function was also found in asiatic acid, which can inhibit osteoclast formation through the RANKL-activated NF- κ B or NFATc1 signaling pathways, thereby reducing bone resorption^[75].

In metabolic disorders, corosolic acid controls lipid metabolism and reduces oxidative stress. It effectively inhibits pancreatic lipase and cholesterol esterase, enzymes responsible for the breakdown of triglycerides and cholesterol, thereby regulating lipid levels in the bloodstream^[76]. Furthermore, corosolic acid improves hypertension, lipid metabolic abnormalities, oxidative stress, and inflammation in spontaneously hypertensive rats^[77], suggesting its potential to prevent atherosclerosis-related diseases.

Corosolic acid has also been found to improve glucose metabolism by promoting glucose uptake in 3T3-L1 adipocytes and inhibiting adipocyte differentiation through downregulation of key adipogenic markers, such as PPAR- γ and C/EBP- α ^[78]. This effect positions corosolic acid as a promising candidate for managing obesity-related conditions and metabolic syndrome.

Finally, the rising prevalence of obesity and diabetes has significantly contributed to the widespread occurrence of non-alcoholic fatty liver disease, which often progresses to hepatocellular carcinoma^[31]. Studies have shown that corosolic acid can markedly inhibit ethanol-induced cell apoptosis while increasing levels of tumor necrosis factor- α and reactive oxygen species, both *in vitro* and *in vivo*^[31,79]. Furthermore, corosolic acid may protect the liver from ethanol-induced damage through the regulation of MAPK signaling pathways and activation of autophagy^[79].

Antimicrobial properties

Corosolic acid has demonstrated significant potential as an antibacterial agent, with multiple studies highlighting its ability to inhibit the growth of various pathogenic bacteria, particularly those associated with hospital-acquired infections. Notably, corosolic acid exhibits stronger antimicrobial activity against *Staphylococcus aureus* (a Gram-positive bacterium) compared to *Escherichia coli* (a Gram-negative bacterium)^[30], which is the same as other triterpenic acids, such as ursolic, asiatic oleanolic, and betulinic acids^[80,81].

A key antimicrobial property of corosolic acid is its ability to prevent biofilm formation, a common feature of bacterial infections that contributes to antibiotic resistance^[82]. Corosolic acid has been identified as an effective biofilm inhibitor^[23,83]. It enhanced the

sensitivity of biofilm-producing *Pseudomonas aeruginosa* to treatment with tobramycin, a widely used antibiotic for resistant infections^[23], and improved the antibacterial activity of cefotaxime against *S. aureus*^[83], suggesting its potential as an adjunct in combination therapy.

In addition to inhibiting biofilm formation and enhancing antibiotic efficacy, corosolic acid plays a synergistic antibacterial role in other ways. Corosolic acid analogs (oleanolic and ursolic acids) have been reported to impact cell morphology and promote autolysis of bacterial cells by influencing peptidoglycan metabolism^[84]. Although less effective against Gram-negative bacteria compared to Gram-positive bacteria, it has been reported to enhance the bactericidal activity of antibiotics against *E. coli*. This effect is attributed to corosolic acid's inhibition of β -lactamase enzymes, such as KPC-2, which mediate antibiotic resistance in *E. coli*. By inhibiting β -lactamase activity, corosolic acid can restore *E. coli*'s sensitivity to carbapenem antibiotics^[24].

Additionally, corosolic acid has been identified as a promising natural product for the treatment of bee diseases, particularly for controlling infections caused by *Melissococcus plutonius* (European foulbrood) and *Bacillus larvae* (American foulbrood)^[85]. These Gram-positive bacterial diseases can be controlled using corosolic acid, further demonstrating its broad-spectrum antimicrobial activity.

In summary, corosolic acid's ability to inhibit bacterial growth, prevent biofilm formation, and restore antibiotic efficacy positions it as a promising plant-derived natural product in the fight against antibiotic-resistant bacterial infections and the development of alternative antimicrobial therapies.

Regarding anti-fungal activities, corosolic acid has demonstrated varying potency depending on the fungal species. Studies have reported a minimum inhibitory concentration of 12.5 $\mu\text{g/mL}$ against *Ascosphaera apis*, a fungal pathogen affecting honeybees^[86]. However, at a concentration of 100 $\mu\text{g/mL}$, corosolic acid exhibited only a 22.16% inhibition rate against *Sclerotinia sclerotiorum*^[87], indicating limited efficacy against this particular fungal strain.

Corosolic acid has also demonstrated notable antiviral activity. *In vitro* studies have shown that corosolic acid exhibits anti-respiratory syncytial virus and anti-herpes simplex virus type 1 activities, with IC₅₀ values of 12.5 μM , comparable to the positive control ribavirin, which has an IC₅₀ of 10 μM ^[88]. The antiviral mechanisms of corosolic acid may involve its interaction with the allosteric site of AMPK. By stimulating AMPK, corosolic acid can inhibit lipid biosynthesis, a process crucial for viral replication^[89], further supporting the potential of corosolic acid as an antiviral agent.

Functionality improvement

The poor water solubility and low bioavailability of corosolic acid are primarily attributed to the rigid skeleton and hydrophobic nature of its pentacyclic triterpenic structure^[90–93], which limits its application as both a dietary supplement and a therapeutic agent. To overcome these challenges, various strategies, including chemical derivatization, microbial transformation, and advanced delivery systems, have been explored. These approaches not only aim to improve the solubility and bioavailability of corosolic acid but also hold the potential to enhance its functional activity.

Chemical derivatization

Chemical modification of natural compounds is a crucial strategy to improve solubility, bioavailability, and biological activity. This approach offers flexibility in designing compounds with improved specificity and efficacy for targeted applications^[94]. The approach has been widely explored in other triterpenic acids, providing

valuable insights into structure-activity relationships. By modifying the C-2, C-3, C-11, C-12, and C-28 positions of the oleanolic acid scaffold, its anti-diabetic, anticancer, and antiviral activities can be enhanced^[90]. Similarly, the introduction of an amino group at the C-28 position of asiatic acid significantly enhances its anticancer activity against several cancer cell lines^[95]. These findings suggest that specific functional group modifications could be a key factor in optimizing the biological activity of corosolic acid. By introducing functional groups or conjugating the compound with bioactive moieties, chemical derivatives can enhance its pharmacological effects while addressing its inherent limitations^[96–97].

