

Hydrogen sulfide: a luminous future in the postharvest preservation of fruits and vegetables

Ya-Qin Zhao, Liang-Yi Zhao, Shun-Qing Hu, Yuan-Yuan Hou, Yong-Hua Zheng, and Peng Jin*

College of Food Science and Technology, Nanjing Agricultural University, 1 Weigang, Nanjing 210095, China

* Corresponding author, E-mail: pjin@njau.edu.cn

Abstract

Hydrogen sulfide (H₂S) has emerged as a signaling molecule that plays a crucial role in the postharvest preservation of fruits and vegetables. This review summarizes the various functions of H₂S such as delaying ripening and senescence, enhancing the resistance to cold and disease, and emphasizes the underlying mechanisms. Appropriate concentrations of H₂S primarily operate through stimulating the antioxidant system, while showing positive effects on physiological metabolism relevant to storage quality and shelf life including energy, sugar, proline, phenolic, membrane lipid and cell wall metabolism. Moreover, H₂S may reduce storage loss by modulating the expression patterns of senescence-related genes, like those linked to ethylene. The coordination of H₂S and nitric oxide (NO) combats ethylene-derived negative effects during ripening and senescence. High concentrations of H₂S not only act as a regulator to induce disease resistance, but also as a fungicide to inhibit the growth and pathogenicity of fungi. The intricate crosstalk between H₂S and other molecules exists via synergistic and antagonistic roles based on protein persulfidation, which is the major signaling process of H₂S and appears to compete with other post-translational modifications (PTMs) for the same cysteine residues. This review summarizes H₂S synthesis pathways and also discusses the correlation between the signals of H₂S, Ca²⁺ and ABA, while highlighting the intrinsic mechanisms of H₂S performing its functions in postharvest preservation.

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Introduction

H₂S, a toxic gas with a rotten egg odor, has recently warranted inclusion as a messenger molecule. H₂S diffuses readily through membranes as small lipophilic molecules, presenting a membrane receptor-independent signaling mechanism^[1]. H₂S signaling in mammalian systems occurs through reactions with metals and oxidants as well as protein persulfidation, which either activates or inhibits enzyme activities directly, or by forming protein adducts^[2]. Moreover, H₂S in plants induces defense systems against biotic and abiotic stress from seed germination to fruit ripening^[3], protein persulfidation in postharvest has also obtained keen interest. After fruits and vegetables are picked, they lose their nutritional supply and gradually senescence during the storage process, so preservation has always been a relevant issue. Recent reports on H₂S application show great potential for delaying senescence in cut flowers, vegetables, climacteric and non-climacteric fruit under shelf and low temperature conditions, including enhancing the antioxidant system, mitigating chilling injury (CI) symptoms, inhibiting fungal growth, regulating senescence-related genes and interacting with other molecules^[4–6]. Nevertheless, the mechanisms of H₂S on postharvest physiological metabolism require further research. The residual levels of H₂S after exogenous treatment are lower than human plasma levels^[6], and there is usually no disagreeable smell in the preparation of plant extracts^[3]. Consequently, a stimulate concentration of H₂S may have

safe applications in postharvest storage and transportation. Intricate interactions between H₂S and other secondary messengers like Ca²⁺, NO, abscisic acid (ABA) and hydrogen peroxide (H₂O₂) form a cross-adaptation signaling network^[7]. The function of H₂S to activate or inhibit target proteins via persulfidation deserves more attention, while possibly contending with other PTMs for the same residue positions, especially NO-mediated S-nitrosation and nitration^[3,8]. Although there have been many reports, the effects of H₂S on postharvest physiology of fruits and vegetables still need comprehensive evaluation. This review focuses on the intrinsic mechanisms of H₂S signaling on senescence, CI and disease resistance, and also highlights the current progress of persulfidation with other modifications.

Biosynthesis of H₂S in plants

Transsulfuration is the major pathway for plants to produce endogenous H₂S. Wilson et al.^[9] first discovered that the emission rate of H₂S in higher plants was light-dependent. It seemed that H₂S was a crucial part of sulfur metabolism and helped plants to remove excessive inorganic sulfur anions^[9]. Since then, researchers have discovered that there are at least five pathways to generate endogenous H₂S in plants (Fig. 1). The five known pathways are as follows: (1) Plants convert the combined sulfate to the free state that is activated to form adenosine 50-phosphosulfate (APS) in the plastids. Then APS is reduced to H₂S by APS reductase (APR) and sulfite

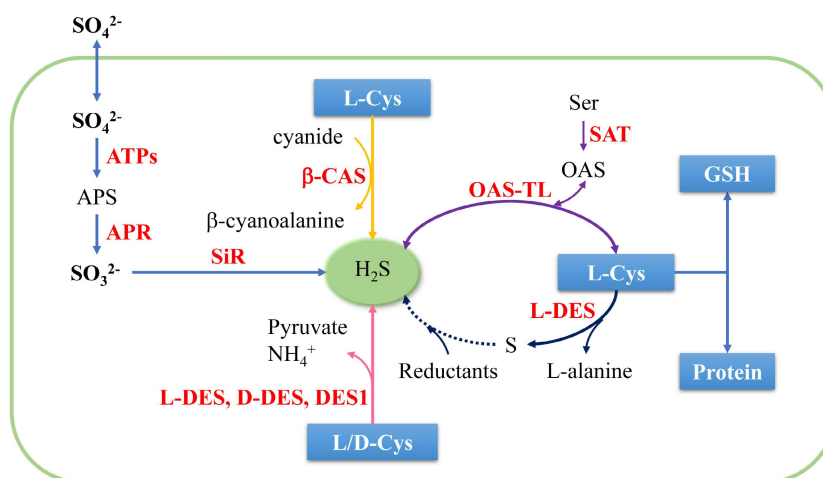


Fig. 1 Endogenous H₂S biosynthesis in higher plants. SO₄²⁻: sulfate ion; SO₃²⁻: sulfite ion; ATPs: ATP sulfurylase; APS: adenosine 50-phosphosulfate; APR: APS reductase; SiR: sulfite reductase; β -CAS: β -cyanoalanine synthase; L-Cys: L-cysteine; D-Cys: D-cysteine; L-DES: L-cysteine desulfhydrase; D-DES: D-cysteine desulfhydrase; DES1: L-cysteine desulfhydrase 1; Ser: serine; SAT: serine acetyltransferase; OAS: Oacetylserine; OAS-TL: O-acetylserine thiol lyase; GSH: reduced glutathione.

reductase (SiR), with reduced glutathione (GSH) as the electron donor^[10]. (2) Cysteine desulfhydrase is the main pathway to generate endogenous H₂S, including L-cysteine desulfhydrase (L-DES), D-cysteine desulfhydrase (D-DES) and L-cysteine desulfhydrase 1 (DES1). L-DES and D-DES are not directly related, they have different substrates and subcellular localization^[5,11]. (3) O-acetylserine thiol lyase (OAS-TL), also known as cysteine synthase (CS), catalyzes sulfide into O-acetylserine (OAS) to form cysteine, its reversible reactions can release H₂S^[12]. (4) Pyridoxal 5'-phosphate-dependent L-DES catalyzes L-cysteine to generate elemental sulfur, which is further reduced to H₂S^[13]. Two candidate genes *AtNFS1* and *AtNFS2* have been identified in *Arabidopsis thaliana*, encoding NifS-like plastidial cysteine desulfhydrase, which are addressed to plastids and chloroplasts, respectively^[14]. (5) β -cyanoalanine synthase (β -CAS) converts L-cysteine into H₂S through the detoxification of cyanide^[15]. NaHS, a commonly used exogenous H₂S donor, gives a rapid burst of H₂S gas in a water solution and keeps a constant concentration, but actual physiological H₂S concentration triggering the defense response is yet unclear^[7].