There was study that reported modifications at the C-28 position, such as the introduction of piperazine-L-amino acid complexes, have yielded derivatives with varied biological activities. However, the α -glucosidase inhibitory activity of these derivatives in DMSO and ethanol-water systems was lower than that of the parent compound^[98] (Fig. 3). In contrast, other studies have shown that derivatives containing piperazine units exhibited superior activity compared to their precursors. Molecular docking simulations further revealed that the incorporation of piperazine enhanced hydrogen bonding interactions with α -glucosidase, thereby improving the biological activity of the derivatives^[99]. The IC₅₀ values of some piperazine derivatives was half of corosolic acid. Additionally, the carbon chain length of the linker between triterpenic acid and 1-DNJ plays a critical role in hypoglycemic activity, with shorter linkers exhibiting stronger inhibition of α -glucosidase than longer ones.

Quinoline derivatives of corosolic acid have also demonstrated promising antitumor properties. For example, 7-aminoquinoline, 5-aminoquinoline, and 8-isoquinoline derivatives exhibited high cytotoxicity, strong tumor cell selectivity, and the ability to overcome drug resistance^[100]. The derivatization could decrease the drug resistance ratio more than twice. Glycosylation of corosolic acid with mono- and disaccharides improves its water solubility, with disaccharides showing superior solubility and α -glucosidase inhibitory activity compared to monosaccharides. However, their inhibitory activity (ranging from IC₅₀ = 428 μM to no inhibition) was still much lower than that of the parent compound (71 μM)^[101].

Furthermore, hybrid derivatives of corosolic acid have shown significant effects on tumor cell cycle progression at doses much lower than those required for the natural triterpenic precursors. Cationic derivatives, such as TPP⁺ and F16-triterpenoid conjugates, outperformed their parent compounds in inducing reactive oxygen species (ROS) and were more effective at reducing mitochondrial membrane potential in isolated rat liver mitochondria, highlighting the therapeutic potential of chemical derivatization to enhance the pharmacological properties of corosolic acid^[27].

Microbial transformation

The second method, microbial transformation, is an attractive approach for expanding the structural diversity of triterpenoids by enabling unique modifications such as methylation, epoxidation, regioselective hydroxylation, and methyl migration^[102–103].

Studies involving *Fusarium equiseti* and *F. solani* demonstrated selective oxidation of corosolic acid at the C-6, C-7, C-15, and C-21 positions, resulting in polar metabolites^[104] (Fig. 4). Although these modifications enhanced the water solubility of corosolic acid, hydroxylation at these positions significantly reduced its anticancer and α -glucosidase inhibitory activities. Similarly, metabolism by *Cunninghamella echinulata* and *C. blakesleeana* generated new hydroxylated and oxidized metabolites, which also exhibited diminished α -glucosidase inhibition^[28]. The study indicated that hydroxylation at the C-6 position and oxidation at the C-24 site of corosolic acid negatively impacted its α -glucosidase activity.

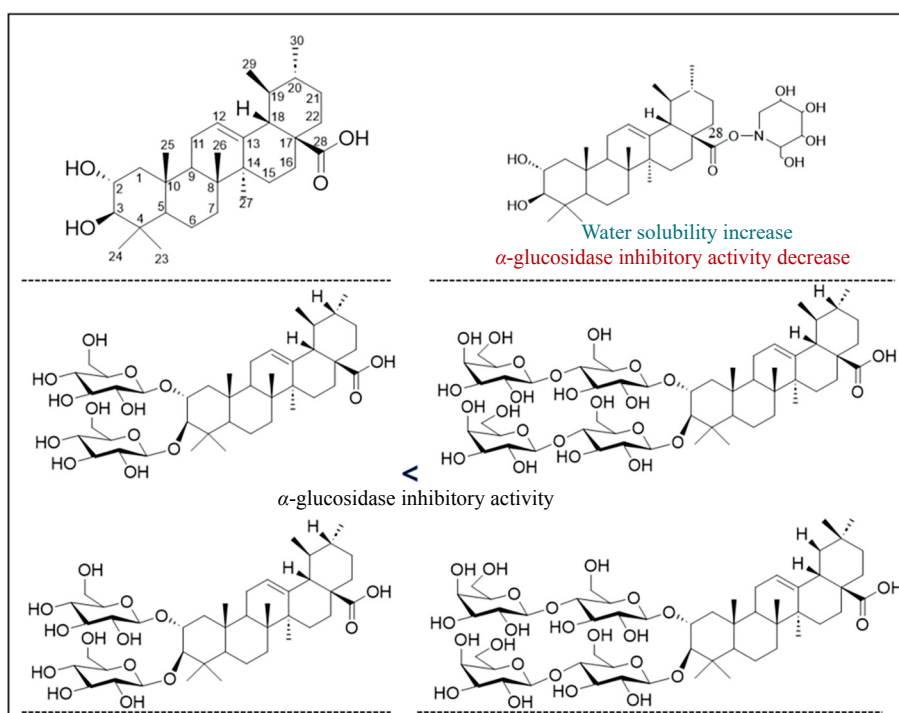


Fig. 3 Structural formula of corosolic acid and some chemical derivatives.