Interaction of H₂S, Ca²⁺ and ABA signaling

The chemical properties of H₂S allow it to diffuse readily through membranes without the necessity of membrane channels or specific transporters, and confers its biological signaling via protein persulfidation^[2,3]. Protein cysteine residues undergo H₂S-derived persulfidation to form persulfides (R-SSH), which is the mechanism by which H₂S performs its biological function^[2]. In plant physiobiochemistry, complex crosstalk between H₂S and phytohormones such as salicylic acid (SA), gibberellins (GAs) and jasmonic acid (JA) have been reported. It also interacts with other messengers like NO, H₂O₂ and carbon monoxide (CO) to regulate plant growth and development^[16]. This review focuses on the interconnection of Ca²⁺, ABA and H₂S. Recent studies show that the relationship between Ca²⁺ signaling and H₂S is not a simple upstream-downstream relationship. Under stress, intra-

cellular Ca²⁺ signaling responded to environmental stimuli to promote endogenous H₂S accumulation, while H₂S mediated Ca²⁺ signaling transduction by up-regulating the transcription of calmodulin (*CaM*), calcineurin B-like (*CBL*) and calcium dependent protein kinase (*CDPKs*) genes. Furthermore, H₂S and Ca²⁺ triggered ascorbic acid-glutathione (AsA-GSH) cycle against heavy metals (HMs)^[17,18] and low temperature stress^[19]. Fang et al.^[20] confirmed how Ca²⁺ promoted endogenous H₂S production under chromium (Cr⁶⁺) stress in *Arabidopsis*. The interaction between Ca²⁺/CaM2 and bZIP transcription factor (TF) *TGA3* activated *LCD* expression by enhancing binding ability of *TGA3* to the motif 'TGACG' of *LCD* promoter, forming a *CaM2-TGA3-pLCD* complex, resulting in H₂S accumulation^[20]. Nevertheless, the correlative studies on Ca²⁺ and H₂S signaling still remain in plant growth and development.

H₂S seems to interplay with ABA in drought responses. Shen et al. found a negative feedback loop between reactive oxygen species (ROS) and ABA signaling. ABA-triggered overproduction of ROS which further produces negative feedback to regulate respiratory burst oxidase homolog protein D (RbohD) and DES1 activity, resulting in ABA de-sensitivity^[21]. Then ABA-triggered *DES1* function contributed to long hypocotyl 1 (*HY1*) retrograde signaling^[22]. Chen et al. also reported that ABA-induced H₂S persulfidated the TF abscisic acid insensitive 4 (*ABI4*) at the residue Cys250, triggering the transactivation of mitogen-activated protein kinase kinase 18 (*MAPKKK18*) involved in autophosphorylation or phosphorylation of the sucrose nonfermenting 1 (SNF1)-related protein kinase 2.6 (SnRK2.6)^[23], while *ABI4* transactivated *DES1* to form a *DES1-ABI4* loop^[24]. Moreover, *HY1* can negatively regulate ABA-induced stomatal closure by activating *ABI4* expression^[22]. Accordingly, there appears to be a redox-based regulatory loop between H₂S signaling and ABA-induced stomatal closure. Furthermore, H₂S mediated ABA signaling by persulfidating SnRK2.6 at Cys131 and Cys137 residues. The interaction between SnRK2.6 and downstream ABA response factor 2 (*ABF2*) was enhanced by persulfidation, hence promoting ABA-induced stomatal

closure^[25]. Ca²⁺ signaling might harmonizes persulfidation and phosphorylation of SnRK2.6, and probably establishes a connection with ABA via ROS signaling^[23]. However, more is pending on how this complex regulatory mechanism controls postharvest ripening and senescence.

Roles of H₂S in postharvest senescence

Antagonism with ethylene

It has been well-established that H₂S modulates ripening and senescence processes by mediating ethylene biosynthesis and signal transduction, and H₂S plays a positive role in both climacteric and non-climacteric fruit (Table 1). After fumigating three green leafy vegetables with exogenous H₂S, then using 0.1 μL L⁻¹ ethylene to ventilate, respiration rate and chlorophyll loss were decreased^[26]. Compared with ethylene treatment, H₂S reduced the accumulation of reactive oxygen species (ROS) by enhancing the antioxidant system, and obviously delayed the color change of tomato fruit^[27]. Consequently, H₂S antagonizes ethylene-induced fruit ripening, which was associated with the antioxidant system. At the transcriptional level, Li et al. found that exogenous H₂S attenuated the expression of ethylene synthesis-related genes, such as *AdSAM*, *AdACS1*, *AdACS2*, *AdACO2* and *AdACO3*^[28]. In addition, H₂S also affects the expression patterns of genes linked to ethylene synthesis and signaling, including ethylene receptor genes and ethylene response factors (*ERFs*). Apple slices treated with 0.4 mM NaHS showed lower expression of *MdACO1*, *MdERS1* and *MdETR1*^[29]. Similarly, H₂S (1.0 mM NaHS) was combined with ethylene (1.0 g L⁻¹ ethephon) to fumigate banana, the released H₂S significantly down-regulated ethylene biosynthesis genes expression (*MaACS1*, *MaACS2* and *MaACO1*) and up-regulated ethylene receptor genes expression (*MaETR*, *MaERS1* and *MaERS2*)^[30]. In this case, ethylene receptors may act as a

negative regulator to inactivate by ethylene binding^[31]. Lin et al. reported that H₂S treatment reduced the expression of ethylene receptor 2 (*ETR2*) and *ERF* genes (*ERF003*, *ERF5* and *ERF016*), but increased the expression of *ERF4* and *ERF113*^[32]. Generally, H₂S counteracts the quality deterioration caused by ethylene via regulating ethylene-related genes, which is essential to postpone postharvest senescence.

There appears to be a cooperative relationship between H₂S and NO against ethylene. The combination of H₂S and NO delayed ripening and decay of strawberry^[33] and peach fruit^[34]. Combinatorial treatment (20 μL L⁻¹ H₂S gas and 15 μL L⁻¹ of NO gas) further attenuated ACC synthase (ACS) and oxidase (ACO) activities, and significantly decreased ACC and MACC content^[34]. Munoz-Vargas et al. also found that H₂S levels were increased during sweet pepper fruit ripening, while NO levels were reduced^[35]. NO binding with ACO to form a ternary complex reduces ethylene generation during the fruit ripening process^[36]. However, it remains uncertain how the intricate crosstalk between H₂S and NO co-regulates ethylene-induced ripening and senescence.

Regulation of senescence-related genes

The genes associated with ethylene mentioned above are also senescence-related genes^[5]. Co-treatment (0.9 mM NaHS and 1.0 g L⁻¹ ethephon) decreased protease activity and sustained a high level of bioactive compounds which were essential for tomato postharvest quality by principal component analysis (PCA), including soluble protein, starch, titratable acids (TA) and ascorbic acid (ASA). Meanwhile, H₂S reduced the transcription of ripening-related TFs (*ERF003* and *DOF22*), and changed the expression pattern of genes encoding beta-amylase and UDP-glycosyltransferase including *BAM3*, *UFGT73* and *UFGT5*^[37]. Consistent results were obtained from studies on fresh-cut pears. Reducing sugar, soluble protein and total amino acids content of pear slices under H₂S fumigation were higher^[38]. After harvest, chloro-

Table 1. H₂S counteracts the effects of ethylene in postharvest fruits and vegetables.

Materials	Treatment	Effect	Reference
Vegetables			
Pak choy (Shanghai)	NaHS fumigation for 4 h (0, 50, 100 and 250 μL L ⁻¹ H ₂ S)	Decreased ethylene production, chlorophyll loss and respiration rate	[26]
Green curly kale (Sabellica)			
Sweet Italian basil (Ocimum basilicum)			
Non-climacteric fruit			
Strawberry (Fengxiang)	0.8 mM NaHS and 5 μM SNP immersion for 10 min	Inhibited respiration rate and maintained crust color	[33]
Climacteric fruit			
Peach (Dahong)	15 μL L ⁻¹ NO gas and 20 μL L ⁻¹ H ₂ S gas fumigation for 20 min	Reduced ACS and ACO activities and inhibited respiration rate	[34]
Kiwifruit (Qinmei)	1.0 mM NaHS fumigation for 8 d	Down-regulated the expression of ethylene synthesis-related genes like <i>AdSAM</i> , <i>AdACS1</i> , <i>AdACS2</i> , <i>AdACO2</i> and <i>AdACO3</i>	[28]
Kiwifruit (Jinkui)	20 μL L ⁻¹ H ₂ S gas fumigation for 0.5 h	Down-regulated the expression of <i>ethylene receptor 2 (ETR2)</i> , <i>ERF003</i> , <i>ERF5</i> and <i>ERF016</i> ; up-regulated the expression of <i>ERF4</i> and <i>ERF113</i>	[32]
Fresh-cut apple (Fuji)	0.4 mM NaHS fumigation for 5 d	Up-regulated <i>MdDHAH</i> expression and down-regulated the expression of <i>MdLOX2</i> , <i>MdPG1</i> , <i>MdPPO</i> , <i>MdACO1</i> , <i>MdERS1</i> and <i>MdETR1</i>	[29]
Banana (Brazil)	1.0 mM NaHS and 1.0 g L ⁻¹ ethephon solution fumigation for 6 d	Inhibited chlorophyll loss; up-regulated the expression of ethylene receptor genes <i>MaETR</i> , <i>MaERS1</i> and <i>MaERS2</i> ; down-regulated the expression of ethylene synthesis-related genes like <i>MaACS1</i> , <i>MaACS2</i> and <i>MaACO1</i>	[30]
Tomato (Micro Tom)	0.9 mM NaHS and 1.0 g L ⁻¹ ethephon solution fumigation for 24 h	Maintained better appearance and postharvest quality and down-regulated the expression of <i>ERF003</i> and <i>DOF22</i>	[37]

phyll and some proteins are gradually degraded under the regulation of senescence-related genes. Extrapolating from recent studies, H₂S affects the transformation of large molecules at the transcriptional level. During postharvest, Li et al. reported a decreased expression of cysteine protease genes *AdCP1* and *AdCP3* in H₂S-treated kiwifruit^[28]. Moreover, H₂S reduced the expression of cysteine protease genes *BoCP1* and *BoCP3* and aspartic protease gene *BoLSC807*, while genes involved in chlorophyll degradation were successively found to be down-regulated by H₂S in postharvest broccoli, such as *BoSGR*, *BoRCCR*, *BoCLH2*, *BoPaO*, *BoNYC*, *BoCLH1* and *BoPPH*^[39,40]. In water spinach, exogenous H₂S fumigation reduced the activities of chlorophyll degradation-related enzymes, including chlorophyllase (Chlase), Mg-dechelatease (MD) and Mg-dechelating substance (MDS), further sustaining chlorophyll levels and slowing the decomposition of chloroplasts^[41]. Studies on banana^[30], pak choy^[26] and broccoli^[42] were consistent with the aforementioned results, where H₂S alleviated senescence-induced yellowing by mitigating chlorophyll loss. Ning et al. found that the TF *lBERF71* formed a *lBERF71-lbMYB340-lbbHLH2* complex to co-target the *lBAN51* promoter, resulting in anthocyanin accumulation in purple-fleshed sweet potato^[43]. Understanding the molecular mechanism by which H₂S regulates pigment metabolism is conducive to comprehending ripening and senescence processes. In addition, Lipoxygenase (LOX) catalyzes the hydroperoxidation of polyunsaturated fatty acids, and according to studies, H₂S treatment can inhibit its activity^[28] and gene expression^[39,40]. In summary, H₂S regulates senescence-related genes to ameliorate ethylene-induced quality deterioration, as well as higher nutritional compounds.

Regulation of cell wall metabolism

Softening and textural changes are modified by several enzymes, and this process maybe different in various types of fruits^[44]. It is well known that cell wall modifying enzymes taking part in fruit ripening are controlled by ethylene. Zhu et al.^[34] found that H₂S treatment (20 $\mu\text{L L}^{-1}$ H₂S gas) attenuated ethylene production and avoided cell wall loosening, while markedly decreasing the activities of cell wall modifying enzymes, such as polygalacturonase (PG), pectin methylesterase (PME) and endo- β -1,4-glucanase (EGase). H₂S-NO co-treatment (15 $\mu\text{L L}^{-1}$ NO gas and 20 $\mu\text{L L}^{-1}$ H₂S gas) further reduced the increases of water-soluble polysaccharides (WSP) and CDTA-soluble polysaccharides (CSP) in peach fruit, as well as the breakdown of Na₂CO₃-soluble polysaccharides (NSP)^[34]. Similarly, in postharvest strawberry, synergistic interactions between H₂S and NO (0.8 mM NaHS and 5 μM SNP) reduced cell wall modifying enzyme activity, and further improved softening and shelf life^[33]. The genes linked to cell wall degradation (EGase, PME and β -galactosidase) were down-regulated under H₂S treatment (20 $\mu\text{L L}^{-1}$ H₂S gas), while the expression pattern of ethylene responsive genes were also changed in kiwifruit, indicating that H₂S modifies the integrity of the cell wall at the transcription level^[32]. Besides, H₂S participates in cell wall metabolism and provides tolerance for cold stress. Lower activities of PG, pectate lyase (PL) and β -galactosidase (β -gal) and lower yields of WSP and CSP under H₂S treatment (1.0 or 2.0 mM NaHS) contributed positively to improve surface pitting symptoms in cold-stored sweet cherry^[45]. Cumulative reports demonstrate that H₂S

inhibits the depolymerization and solubilization of cell wall polysaccharides. In addition, Forlani et al. proposed that ABA is also a key ripening-associated regulator, which can promote ethylene biosynthesis^[44]. Whether the synergistic or antagonistic effects of H₂S and other molecules in fruit softening and senescence regulate cell wall metabolism directly or indirectly needs to be explored.

Roles of H₂S in antioxidant system during postharvest

The ROS scavenging system is divided into non-enzymatic and enzymatic. The enzymatic antioxidant system is composed of antioxidant enzymes like glutathione reductase (GR), catalase (CAT), superoxide dismutase (SOD), ascorbate peroxidase (APX), dehydroascorbate reductase (DHAR) and glutathione S-transferase (GST), and the non-enzymatic antioxidant system consists of antioxidants, such as GSH, AsA, polyphenols, and carotenoids^[46]. Abiotic stresses can destroy the dynamic balance of reactive oxygen species and reactive nitrogen species (ROS/RNS), which may ultimately cause irreversible nitro-oxidative damage^[3]. Accumulating evidence verifies that H₂S counteracts ROS accumulation to reduce oxidative damage. Ni et al.^[47] reported that high concentrations of H₂S (NaHS as donor) had toxic effects on Kyoho grape, described as severe decay and threshing rate. The optimal concentration of H₂S (1.0 mM NaHS) increased antioxidant content like AsA, flavonoid and total phenolics, as well as higher APX and CAT activity, while inhibiting H₂O₂, superoxide anion (O₂⁻) and malondialdehyde (MDA) accumulation due to its higher antioxidant capacity^[47]. Furthermore, it was reported that H₂S treatment (0.7 mM NaHS) showed higher relative expression of genes encoding SOD, CAT and GST, which ameliorated surface browning of fresh-cut apple by stimulating non-enzymatic and enzymatic antioxidant systems^[48]. Similarly, Yao et al. found that additional H₂S (0.9 mM NaHS) enhanced the activities of POD, APX, CAT and SOD, and also up-regulated the relative expression of *SIAPX2*, *SICAT1*, *SIPOD12* and *SICuZnSOD*^[27]. In a nutshell, this is a possible mechanism for H₂S to delay postharvest senescence by sustaining the redox balance.