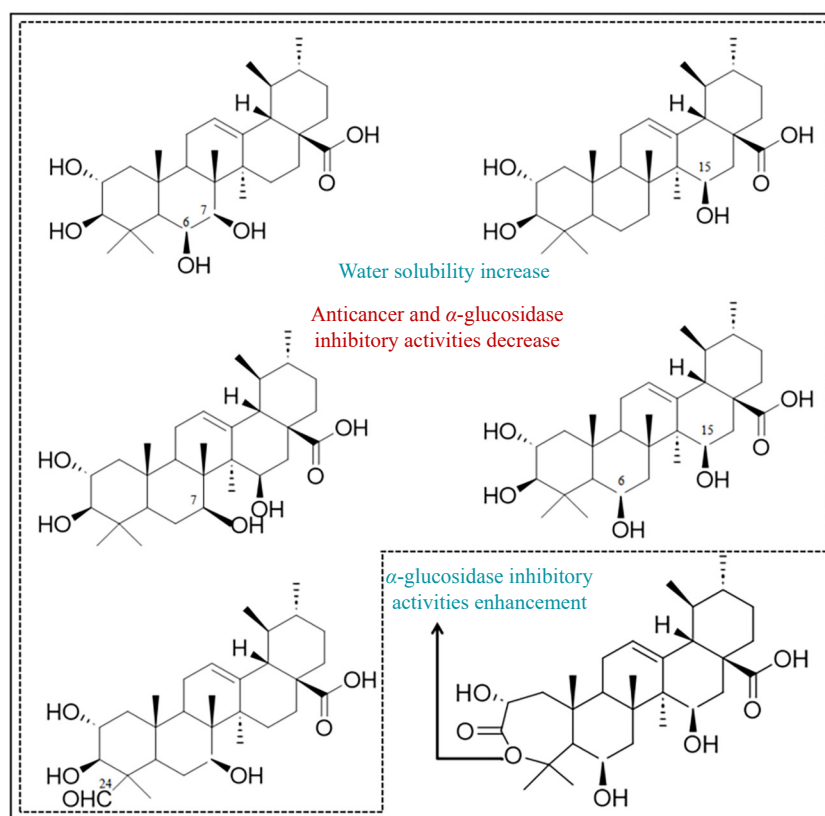


Fig. 4 Structural formula of some biotransformation products of corosolic acid.

In contrast, another study showed that while C-15, C-21, and C-30 hydroxylation and C-21 carbonylation of corosolic acid reduced its α -glucosidase inhibitory activity, lactonization within the A-ring could enhance this activity^[28]. Most microbial derivatives exhibited reduced toxicity toward cancer cells, likely due to decreased substrate toxicity resulting from microbial metabolism^[105].

Based on these studies, microbial transformation can improve the bioavailability of corosolic acid, but it generally decreases its α -glucosidase inhibitory activity. However, analyzing the effects of these structural changes on the compound's function may offer valuable insights into optimizing the modification of corosolic acid for therapeutic applications^[106].

Delivery system

Advancements in delivery systems have introduced novel strategies to enhance the solubility, stability, and bioavailability of triterpenic acids, including corosolic acid^[107–109]. The supramolecular properties of corosolic acid allow it to molecular self-assembly without structural modifications supporting subsequent application^[110–111]. For example, it self-assembles into vesicles or supramolecular gels in aqueous organic solvents^[112], demonstrating potential for applications such as drug encapsulation, controlled release, and fluorescence labeling. Additionally, corosolic acid can spontaneously form inclusion complexes with hydroxypropyl- β -cyclodextrin, significantly improving its water solubility and enhancing its bioavailability^[113].

Nanoemulsions of corosolic acid further enhance its functionality by reducing hydrophobicity, thus improving its antibacterial activity against Gram-positive bacteria^[30]. Corosolic acid structural analogs, oleanolic acid, and ursolic acid, have also been confirmed as stabilizers for the preparation of Pickering emulsions^[114,115]. This makes corosolic acid-based emulsions promising for applications in food, cosmetics, and pharmaceuticals. Cholesterol-free lipid nanoparticles derived from corosolic acid exhibit superior tumor cell uptake and endosomal membrane fusion capabilities, enabling more efficient cytoplasmic delivery of mRNA and siRNA^[116].

In anticancer applications, corosolic acid-based liposomes demonstrate several advantages over conventional cholesterol-based liposomes. For instance, doxorubicin-loaded corosolic acid-based liposomes enhance membrane fusion, and cellular uptake, and inhibit STAT3 activation and macrophage recruitment in the tumor microenvironment^[29]. Similarly, corosolic acid-loaded liposomes containing paclitaxel significantly improve its anticancer efficacy by overcoming tumor biological barriers, enhancing immunogenic cell death, and achieving satisfactory therapeutic outcomes^[117]. These studies highlight the potential of corosolic acid-based delivery systems in improving solubility, stability, antibacterial, and anticancer activities. Such systems offer promising alternatives to traditional drug carriers, opening avenues for wide-ranging applications in medicine, food, and cosmetics.

Conclusions

This review offers a comprehensive and current analysis of the functional activities and biological activity enhancement strategies for corosolic acid. It's demonstrated therapeutic potential spans anti-diabetic, anti-cancer, anti-inflammatory, and antibacterial properties, with notable activities including enzyme inhibition (e.g., α -amylase and α -glucosidase), regulation of insulin signaling pathways, induction of ferroptosis, promotion of apoptosis, and suppression of pro-inflammatory cytokines and biofilm formation. These attributes position corosolic acid as a promising candidate for applications in dietary supplements and pharmaceuticals. However, its poor water solubility and limited bioavailability remain significant barriers to broader utilization. Strategies such as chemical modifications, microbial transformations, and advanced delivery systems (e.g., liposomes and nanoemulsions) have shown promise in overcoming these challenges, enhancing therapeutic efficacy, and broadening functionality. Advanced delivery systems, in particular, demonstrate significant potential to simultaneously improve solubility, stability, and biological activities. These advancements underscore the importance of optimizing corosolic acid's pharmacological profile for practical applications in food and medicine.

Future research should prioritize refining these enhancement strategies, elucidating the molecular mechanisms underlying its diverse bioactivities, and ensuring the long-term safety and efficacy of corosolic acid. Addressing these challenges will pave the way for

corosolic acid to emerge as a highly valuable bioactive compound with broad applications in health and wellness.

Author contributions

The authors confirm contribution to the paper as follows: formal analysis, writing - original draft: Shi B; investigation: Shi B, Sun H; supervision: Jia Q, Luo Z; validation: Mao Y; conceptualization; funding acquisition: Luo Z; writing - review and editing: Sun Z, Zhang H, Luo Z. All authors reviewed the results and approved the final version of the manuscript.

Data availability

The datasets generated during and/or analyzed during the current study are available from the corresponding author on reasonable request.

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Conflict of interest

The authors declare that they have no conflict of interest.

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References

1. Samananda Singh L, Singh WS. 2024. Multifaceted therapeutic potential of corosolic acid: a novel bioactive compound. *Obesity Medicine* 49:100548
2. Jin M, Li J, Zheng L, Huang M, Wu Y, et al. 2024. Corosolic acid delivered by exosomes from *Eriobotrya japonica* decreased pancreatic cancer cell proliferation and invasion by inducing SAT1-mediated ferroptosis. *International Immunopharmacology* 132:111939
3. Phakeovilay C, Bourgeade-Delmas S, Perio P, Valentin A, Chassagne F, et al. 2019. Antileishmanial compounds isolated from *Psidium guajava* L. using a metabolomic approach. *Molecules* 24(24):4536
4. Sharma Y, Velamuri R, Fagan J, Schaefer J. 2020. Full-spectrum analysis of bioactive compounds in rosemary (*Rosmarinus officinalis* L.) as influenced by different extraction methods. *Molecules* 25(20):4599
5. Ladurner A, Zehl M, Grienke U, Hofstadler C, Faur N, et al. 2017. Allspice and clove as source of triterpene acids activating the g Protein-Coupled bile acid receptor TGR5. *Frontiers in Pharmacology* 8:468
6. Xue L, Carreiro B, Mia MS, Paetau-Robinson I, Khoo C, et al. 2024. Pentacyclic triterpenoid content in cranberry raw materials and products. *Foods* 13(19):3136
7. Hu F, Liao X, Guo Y, Yamaki S, Li X, et al. 2020. Fast determination of isomeric triterpenic acids in *Osmanthus fragrans* (Thunb.) Lour. fruits by UHPLC coupled with triple quadrupole mass spectrometry. *Food Chemistry* 322:126781
8. Olszewska M. 2008. Optimization and validation of an HPLC-UV method for analysis of corosolic, oleanolic, and ursolic acids in plant material: Application to *Prunus serotina* Ehrh. *Acta Chromatographica* 20(4):643–59
9. Qian XP, Zhang XH, Sun LN, Xing WF, Wang Y, et al. 2021. Corosolic acid and its structural analogs: a systematic review of their biological activities and underlying mechanism of action. *Phytomedicine* 91:153696