Like senescence, CI also induces ROS burst^[46]. Application of H₂S induced chilling tolerance by enhancing antioxidant capacity as reported in banana^[49], hawthorn^[50] and kiwifruit^[51] during postharvest (Table 1). It is indicated that H₂S alleviates CI-induced quality deterioration during postharvest, which is associated with the antioxidant system. Interestingly, water spinach treated with NaHS enhanced L-DES and D-DES activity that generated more endogenous H₂S. However, it was recovered by using DL-propargylglycine (PAG, L-DES inhibitor), while the hypotaurine (HT, H₂S-scavenger) group was unaffected. Hence, Hu et al. deduced that endogenous H₂S was involved in the regulation of senescence^[41]. During cold storage, NaHS application on hawthorn fruit^[50] and mulberry fruit^[52] also exhibited higher levels of endogenous H₂S, as well as H₂S-generating enzyme activity (L-DES and D-DES). Moreover, low temperature induced endogenous H₂S accumulation and higher expression levels of H₂S synthesis-related genes like *CsaLCD* (*Csa2G034800.1*), *CsaDES1* (*Csa1G574800.1*) and *CsaDES2*

(*Csa1G574810.1*)^[53]. As mentioned earlier, endogenous H₂S regulates ethylene-induced ripening, which indicates that exogenous H₂S triggers endogenous H₂S production to further regulate ripening, senescence and chilling injury processes during storage.

Roles of H₂S in chilling injury

Regulation of energy metabolism

If chilling stress occurs over time, it will induce membrane phase transitions and increase membrane permeability, resulting in a series of adverse effects such as intracellular ATP (iATP) shortages, ROS accumulation and lipid peroxidation^[55]. Therefore, sufficient iATP plays a pivotal role in postharvest fruits and vegetables. During cold storage and ripening period, banana fruit treated with 0.5 mM NaHS showed higher ATP levels, and increased the activities of energy-related enzymes like H⁺-ATPase, Ca²⁺-ATPase, cytochrome C oxidase (CCO) and succinate dehydrogenase (SDH)^[56]. Thus, H₂S seemingly induced chilling tolerance of banana fruit by regulating energy metabolism. In water spinach, H₂S treatment (2.4 mM NaHS) delayed dark-induced senescence resulting from maintenance of energy status, along with higher ATP and ADP levels and lower AMP levels^[41]. Sugar is the substrate of energy metabolism. Li et al. reported that H₂S treatment (0.8 mM NaHS) not only triggered energy metabolism, but also increased the activity of enzymes involved in embden-meyerhof-parnas (EMP) and pentose phosphate pathway (PPP), such as glucokinase (GK), fructokinase (FK), glucose-6-phosphate dehydrogenase (G6PDH) and 6-phospho-gluconate dehydrogenase (6PGDH), while the

broccol treated with 0.5 mM PAG exhibited a negative role^[42]. H₂S participates in the inclusion of sugar and energy metabolism, resulting in adequate energy availability and better postharvest quality, but how H₂S regulates these enzymes still requires further investigation.

GABA, a 'sensor' for intracellular energy status, indicates plants whether to need or institute processes for more energy^[57]. GABA shunt metabolism reduces oxidative damage and ensures ATP supply by providing NADH and succinate to the tricarboxylic acid (TCA) cycle and electron transfer system (ETS)^[55]. Moreover, recent studies proposed that GABA is not only a plant metabolite, but also an endogenous signaling molecule in response to stress^[58]. NO has been reported to promote coordinated biosynthesis of polyamines (PAs), proline and GABA in banana fruit, thereby blocking the development of CI symptoms^[59]. Nevertheless, whether H₂S and GABA have a positive response to cold stress has not been reported.

Roles of H₂S in membrane integrity and membrane lipid peroxidation

Chilling attenuates the fluidity of the cytomembrane, resulting in electrolyte leakage, MDA accumulation, eventually resulting in irreversible oxidative damage and CI symptoms^[60]. Most chilling-induced changes are associated with the integrity of the cell membrane. Table 2 shows that exogenous H₂S reduces ROS accumulation and membrane lipid peroxidation during shelf life and cold storage. Supplementation with 0.5 mM NaHS conferred chilling tolerance of banana fruit by reducing electrolyte leakage, MDA and ROS, and because of higher CAT, POD, APX, SOD and GR activities. Meanwhile, H₂S regulated Δ^1 -pyrroline-5-carboxylate synthe-

Table 2. Effects of H₂S on antioxidant system in postharvest.

Materials	Treatment	Results	Reference
Delay ripening and senescence			
Tomato (Micro Tom)	0.9 mM NaHS and 1.0 g L ⁻¹ ethephon fumigation for 24 h	Enhanced CAT, POD, APX, SOD and PAL activities; reduced LOX and PPO activities; inhibited H ₂ O ₂ , O ₂ ⁻ and MDA accumulation	[27]
Fresh-cut pear (Dangshan)	2.0 mM NaHS fumigation for 6 d	Enhanced CAT, APX and POD activities; reduced PAL, PPO and LOX activities; reduced H ₂ O ₂ , O ₂ ⁻ and MDA content	[38]
Water spinach (<i>Ipomoea aquatica</i>)	2.4 mM NaHS fumigation for 8 d	Enhanced CAT, POD and SOD activities, as well as higher DPPH scavenging activity; reduced O ₂ ⁻ and MDA content; increased endogenous H ₂ S content by higher LCD and DCD activities	[41]
Grape (Kyoho)	1.0 mM NaHS fumigation for 7 d	Enhanced CAT and APX activities; reduced LOX activities; reduced H ₂ O ₂ , O ₂ ⁻ and MDA content	[47]
Apple (Red Fuji)	0.7 mM NaHS fumigation for 24 h	Enhanced CAT, SOD, PAL and GR activities; increased antioxidants content like AsA and phenolic; reduced PPO, POD and APX activities, as well as H ₂ O ₂ accumulation	[48]
Mulberry (Dianmian-1)	0.8 mM NaHS fumigation for 6 d	Enhanced DPPH and O ₂ ⁻ scavenging activity by higher CAT, POD and SOD activities; reduced O ₂ ⁻ and MDA content; increased endogenous H ₂ S levels by higher LCD and DCD activities	[52]
Fresh-cut lotus roots (Taikong Lian III)	15 μ L L ⁻¹ H ₂ S gas fumigation for 30 min	Enhanced total phenols and antioxidant capacity; higher PAL and CAT activities; lower PPO and POD activities; reduced H ₂ O ₂ and O ₂ ⁻ production	[54]
Alleviate chilling injury			
Banana (Brazil)	0.5 mM NaHS fumigation for 24 h	Enhanced CAT, POD, APX, SOD and GR activities; reduced H ₂ O ₂ , O ₂ ⁻ and MDA content; alleviated electrolyte leakage and CI	[49]
Hawthorn (<i>Crataegus monogyna</i>)	1.5 mM NaHS fumigation for 48 h	Alleviated CI; enhanced CAT, APX, SOD and PAL activities; reduced H ₂ O ₂ and MDA content; enhanced LCD and DCD activities, while increased endogenous H ₂ S levels	[50]
Kiwifruit (Jinkui)	45 μ M H ₂ S immersion for 30 min	Enhanced CAT, POD and SOD activities; reduced H ₂ O ₂ and O ₂ ⁻ content; inhibited electrolyte leakage	[51]

tase (P5CS) and proline dehydrogenase (PDH) activity that synthesized more proline^[49]. Proline, a major osmolyte, can sustain membrane permeability^[61], and stabilize antioxidant enzymes or stimulate additional ROS scavenging pathways in response to abiotic and biotic stresses^[62]. Thus, it is implied that H₂S mediates the production of ROS and proline to mitigate CI. Soluble protein and soluble sugar are also important osmotic substances. As mentioned earlier, they can be improved by H₂S. Besides, H₂S up-regulated most differentially expressed unigenes (DEGs) associated with antioxidant and energy, while down-regulating the expression of genes encoding membrane-degrading enzymes like PLD and LOX^[48]. Thus, H₂S postpones membrane lipid metabolism to reduce the damage of lipid peroxidation at the transcriptional level. In combination with the above, the roles of H₂S in CI may be achieved by regulating physiological metabolism including (i) ROS, (ii) energy, (iii) membrane lipid, (iv) proline and (v) cell wall metabolism, there are still vast areas to be studied.