10. Mioc M, Milan A, Malița D, Mioc A, Prodea A, et al. 2022. Recent advances regarding the molecular mechanisms of triterpenic acids: a review (Part I). *International Journal of Molecular Sciences* 23(14):7740
11. Mioc M, Prodea A, Racoviceanu R, Mioc A, Ghiulai R, et al. 2022. Recent advances regarding the molecular mechanisms of triterpenic acids: a review (Part II). *International Journal of Molecular Sciences* 23(16):8896
12. Ni M, Pan J, Hu X, Gong D, Zhang G. 2019. Inhibitory effect of corosolic acid on α -glucosidase: kinetics, interaction mechanism, and molecular simulation. *Journal of the Science of Food and Agriculture* 99(13):5881–89
13. Zhang BW, Xing Y, Wen C, Yu XX, Sun WL, et al. 2017. Pentacyclic triterpenes as α -glucosidase and α -amylase inhibitors: structure-activity relationships and the synergism with acarbose. *Bioorganic & Medicinal Chemistry Letters* 27(22):5065–70
14. Shi L, Zhang W, Zhou YY, Zhang YN, Li JY, et al. 2008. Corosolic acid stimulates glucose uptake via enhancing insulin receptor phosphorylation. *European Journal of Pharmacology* 584(1):21–29
15. Yang J, Leng J, Li JJ, Tang JF, Li Y, et al. 2016. Corosolic acid inhibits adipose tissue inflammation and ameliorates insulin resistance via AMPK activation in high-fat fed mice. *Phytomedicine* 23(2):181–90
16. Zhang C, Gao L, Zhang Y, Jin X, Wang M, et al. 2024. Corosolic acid inhibits EMT in lung cancer cells by promoting YAP-mediated ferroptosis. *Phytomedicine* 135:156110
17. Peng Y, Li N, Tang F, Qian C, Jia T, et al. 2022. Corosolic acid sensitizes ferroptosis by upregulating HERPUD1 in liver cancer cells. *Cell Death Discovery* 8(1):376
18. Lee MS, Cha EY, Thuong PT, Kim JY, Ahn MS, et al. 2010. Down-Regulation of human epidermal growth factor receptor 2/neu oncogene by corosolic acid induces cell cycle arrest and apoptosis in NCI-N87 human gastric cancer cells. *Biological & Pharmaceutical Bulletin* 33(6):931–37
19. Tang F, Peng Y, Liu J, Gao W, Xu Y. 2023. Integrating network pharmacology and experimental models to examine the mechanisms of corosolic acid in preventing hepatocellular carcinoma progression through activation PERK-eIF2 α -ATF4 signaling. *Naunyn-Schmiedeberg's Archives of Pharmacology* 396(12):3671–82
20. Wu TK, Hung TW, Chen YS, Pan YR, Hsieh YH, et al. 2024. Corosolic acid inhibits metastatic response of human renal cell carcinoma cells by modulating ERK/MMP2 signaling. *Environmental Toxicology* 39(2):857–68
21. Pundalik S, Hanumappa KR, Giresha AS, Urs D, Rajashekarappa S, et al. 2022. Corosolic acid inhibits secretory phospholipase A₂IIa as an Anti-Inflammatory function and exhibits anti-tumor activity in ehrlich ascites carcinoma bearing mice. *Journal of Inflammation Research* 15:6905–21
22. Aguirre MC, Delporte C, Backhouse N, Erazo S, Letelier ME, et al. 2006. Topical anti-inflammatory activity of 2 α -hydroxy pentacyclic triterpene acids from the leaves of *Ugni molinae*. *Bioorganic & Medicinal Chemistry* 14(16):5673–77
23. Garo E, Eldridge GR, Goering MG, DeLancey Pulcini E, Hamilton MA, et al. 2007. Asiatic acid and corosolic acid enhance the susceptibility of *Pseudomonas aeruginosa* biofilms to tobramycin. *Antimicrobial Agents and Chemotherapy* 51(5):1813–17
24. Zhou Y, Lv X, Chen M, Guo Y, Ding R, et al. 2020. Characterization of corosolic acid as a KPC-2 inhibitor that increases the susceptibility of KPC-2-positive bacteria to carbapenems. *Frontiers in Pharmacology* 11:1047
25. Romsuk J, Srisawat P, Robertlee J, Yasumoto S, Miura K, et al. 2024. Heterologous production of corosolic acid, a phyto-insulin, in agroinfiltrated *Nicotiana benthamiana* leaves. *Plant Biotechnology* 41(3):277–88
26. Furtado NJC, Pirson L, Edelberg H, Miranda LM, Loira-Pastoriza C, et al. 2017. Pentacyclic triterpene bioavailability: an overview of in vitro and in vivo studies. *Molecules* 22(3):400
27. Spivak AY, Kuzmina US, Nedopekina DA, Dubinin MV, Khalitova RR, et al. 2024. Synthesis and comparative analysis of the cytotoxicity and mitochondrial effects of triphenylphosphonium and F16 maslinic and corosolic acid hybrid derivatives. *Steroids* 209:109471
28. Feng X, Lu YH, Liu Z, Li DP, Zou YX, et al. 2017. Microbial transformation of the anti-diabetic agent corosolic acid by *Cunninghamella echinulata*. *Journal of Asian Natural Products Research* 19(7):645–50