Roles of H₂S in disease resistance

Inhibition of fungal growth

Fruits and vegetables often rot and deteriorate due to fungal infection during the process of transportation and storage. Recent studies have verified that H₂S-induced reduction in postharvest decay primarily operates through inhibiting fungal growth (Table 3). For example, Tang et al. reported that H₂S fumigation (1.0–2.5 mM NaHS) effectively inhibited the growth of fungal pathogens on medium and sweet potato slices in a dose-dependent manner, while reducing the incidence of black rot or soft rot^[63]. Additionally, low doses of H₂S also controlled the mycelium growth on medium and pears^[38]. How does H₂S rely on dosage to combat fungi? This study seems to explain, 0.5 mM NaHS treatment inhibited the growth of *Aspergillus niger* and

Penicillium italicum on medium and inoculated fruits, and this facilitation may be attributed to lower CAT and SOD activities and down-regulation of relative genes in *A. niger*. H₂S in high concentrations had a bactericidal effect on the growth of some foodborne pathogens, among which *Staphylococcus aureus* was more sensitive to H₂S^[64]. Taken together, the potential mechanism by which H₂S, in low doses, exert antibacterial effects is to induce ROS overproduction in fungi by attenuating antioxidant capacity, and high concentrations of H₂S principally act as a fungicide to inhibit spore germination, germ tube elongation and mycelial growth (Table 3).

In mammals and yeast, cystathionine- β -synthase (CBS) and cystathionine- γ -lyase (CSE) are key enzymes involved in H₂S synthesis from sulfur metabolism. As shown in Fig. 2a, CBS catalyzes the dehydration condensation of homocysteine (Hcy) with Ser or Cys to form H₂O or H₂S, respectively. Additionally, CBS also converts Cys to H₂S through the β -replacement reaction of Cys with thiols, and CSE produces H₂S via the γ -replacement reaction between Hcy molecules and the α,β -elimination reaction with cystine (Fig. 2b)^[65,66]. Interesting results suggest that H₂S in fungi is also produced through the transsulfuration pathway. Guo et al.^[67] reported that the deletion of the *An05g00160* gene (encoding CBS, named *cbsA*) in *A. niger* promoted Cys and GSH levels while decreasing endogenous H₂S content, which is responsible for enhancing cadmium (Cd²⁺) tolerance and pathogenicity. Pear fruits infected with the *cbsA* mutant brought about an excessive production of ROS. In accordance with these results, it is suggested that CBS mainly decomposed Cys to produce H₂S, and *cbsA* deletion harmonized redox homeostasis in *A. niger* to resist ROS burst of the host^[67]. This maybe the reason why fungi can still grow on fruits, the detailed mechanisms require further research.

Regulation of defense-related proteins

Under abiotic and biotic stresses, pathogenesis-related proteins (PRs) are triggered through the signaling network

Table 3. Effects of H₂S on disease resistance in postharvest fruits and vegetables.

Materials	Pathogens	Treatment	Effect	Reference
Pear (Dangshan)	<i>Aspergillus niger</i> <i>Penicillium expansum</i>	NaHS fumigation for 6 d (2.0 mM)	Enhanced antioxidant capacity of fresh-cut pear and inhibited ROS accumulation; increased POD activity; reduced PAL and PPO activities; inhibited mycelium growth and lesion diameter on pears	[38]
Fresh-cut sweet potato (Xushu 18)	<i>Mucor rouxianus</i> <i>Rhizopus nigricans</i> <i>Geotrichum candidum</i>	NaHS fumigation for 4 d (2.0 mM)	decreased PPO and LOX activities; lower POD activity; inhibited fungal growth and its pathogenicity	[63]
Apple (<i>Malus domestica</i>), mandarin (<i>Citrus reticulata</i>), sweet orange (<i>Citrus sinensis</i>), kiwifruits (<i>Actinidia deliciosa</i>), pear (<i>Pyrus bretschneideri</i> Rehd), tomato (<i>Lycopersicon esculentum</i>)	<i>Aspergillus niger</i> <i>Penicillium italicum</i>	NaHS fumigation for 9 d (0.5 mM)	Inhibited spore germination, germ tube elongation, mycelial growth; promoted ROS accumulation by reducing SOD and CAT activities in <i>A. niger</i> , as well as higher expression of SOD and CAT	[64]
Chinese white pear (<i>Pyrus bretschneideri</i>)	<i>Aspergillus niger</i>	Deletion of the <i>cbsA</i> Gene in <i>A. niger</i>	<i>niger</i> MA: induced higher resistance to cadmium stress and stronger infectivity to pears; increased Cys and GSH levels; decreased endogenous H ₂ S content	[67]
Peach (Mengyin)	<i>Monilinia fructicola</i>	infection with above mutant NaHS fumigation for 2 h (50 mM)	Pear fruit: increased the accumulation of H ₂ O ₂ , O ₂ ⁻ and MDA; induced larger lesion diameter <i>M. fructicola</i> : inhibited spore germination, mycelial growth and pathogenicity Peach fruit: enhanced CHT, GLU and PAL activities, while up-regulated the expression of related genes; reduced disease incidence and lesion diameter	[70]

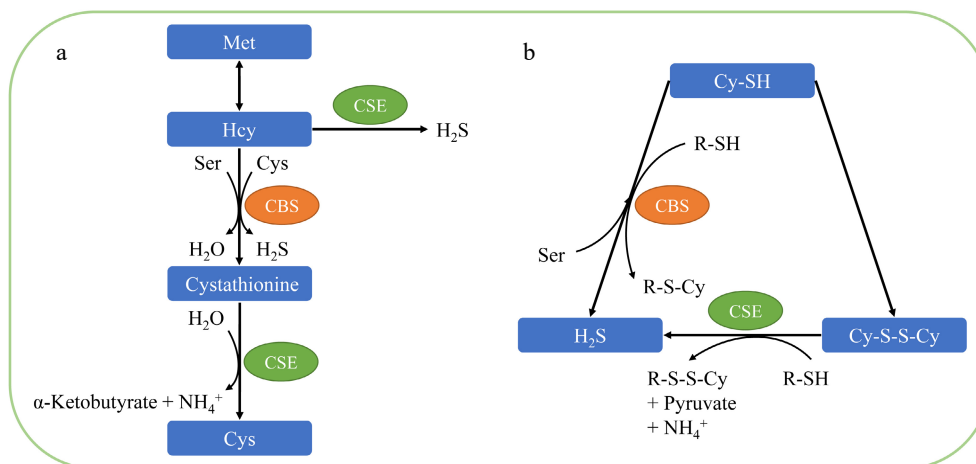


Fig. 2 Generation of H₂S in mammals and yeast. (a) H₂S is produced by sulfur metabolism, wherein CSE and CBS participate in each step. (b) Cys with thiols can be further interacted by CSE and CBS to synthesize H₂S endogenously through triggering replacement reactions. Met, methionine; Hcy, homocysteine; CSE, cystathionine-γ-lyase; CBS, cystathionine-β-synthase; Ser, serine; Cys, cysteine; Cy-SH, cysteine; Cy-S-S-Cy, disulfide.

like SA, JA, and ethylene. PRs are divided into 17 families according to their biological characteristics, among which PR-2 (β-1,3-glucanase, GLU) and PR-11 (chitinases, CHT) proteins have antifreeze^[68] and fungicide activities^[69]. GLU and CHT enzymes could decomposed the basic components of the fungal cell wall to deal with fungal infection. Zhang et al. observed that H₂S and NO in association, caused an overall reduction in the decay incidence of strawberry, associated with an increase in GLU and CHT activity^[33]. Analogous results were obtained on peach fruit, where H₂S and HT application both effectively controlled lesion development diameter by enhancing GLU, CHT and PAL activities, with accompanying higher gene expression (*U49454*, *AF20663* and *KC757351*). But HT treatment had no obvious impact on the incidence of brown rot compared with control^[70]. HT function usually acts as a H₂S scavenger, but the assumptions of how H₂S and HT promote PRs responsible in disease resistance remains unclear. Related proteins triggering in the absence of fungal infection were considered as 'PR-like' proteins such as PPO, POD and PAL^[69], also known as PRs. The metabolites lignin and phenols produced through phenylpropanoid metabolism are correlated with disease resistance, and the metabolic network composed of PAL, POD, PPO, 4-coumarate coenzyme A ligase (4CL), cinnamate-4-hydroxylase (C4H) and chalcone isomerase (CHI) co-regulate their synthesis^[71,72]. POD and PPO activity are also major drivers of the fungicidal substance quinone. Vacuum infiltrated H₂S (1.0 and 2.0 mM) increased total phenols, diminished quinone accumulation and further mitigated pericarp browning in cold-stored Litchi by altering the activity of PAL, POD and PPO. Interestingly, POD and PPO activity were higher than control at the later stages^[73]. Likewise, in fresh-cut lotus root, 15 μL L⁻¹ H₂S showed positive effects on reducing browning degree, which corresponded to higher phenolic content and antioxidant capacity^[54]. Moreover, Chen et al. found that H₂S incompletely suppressed POD activity and relative expression in fresh-cut apple, and the reduction of enzymatic browning by H₂S is not realized by reducing phenolic substrates^[48]. It is worth noting that phenols are not only regulators of disease resistance, but

also antioxidants to delay postharvest senescence. However, how H₂S induces phenolic synthesis involved in the equilibrium between antioxidant and disease resistance is vague, less is known about whether H₂S regulates other key enzymes of phenolic metabolism.

H₂S-derived protein persulfidation

H₂S triggers antioxidant system and controls ROS/RNS to harmonize the redox homeostasis, which may be pivotal to its functions^[3]. It occurs primarily through mediating PTMs of proteins that convert cysteine (Cy-SH) into R-SSH^[74]. This process was initially called S-sulfidation which can not signify hydration, but persulfidation is more suitable^[3]. Cysteine residues are the 'switch' of redox and undergo other PTMs such as S-nitrosation (NO), S-glutathionylation (GSH), S-cyanation (cyanide) and S-acylation (fatty acids) during different stress periods, which compete with H₂S for thiol (-SH) groups^[75]. Persulfidation modifies the structure, function and subcellular localization of target proteins. Proteomic analysis showed that the number of targets identified in *Arabidopsis* was in the following order: persulfidation > S-nitrosation > S-glutathionylation^[8]. This implies that H₂S-derived persulfidation plays a crucial role in biology. Two bHLH TFs, *Csa5G156220* and *Csa5G157230*, regulated the expression of the gene cluster involved in cucurbitacin C (CuC) biosynthesis^[76]. H₂S-derived persulfidation enhanced binding ability of two TFs with *bitter leaf* (*Bi*) promoter, further resulting in CuC accumulation and helping cucumber to withstand biotic and cold stress^[53]. Likewise, H₂S persulfidated mitogen-activated protein kinase MPK4 to resist cold stress in *Arabidopsis*^[77]. Jia et al. also proposed that ethylene-induced H₂S feedback regulated ethylene generation by reducing ACO activities via persulfidation under osmotic stress, and the residue Cys60 was a potential binding site in LeACO1^[78].

As mentioned previously, H₂S forms a regulatory loop with ABA and redox signaling based on protein persulfidation. There is growing evidence that the interconnection between H₂S and ROS signaling operates through persulfidation. Rboh,

also called NADPH oxidase (NOX), takes part in ROS/RNS metabolism. During sweet pepper fruit ripening, NOX activity is suppressed by S-nitrosation, nitration and S-glutathionylation^[79]. NADPH is a key cofactor in ROS/RNS metabolism, where some enzymes like GR, NOX and NO synthase-like (NOS) are NADPH-dependent to perform their functions^[80]. H₂S and NO regulate NADPH-generating enzymes based on PTMs to harmonize cellular redox homeostasis^[81]. G6PDH, 6PGDH, NADP-malic enzyme (NADP-ME) and NADP-depend

ent isocitrate dehydrogenase (NADP-ICDH) enzymes are responsible for NADPH supply. NADP-ICDH, a potential H₂S target, was probably suppressed by S-nitrosylation at Cys133 and nitration at Tyr450 during the ripening phase^[35]. Moreover, further studies showed that higher H₂S and NO levels were designed to diminish NADP-ME activity to modulate pepper fruit ripening, while 6PGDH activity was unaffected^[82]. Cumulative reports extrapolate that NADPH generation involved in fruit ripening is a good example of the association