29. Li X, Widjaya AS, Liu J, Liu X, Long Z, et al. 2020. Cell-penetrating corosolic acid liposome as a functional carrier for delivering chemotherapeutic drugs. *Acta Biomaterialia* 106:301–13
30. Li H, Tan X, Qin L, Gatasheh MK, Zhang L, et al. 2024. Preparation, process optimisation, stability and bacteriostatic assessment of composite nanoemulsion containing corosolic acid. *Heliyon* 10(19):e38283
31. Zhao J, Zhou H, An Y, Shen K, Yu L. 2021. Biological effects of corosolic acid as an anti-inflammatory, anti-metabolic syndrome and anti-neoplastic natural compound. *Oncology Letters* 21(2):84
32. Sivakumar G, Vail DR, Nair V, Medina Bolivar F, Lay JO. 2009. Plant-based corosolic acid: Future anti-diabetic drug? *Biotechnology Journal* 4(12):1704–11
33. Rangel-Galván M, Pacheco-Hernández Y, Lozoya-Gloria E, Villa-Ruano N. 2024. Dietary natural products as inhibitors of α -amylase and α -glucosidase: an updated review of ligand-receptor correlations validated by docking studies. *Food Bioscience* 62:105456
34. Khalid H, Butt MH, Ur Rehman Aziz A, Ahmad I, Iqbal F, et al. 2024. Phytobioinformatics screening of ayurvedic plants for potential α -glucosidase inhibitors in diabetes management. *Current Plant Biology* 40:100404
35. Aydin S, Tekinalp SG, Tuzcu B, Cam F, Sevik MO, et al. 2025. The role of AMP-activated protein kinase activators on energy balance and cellular metabolism in type 2 diabetes mellitus. *Obesity Medicine* 53:100577
36. Kwon EB, Kang MJ, Ryu HW, Lee S, Lee JW, et al. 2020. Acacetin enhances glucose uptake through insulin-independent GLUT4 translocation in L6 myotubes. *Phytomedicine* 68:153178
37. Xu S, Yu W, Zhang X, Wang W, Wang X. 2022. The regulatory role of Gnao1 protein in diabetic encephalopathy in KK-Ay mice and streptozotocin-induced diabetic rats. *Brain Research* 1792:148012
38. Miura T, Ueda N, Yamada K, Fukushima M, Ishida T, et al. 2006. Antidiabetic effects of corosolic acid in KK-Ay diabetic mice. *Biological & Pharmaceutical Bulletin* 29(3):585–87
39. Hibi M, Matsui Y, Niwa S, Oishi S, Yanagimoto A, et al. 2022. Corosolic acid improves glucose and insulin responses in middle-aged men with impaired fasting glucose: A randomized, double-blinded, placebo-controlled crossover trial. *Journal of Functional Foods* 97:105256
40. Zhang W, Men X, Lei P. 2014. Review on anti-tumor effect of triterpene acid compounds. *Journal of Cancer Research and Therapeutics* 10:14–19
41. Bahadori MB, Vandghanooni S, Dinparast L, Eskandani M, Ayatollahi SA, et al. 2019. Triterpenoid corosolic acid attenuates HIF-1 stabilization upon cobalt (II) chloride-induced hypoxia in A549 human lung epithelial cancer cells. *Fitoterapia* 134:493–500
42. Woo SM, Seo SU, Min KJ, Im SS, Nam JO, et al. 2018. Corosolic acid induces Non-Apoptotic cell death through generation of lipid reactive oxygen species production in human renal carcinoma cells. *International Journal of Molecular Sciences* 19(5):1309
43. Tu S, Zou Y, Yang M, Zhou X, Zheng X, et al. 2025. Ferroptosis in hepatocellular carcinoma: mechanisms and therapeutic implications. *Biomedicine & Pharmacotherapy* 182:117769
44. Li B, Li Y, Wang Q, Li F, Li F. 2019. Corosolic acid impairs human lung adenocarcinoma A549 cells proliferation by inhibiting cell migration. *Oncology Letters* 17(6):5747–53
45. Ku CY, Wang YR, Lin HY, Lu SC, Lin JY. 2015. Corosolic acid inhibits hepatocellular carcinoma cell migration by targeting the VEGFR2/Src/FAK pathway. *PLoS One* 10(5):e126725
46. Zhang C, Niu Y, Wang Z, Xu X, Li Y, et al. 2021. Corosolic acid inhibits cancer progression by decreasing the level of CDK19-mediated O-GlcNAcylation in liver cancer cells. *Cell Death & Disease* 12(10):811–89
47. Park JB, Lee JS, Lee MS, Cha EY, Kim S, et al. 2018. Corosolic acid reduces 5-FU chemoresistance in human gastric cancer cells by activating AMPK. *Molecular Medicine Reports* 18(3):2880–88
48. Chen J, Kang J, Yuan S, O Connell P, Zhang Z, et al. 2024. Exploring the mechanisms of traditional chinese herbal therapy in gastric cancer: A comprehensive network pharmacology study of the Tiao-Yuan-Tong-Wei decoction. *Pharmaceuticals* 17(4):414
49. Kim JH, Kim YH, Song GY, Kim DE, Jeong YJ, et al. 2014. Ursolic acid and its natural derivative corosolic acid suppress the proliferation of APC-mutated colon cancer cells through promotion of β -catenin degradation. *Food and Chemical Toxicology* 67:87–95