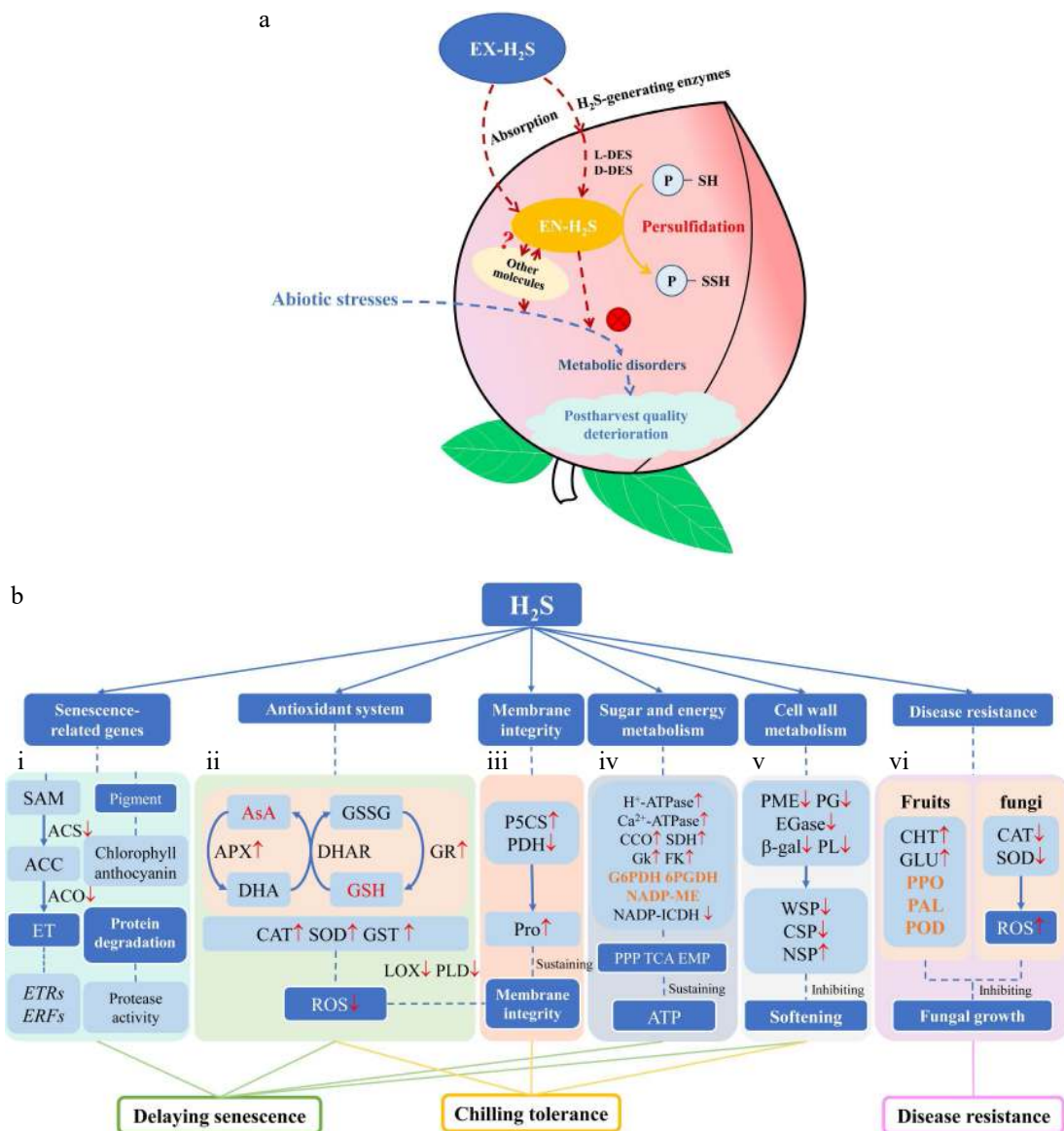


Fig. 3 Exogenous H₂S regulates numerous physiological processes to improve postharvest quality of fruits and vegetables. (a) A simple model showing a cascade of events after exogenous H₂S, which may stimulate various components of enzyme systems to maintain cellular homeostasis, but it is still not well understood how H₂S works with other molecules like ABA, NO, Ca²⁺ and ROS, etc. Broken red arrows represent processes triggered by H₂S, and broken blue arrows indicate abiotic stress-induced. (b) Various functions of H₂S in metabolic enzyme systems. (b-i) H₂S regulates related genes to delay senescence, especially ethylene (ET) biosynthesis and signaling transduction. (b-ii) The roles of H₂S in the antioxidant system, which is essential for H₂S-derived beneficial effects. (b-iii) H₂S sustains the integrity of cell membrane. (b-iv) H₂S facilitates ATP supply. (b-v) Inhibitory effect of H₂S on cell wall loosening. (b-vi) H₂S shows different mechanisms of redox homeostasis in hosts and fungi. Red arrows indicate that exogenous H₂S enhanced the enzyme activity. The orange font implies that different results are obtained after H₂S treatment, which correlates with whether the analyses are performed under cellular/subcellular or cell-free conditions (purified proteins)^[81].

between H₂S and NO via PTMs. Similarly, APX and CAT are targets of both persulfidation and S-nitrosylation at the same position, where H₂S acts upstream or downstream of NO in response to different stimuli^[83,84]. H₂S and NO co-regulate root development, stomatal closure, and programmed cell death (PCD) in plants, where ABA and ROS signaling play an integral role^[85]. These incorporating reports mean that synergistic or antagonistic properties of H₂S and NO under various stresses are based on PTMs, which is in agreement with previous studies. Nonetheless, the precise molecular mechanism of how they work remains ambiguous.

Conclusion and perspective

There is growing proof that exogenous H₂S promotes endogenous H₂S directly or by synthetic enzymes to further oppose metabolic disorders in fruits and vegetables caused by abiotic stress, which appears to affect enzyme activity via persulfidation (Fig. 3a). However, large numbers of reports suggest that H₂S can regulate various metabolic enzyme systems to improve postharvest quality (Fig. 3b), but fewer persulfidated target proteins have been identified. Many issues have emerged that require urgent attention. H₂S-induced chilling tolerance remains an extensive area of research, and it is necessary to investigate the effects of H₂S application on GABA and PAs catabolism. Whether H₂S stimulates the expression of cold responsive proteins at low temperature is also a research interest. Multiple studies have corroborated that H₂S can delay ripening and senescence and arrest ethylene-induced quality deterioration, but the accurate analysis of the cooperation between H₂S and other molecules remains obscure (Fig. 3a), especially H₂S and NO seem to provide mutual regulation. Moreover, future work should focus on the interaction network of H₂S, ABA and Ca²⁺ signaling during postharvest. H₂S inhibits fungal growth in a dose-dependent manner, the reasons why H₂S has different impacts on the redox homeostasis of fruits and fungi needs further discussion. Some enzymes involved in ripening have been reported to be persulfidated by H₂S, but it is essential to understand how H₂S recognizes specific targets. Cysteine residues also undergo other PTMs, whose interactions benefit our understanding of how H₂S responds to various stresses and explore its application value postharvest. Nevertheless, maximum residue limit (MRL) of H₂S has not been reported^[6], which limits its commercial applicability, but H₂S has huge potential for postharvest commercialization.

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Conflict of interest

The authors declare that they have no conflict of interest.

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REFERENCES

1. Wang R. 2002. Two's company, three's a crowd: can H₂S be the third endogenous gaseous transmitter? *The FASEB Journal* 16:1792–98
2. Kuschman HP, Palczewski MB, Thomas DD. 2021. Nitric oxide and hydrogen sulfide: Sibling rivalry in the family of epigenetic regulators. *Free Radical Biology Medicine* 170:34–43
3. Corpas FJ. 2019. Hydrogen sulfide: A new warrior against abiotic stress. *Trends in Plant Science* 24:983–88
4. Ziogas V, Molassiotis A, Fotopoulos V, Tanou G. 2018. Hydrogen sulfide: A potent tool in postharvest fruit biology and possible mechanism of action. *Frontiers in Plant Science* 9:1375
5. Huo J, Huang D, Zhang J, Fang H, Wang B, et al. 2018. Hydrogen sulfide: A gaseous molecule in postharvest freshness. *Frontiers in Plant Science* 9:1172
6. Ali S, Nawaz A, Ejaz S, Haider ST-A, Alam MW, et al. 2019. Effects of hydrogen sulfide on postharvest physiology of fruits and vegetables: An overview. *Scientia Horticulturae* 243:290–99
7. Li Z, Min X, Zhou Z. 2016. Hydrogen sulfide: A signal molecule in plant cross-adaptation. *Frontiers in Plant Science* 7:1621
8. Aroca A, Gotor C, Romero LC. 2018. Hydrogen sulfide signaling in plants: Emerging roles of protein persulfidation. *Frontiers in Plant Science* 9:1369
9. Wilson LG, Bressan RA, Filner P. 1978. Light-dependent emission of hydrogen sulfide from plants. *Plant Physiology* 61:184–89
10. Li Z. 2015. Analysis of some enzymes activities of hydrogen sulfide metabolism in plants. In *Methods in Enzymology, Hydrogen Sulfide in Redox Biology, Part B*, eds. Cadenas E, Packer L. 555: 356. USA: Academic Press. pp. 253–69 <https://doi.org/10.1016/bs.mie.2014.11.035>
11. Rennenberg H. 1983. Cysteine desulphydrase activity in cucurbit plants: stimulation by preincubation with L- or D-cysteine. *Phytochemistry* 22:1557–60
12. Riemenschneider A, Riedel K, Hoefgen R, Papenbrock J, Hesse H. 2005. Impact of reduced O-acetylserine(thiol)lyase isoform contents on potato plant metabolism. *Plant Physiology* 137:892–900
13. Papenbrock J, Riemenschneider A, Kamp A, Schulz-Vogt HN, Schmidt A. 2007. Characterization of cysteine-degrading and H₂S-releasing enzymes of higher plants - from the field to the test tube and back. *Plant Biology* 9:582–88
14. Léon S, Touraine B, Briat JF, Lobreau S. 2002. The *AtNFS2* gene from *Arabidopsis thaliana* encodes a Nifs-like plastidial cysteine desulphurase. *Biochemical Journal* 366:557–64
15. Hatzfeld Y, Maruyama A, Schmidt A, Noji M, Ishizawa K, et al. 2000. β -Cyanoalanine synthase is a mitochondrial cysteine synthase-like protein in spinach and *Arabidopsis*. *Plant Physiology* 123:1163–72
16. Arif Y, Hayat S, Yusuf M, Bajguz A. 2021. Hydrogen sulfide: A versatile gaseous molecule in plants. *Plant Physiology and Biochemistry* 158:372–84
17. Fang H, Jing T, Liu Z, Zhang L, Jin Z, et al. 2014. Hydrogen sulfide interacts with calcium signaling to enhance the chromium tolerance in *Setaria italica*. *Cell Calcium* 56:472–81
18. Valivand M, Amooaghaie R, Ahadi A. 2019. Seed priming with H₂S and Ca²⁺ trigger signal memory that induces cross-adaptation against nickel stress in zucchini seedlings. *Plant Physiology and Biochemistry* 143:286–98
19. Wu GX, Li DD, Sun CC, Sun SN. 2017. Hydrogen sulfide interacts with Ca²⁺ to enhance chilling tolerance of cucumber seedlings. *Chinese Journal of Biochemistry and Molecular Biology* 33:1037–46
20. Fang H, Liu Z, Long Y, Liang Y, Jin Z, et al. 2017. The Ca²⁺/calmodulin-binding transcription factor TGA3 elevates LCD expression and H₂S production to bolster Cr⁶⁺ tolerance in *Arabidopsis*. *The Plant Journal* 91:1038–50