50. Han S, Lim SL, Kim H, Choi H, Lee MY, et al. 2024. Ethyl acetate fraction of *Osmanthus fragrans* var. *Aurantiacus* and its triterpenoids suppress proliferation and survival of colorectal cancer cells by inhibiting NF- κ B and COX2. *Journal of Ethnopharmacology* 319:117362
51. Yoo KH, Park JH, Lee DY, Hwang-Bo J, Baek NI, et al. 2015. Corosolic acid exhibits anti-angiogenic and anti-lymphangiogenic effects on *in vitro* endothelial cells and on an *in vivo* CT-26 colon carcinoma animal model. *Phytotherapy Research* 29(5):714–23
52. Luo X, Ye Z, Xu C, Chen H, Dai S, et al. 2024. Corosolic acid enhances oxidative stress-induced apoptosis and senescence in pancreatic cancer cells by inhibiting the JAK2/STAT3 pathway. *Molecular Biology Reports* 51(1):176
53. Jasim SA, Khalaf OZ, Alshahrani SH, Hachem K, Ziyadullaev S, et al. 2023. An *in vitro* investigation of the apoptosis-inducing activity of corosolic acid in breast cancer cells. *Iranian Journal of Basic Medical Sciences* 26(4):453–60
54. Cui A, Li X, Ma X, Song Z, Wang X, et al. 2023. Quantitative transcriptomic and proteomic analysis reveals corosolic acid inhibiting bladder cancer via suppressing cell cycle and inducing mitophagy *in vitro* and *in vivo*. *Toxicology and Applied Pharmacology* 480:116749
55. Fujiwara Y, Takaishi K, Nakao J, Ikeda T, Katabuchi H, et al. 2013. Corosolic acid enhances the antitumor effects of chemotherapy on epithelial ovarian cancer by inhibiting signal transducer and activator of transcription 3 signaling. *Oncology Letters* 6(6):1619–23
56. Chen JL, Lai CY, Ying TH, Lin CW, Wang PH, et al. 2021. Modulating the ERK1/2–MMP1 axis through corosolic acid inhibits metastasis of human oral squamous cell carcinoma cells. *International Journal of Molecular Sciences* 22(16):8641
57. Cai X, Zhang H, Tong D, Tan Z, Han D, et al. 2011. Corosolic acid triggers mitochondria and caspase-dependent apoptotic cell death in osteosarcoma MG-63 cells. *Phytotherapy Research* 25(9):1354–61
58. Tang FF, Liu L, Tian XT, Li N, Peng YX, et al. 2023. Network pharmacological analysis of corosolic acid reveals P4HA2 inhibits hepatocellular carcinoma progression. *BMC Complementary Medicine and Therapies* 23:171
59. Lee MS, Lee CM, Cha EY, Thuong PT, Bae K, et al. 2010. Activation of AMP-activated protein kinase on human gastric cancer cells by apoptosis induced by corosolic acid isolated from *Weigela subsessilis*. *Phytotherapy Research* 24(12):1857–61
60. Lee HS, Park JB, Lee MS, Cha EY, Kim JY, et al. 2015. Corosolic acid enhances 5-fluorouracil-induced apoptosis against SNU-620 human gastric carcinoma cells by inhibition of mammalian target of rapamycin. *Molecular Medicine Reports* 12(3):4782–88
61. Chan EWC, Soon CY, Tan JBL, Wong SK, Hui YW. 2019. Ursolic acid: an overview on its cytotoxic activities against breast and colorectal cancer cells. *Journal of Integrative Medicine* 17(3):155–60
62. Hudlikar RR, Sargsyan D, Wu R, Su S, Zheng M, et al. 2020. Triterpenoid corosolic acid modulates global CpG methylation and transcriptome of tumor promoter TPA induced mouse epidermal JB6 P+ cells. *Chemico-Biological Interactions* 321:109025
63. Ooi KX, Poo CL, Subramaniam M, Cordell GA, Lim YM. 2023. Maslinic acid exerts anticancer effects by targeting cancer hallmarks. *Phytomedicine* 110:154631
64. Xu YQ, Zhang JH, Yang XS. 2016. Corosolic acid induces potent anticancer effects in CaSki cervical cancer cells through the induction of apoptosis, cell cycle arrest and PI3K/Akt signalling pathway. *Bangladesh Journal of Pharmacology* 11(2):453
65. Li XQ, Tian W, Liu XX, Zhang K, Huo JC, et al. 2016. Corosolic acid inhibits the proliferation of glomerular mesangial cells and protects against diabetic renal damage. *Scientific Reports* 6:26854
66. Zhang J, Zhao Y, Yan L, Tan M, Jin Y, et al. 2024. Corosolic acid attenuates cardiac ischemia/reperfusion injury through the PHB2/PINK1/parkin/mitophagy pathway. *iScience* 27(8):110448
67. Alkholifi FK, Devi S, Yusufoglu HS, Alam A. 2023. The cardioprotective effect of corosolic acid in the diabetic rats: A possible mechanism of the PPAR- γ pathway. *Molecules* 28(3):929
68. Wang ZP, Che Y, Zhou H, Meng YY, Wu HM, et al. 2020. Corosolic acid attenuates cardiac fibrosis following myocardial infarction in mice. *International Journal of Molecular Medicine* 45(5):1425–35
69. Che Y, Wang Z, Yuan Y, Zhou H, Wu H, et al. 2022. By restoring autophagic flux and improving mitochondrial function, corosolic acid protects against Dox-induced cardiotoxicity. *Cell Biology and Toxicology* 38(3):451–67
70. Luna-Vázquez FJ, Ibarra-Alvarado C, Del Rayo Camacho-Corona M, Rojas-Molina A, Rojas-Molina JI, et al. 2018. Vasodilator activity of compounds isolated from plants used in Mexican traditional medicine. *Molecules* 23(6):1474
71. Yamamura A, Fujiwara M, Kawade A, Amano T, Hossain A, et al. 2024. Corosolic acid attenuates platelet-derived growth factor signaling in macrophages and smooth muscle cells of pulmonary arterial hypertension. *European Journal of Pharmacology* 973:176564
72. Sun Q, He M, Zhang M, Zeng S, Chen L, et al. 2020. Ursolic acid: A systematic review of its pharmacology, toxicity and rethink on its pharmacokinetics based on PK-PD model. *Fitoterapia* 147:104735
73. Han H, Chen M, Li Z, Zhou S, Wu Y, et al. 2022. Corosolic acid protects rat chondrocytes against IL-1 β -induced ECM degradation by activating autophagy via PI3K/AKT/mTOR pathway and ameliorates rat osteoarthritis. *Drug Design, Development and Therapy* 16:2627–37
74. Peng M, Qiang L, Xu Y, Li C, Li T, et al. 2019. Inhibition of JNK and activation of the AMPK-Nrf2 axis by corosolic acid suppress osteolysis and oxidative stress. *Nitric Oxide* 82:12–24
75. Hong G, Zhou L, Han X, Sun P, Chen Z, et al. 2020. Asiatic acid inhibits OVX-induced osteoporosis and osteoclastogenesis via regulating RANKL-mediated NF- κ B and Nfatc1 signaling pathways. *Frontiers in Pharmacology* 11:331
76. Shen H, Wang J, Ao J, Ye L, Shi Y, et al. 2023. The inhibitory mechanism of pentacyclic triterpenoid acids on pancreatic lipase and cholesterol esterase. *Food Bioscience* 51:102341
77. Yamaguchi Y, Yamada K, Yoshikawa N, Nakamura K, Haginaka J, et al. 2006. Corosolic acid prevents oxidative stress, inflammation and hypertension in SHR/NDmcr-cp rats, a model of metabolic syndrome. *Life Sciences* 79(26):2474–79
78. Zong W, Zhao G. 2007. Corosolic acid isolation from the leaves of *Eriobotrya japonica* showing the effects on carbohydrate metabolism and differentiation of 3T3-L1 adipocytes. *Asia Pacific Journal of Clinical Nutrition* 16(Suppl1):346–52
79. Guo X, Cui R, Zhao J, Mo R, Peng L, et al. 2016. Corosolic acid protects hepatocytes against ethanol-induced damage by modulating mitogen-activated protein kinases and activating autophagy. *European Journal of Pharmacology* 791:578–88
80. Fontanay S, Grare M, Mayer J, Finance C, Duval RE. 2008. Ursolic, oleanolic and betulinic acids: antibacterial spectra and selectivity indexes. *Journal of Ethnopharmacology* 120(2):272–76
81. Sycz Z, Tichaczek-Goska D, Wojnicz D. 2022. Anti-planktonic and anti-biofilm properties of pentacyclic triterpenes—asiatic acid and ursolic acid as promising antibacterial future pharmaceuticals. *Biomolecules* 12(1):98
82. Yum SJ, Kim SM, Yu YC, Jeong HG. 2017. Inhibition of growth and biofilm formation of *Staphylococcus aureus* by corosolic acid. *Korean Society of Food Science and Technology* 49(2):146–50
83. Sinelius S, Lady J, Yunardy M, Tjoa E, Nurcahyanti ADR. 2023. Antibacterial activity of *Lagerstroemia speciosa* and its active compound, corosolic acid, enhances cefotaxime inhibitory activity against *Staphylococcus aureus*. *Journal of Applied Microbiology* 134(8):d171
84. Kurek A, Grudniak AM, Szwed M, Klicka A, Samluk L, et al. 2010. Oleanolic acid and ursolic acid affect peptidoglycan metabolism in *Listeria monocytogenes*. *Antonie van Leeuwenhoek* 97(1):61–68
85. Kim J, Park S, Shin YK, Kang H, Kim KY. 2018. *In vitro* antibacterial activity of macelignan and corosolic acid against the bacterial bee pathogens *Paenibacillus larvae* and *Melissococcus plutonius*. *Acta Veterinaria Brno* 87(3):277–84
86. Shin YK, Kim KY. 2016. Macelignan inhibits bee pathogenic fungi *Ascophaera apis* growth through HOG1 pathway. *Brazilian Journal of Medical and Biological Research [Revista Brasileira de Pesquisas Medicas e Biologicas]* 49(7):e5313
87. Zhao WB, Zhao ZM, Ma Y, Li AP, Zhang ZJ, et al. 2022. Antifungal activity and preliminary mechanism of pristimerin against *Sclerotinia sclerotiorum*. *Industrial Crops and Products* 185:115124

88. Zhao HY, Zhu HY, Tang Q, Lin Q, Hao YK, et al. 2024. Anti-inflammatory and antiviral activities of compounds from the fruit of *Pouteria caimito*. *Cogent Food & Agriculture* 10:1
89. Zhou JF, Zhang MR, Wang Q, Li MZ, Bai JS, et al. 2024. Two novel compounds inhibit Flavivirus infection *in vitro* and *in vivo* by targeting lipid metabolism. *Journal of Virology* 98(9):e0063524
90. Yang YH, Dai SY, Deng FH, Peng LH, Li C, et al. 2022. Recent advances in medicinal chemistry of oleanolic acid derivatives. *Phytochemistry* 203:113397
91. Chen L, Gong J, Yong X, Li Y, Wang S. 2024. A review of typical biological activities of glycyrrhetic acid and its derivatives. *RSC Advances* 14(1):6557–97
92. Farina C, Pinza M, Pifferi G. 1998. Synthesis and anti-ulcer activity of new derivatives of glycyrrhetic, oleanolic and ursolic acids. *Il Farmaco* 53(1):22–32
93. Yan R, Liu L, Huang X, Quan ZS, Shen QK, et al. 2024. Bioactivities and structure-activity relationships of maslinic acid derivatives: a review. *Chemistry & Biodiversity* 21(2):e202301327
94. Hussain H, Ali I, Wang D, Hakkim FL, Westermann B, et al. 2021. Glycyrrhetic acid: a promising scaffold for the discovery of anti-cancer agents. *Expert Opinion on Drug Discovery* 16(12):1497–516
95. Lv J, Sharma A, Zhang T, Wu Y, Ding X. 2018. Pharmacological review on asiatic acid and its derivatives: A potential compound. *SLAS Technology* 23(2):111–27
96. Zhao ZX, Zou QY, Ma YH, Morris-Natschke SL, Li XY, et al. 2025. Recent progress on triterpenoid derivatives and their anticancer potential. *Phytochemistry* 229:114257
97. Yang H, Deng M, Jia H, Zhang K, Liu Y, et al. 2024. A review of structural modification and biological activities of oleanolic acid. *Chinese Journal of Natural Medicines* 22(1):15–30
98. Huang J, Zang X, Yang W, Yin X, Huang J, et al. 2021. Pentacyclic triterpene carboxylic acids derivatives integrated piperazine-amino acid complexes for α -glucosidase inhibition *in vitro*. *Bioorganic Chemistry* 115:105212
99. Liu X, Zang X, Yin X, Yang W, Huang J, et al. 2020. Semi-synthesis of C28-modified triterpene acid derivatives from maslinic acid or corosolic acid as potential α -glucosidase inhibitors. *Bioorganic Chemistry* 97:103694
100. Heise NV, Csuk R, Mueller T. 2024. (Iso)quinoline amides derived from corosolic acid exhibit high cytotoxicity, and the potential for overcoming drug resistance in human cancer cells. *European Journal of Medicinal Chemistry Reports* 12:100198
101. Xu J, Nie X, Hong Y, Jiang Y, Wu G, et al. 2016. Synthesis of water soluble glycosides of pentacyclic dihydroxytriterpene carboxylic acids as inhibitors of α -glucosidase. *Carbohydrate Research* 424:42–53
102. Kumari H, Ganjoo A, Shafeeq H, Ayoub N, Babu V, et al. 2024. Microbial transformation of some phytochemicals into value-added products: A review. *Fitoterapia* 178:106149
103. Abdel Bar FM, Elekhawy E, Salkini AA, Soliman AF. 2024. Enhancement of antibacterial and antibiofilm properties of proximadiol through microbial transformation by *Rhizopus oryzae*. *South African Journal of Botany* 172:236–41
104. Li DP, Feng X, Chu ZY, Guo FF, Zhang ZS. 2013. Microbial transformation of corosolic acid by *Fusarium equiseti* and *Gliocladium catenulatum*. *Journal of Asian Natural Products Research* 15(7):789–808
105. Xu SH, Zhang C, Wang WW, Yu BY, Zhang J. 2017. Site-selective biotransformation of ursane triterpenes by *Streptomyces griseus* ATCC 13273. *RSC Advances* 7(34):20754–59
106. Kumar P, Bhadauria AS, Singh AK, Saha S. 2018. Betulinic acid as apoptosis activator: Molecular mechanisms, mathematical modeling and chemical modifications. *Life Sciences* 209:24–33
107. Cui N, Li MJ, Wang YW, Meng Q, Shi YJ, et al. 2024. Boswellic acids: a review on its pharmacological properties, molecular mechanism and bioavailability. *Traditional Medicine Research* 9(10):60
108. Fernandes S, Vieira M, Prudêncio C, Ferraz R. 2024. Betulinic acid for glioblastoma treatment: Reality, challenges and perspectives. *International Journal of Molecular Sciences* 25(4):2108
109. Rehman Sheikh A, Wu-Chen RA, Matloob A, Mahmood MH, Javed M. 2024. Nanoencapsulation of volatile plant essential oils: a paradigm shift in food industry practices. *Food Innovation and Advances* 3(3):305–19
110. Malik M, Velechovský J, Tlustoš P. 2021. Natural pentacyclic triterpenoid acids potentially useful as biocompatible nanocarriers. *Fitoterapia* 151:104845
111. Bildziukevich U, Özdemir Z, Wimmer Z. 2019. Recent achievements in medicinal and supramolecular chemistry of betulinic acid and its derivatives. *Molecules* 24(19):3546
112. Bag BG, Garai S, Ghorai S. 2019. Vesicular self-assembly of a natural ursane-type dihydroxy-triterpenoid corosolic acid. *RSC Advances* 9(27):15190–95
113. Bao H, Sun W, Sun H, Jin Y, Gong X, et al. 2022. Liquid chromatographic study of two structural isomeric pentacyclic triterpenes on reversed-phase stationary phase with hydroxypropyl- β -cyclodextrin as mobile phase additive. *Journal of Pharmaceutical and Biomedical Analysis* 207:114420
114. Liu S, Liu Y, Li Q, Song Y, Zhang L, et al. 2024. Oleanolic acid nanoparticles-stabilized W/O pickering emulsions: fabrication, characterization, and delivery application. *Food Chemistry* 444:138598
115. Liu Y, Xia H, Guo S, Lu X, Zeng C. 2022. Development and characterization of a novel naturally occurring pentacyclic triterpene self-stabilized pickering emulsion. *Colloids and Surfaces A: Physicochemical and Engineering Aspects* 634:127908
116. Liu Y, Zhang R, Yang Y, Liu X, Jiang Y. 2025. Corosolic acid derivative-based lipid nanoparticles for efficient RNA delivery. *Journal of Controlled Release* 378:1–17
117. Widjaya AS, Liu Y, Yang Y, Yin W, Liang J, et al. 2022. Tumor-permeable smart liposomes by modulating the tumor microenvironment to improve the chemotherapy. *Journal of Controlled Release* 344:62–79



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