21. Shen J, Zhang J, Zhou M, Zhou H, Cui B, et al. 2020. Persulfidation-based modification of cysteine desulphydrase and the NADPH oxidase RBOHD controls guard cell abscisic acid signaling. *The Plant Cell* 32:1000–17
22. Zhang J, Zhou MJ, Ge ZL, Shen I, Zhou C, et al. 2020. Abscisic acid-triggered guard cell L-cysteine *desulphydrase* function and in situ hydrogen sulfide production contributes to heme oxygenase-modulated stomatal closure. *Plant, Cell & Environment* 43:624–36
23. Chen J, Zhou H, Xie Y. 2021. SnRK2.6 phosphorylation/persulfidation: where ABA and H₂S signaling meet. *Trends in Plant Science* 26:1207–9
24. Zhou M, Zhang J, Shen J, Zhou H, Zhao D, et al. 2021. Hydrogen sulfide-linked persulfidation of ABI4 controls ABA responses through the transactivation of MAPKKK18 in *Arabidopsis*. *Molecular Plant* 14:921–36
25. Chen S, Jia H, Wang X, Shi C, Wang X, et al. 2020. Hydrogen sulfide positively regulates abscisic acid signaling through persulfidation of SnRK2.6 in guard cells. *Molecular Plant* 13:732–44
26. Al Ubeed HMS, Wills RBH, Bowyer MC, Vuong QV, Golding JB. 2017. Interaction of exogenous hydrogen sulphide and ethylene on senescence of green leafy vegetables. *Postharvest Biology and Technology* 133:81–7
27. Yao G, Wei Z, Li T, Tang J, Huang Z, et al. 2018. Modulation of enhanced antioxidant activity by hydrogen sulfide antagonization of ethylene in tomato fruit ripening. *Journal of Agricultural and Food Chemistry* 66:10380–7
28. Li T, Li Z, Hu K, Hu L, Chen X, et al. 2017. Hydrogen sulfide alleviates kiwifruit ripening and senescence by antagonizing effect of ethylene. *HortScience* 52:1556–62
29. Zheng J, Hu L, Hu K, Wu J, Yang F, et al. 2016. Hydrogen sulfide alleviates senescence of fresh-cut apple by regulating antioxidant defense system and senescence-related gene expression. *HortScience* 51:51–58
30. Ge Y, Hu K, Wang S, Hu L, Chen X, et al. 2017. Hydrogen sulfide alleviates postharvest ripening and senescence of banana by antagonizing the effect of ethylene. *PLoS One* 12:e0180113
31. Wang KLC, Li H, Ecker JR. 2002. Ethylene biosynthesis and signaling networks. *The Plant Cell* 14:S131–S151
32. Lin X, Yang R, Dou Y, Zhang W, Du H, et al. 2020. Transcriptome analysis reveals delaying of the ripening and cell-wall degradation of kiwifruit by hydrogen sulfide. *Journal of the Science of Food and Agriculture* 100:2280–7
33. Zhang C, Shi JY, Zhu LQ. 2014. Cooperative effects of hydrogen sulfide and nitric oxide on delaying softening and decay of strawberry. *International Journal of Agricultural and Biological Engineering* 7:114–22
34. Zhu L, Du H, Wang W, Zhang W, Shen Y, et al. 2019. Synergistic effect of nitric oxide with hydrogen sulfide on inhibition of ripening and softening of peach fruits during storage. *Scientia Horticulturae* 256:108591
35. Muñoz-Vargas MA, González-Gordo S, Cañas A, López-Jaramillo J, Palma JM, et al. 2018. Endogenous hydrogen sulfide (H₂S) is up-regulated during sweet pepper (*Capsicum annuum* L.) fruit ripening. In vitro analysis shows that NADP-dependent isocitrate dehydrogenase (ICDH) activity is inhibited by H₂S and NO. *Nitric Oxide* 81:36–45
36. Mukherjee S. 2019. Recent advancements in the mechanism of nitric oxide signaling associated with hydrogen sulfide and melatonin crosstalk during ethylene-induced fruit ripening in plants. *Nitric Oxide* 82:25–34
37. Yao G, Li C, Sun K, Tang J, Huang Z, et al. 2020. Hydrogen sulfide maintained the good appearance and nutrition in post-harvest tomato fruits by antagonizing the effect of ethylene. *Frontiers in Plant Science* 11:584
38. Hu KD, Wang Q, Hu LY, Gao SP, Wu J, et al. 2014. Hydrogen sulfide prolongs postharvest storage of fresh-cut pears (*Pyrus pyrifolia*) by alleviation of oxidative damage and inhibition of fungal growth. *PLoS One* 9:e85524
39. Li SP, Hu KD, Hu LY, Li YH, Jiang AM, et al. 2014. Hydrogen sulfide alleviates postharvest senescence of broccoli by modulating antioxidant defense and senescence-related gene expression. *Journal of Agricultural and Food Chemistry* 62:1119–29
40. Li Z, Hu K, Zhang F, Li S, Hu L, et al. 2015. Hydrogen sulfide alleviates dark-promoted senescence in postharvest broccoli. *HortScience* 50:416–20
41. Hu H, Liu D, Li P, Shen W. 2015. Hydrogen sulfide delays leaf yellowing of stored water spinach (*Ipomoea aquatica*) during dark-induced senescence by delaying chlorophyll breakdown, maintaining energy status and increasing antioxidant capacity. *Postharvest Biology and Technology* 108:8–20
42. Li D, Li L, Ge Z, Limwachiranon J, Ban Z, et al. 2017. Effects of hydrogen sulfide on yellowing and energy metabolism in broccoli. *Postharvest Biology and Technology* 129:136–42
43. Ning Z, Hu K, Zhou Z, Zhao D, Tang J, et al. 2021. *IbERF71*, with *IbMYB340* and *IbbHLH2*, coregulates anthocyanin accumulation by binding to the *IbANS1* promoter in purple-fleshed sweet potato (*Ipomoea batatas* L.). *Plant Cell Reports* 40:157–69
44. Forlani S, Masiero S, Mizzotti C. 2019. Fruit ripening: the role of hormones, cell wall modifications, and their relationship with pathogens. *Journal of Experimental Botany* 70:2993–3006
45. Zhi H, Dong Y. 2018. Effect of hydrogen sulfide on surface pitting and related cell wall metabolism in sweet cherry during cold storage. *Journal of Applied Botany and Food Quality* 91:109–13
46. Zhang W, Cao J, Fan X, Jiang W. 2020. Applications of nitric oxide and melatonin in improving postharvest fruit quality and the separate and crosstalk biochemical mechanisms. *Trends in Food Science & Technology* 99:531–41
47. Ni ZJ, Hu KD, Song CB, Ma RH, Li ZR, et al. 2016. Hydrogen sulfide alleviates postharvest senescence of grape by modulating the antioxidant defenses. *Oxidative Medicine and Cellular Longevity* 2016:4715651
48. Chen C, Jiang A, Liu C, Wagstaff C, Zhao Q, et al. 2021. Hydrogen sulfide inhibits the browning of fresh-cut apple by regulating the antioxidant, energy and lipid metabolism. *Postharvest Biology and Technology* 175:111487
49. Luo Z, Li D, Du R, Mou W. 2015. Hydrogen sulfide alleviates chilling injury of banana fruit by enhanced antioxidant system and proline content. *Scientia Horticulturae* 183:144–51
50. Aghdam MS, Mahmoudi R, Razavi F, Rabiei V, Soleimani A. 2018. Hydrogen sulfide treatment confers chilling tolerance in hawthorn fruit during cold storage by triggering endogenous H₂S accumulation, enhancing antioxidant enzymes activity and promoting phenols accumulation. *Scientia Horticulturae* 238:264–71
51. Shen Y, Wang W, Zhang W, Zhu L, Li B. 2015. Hydrogen sulfide facilitating enhancement of antioxidant ability and maintenance of fruit quality of kiwifruits during low-temperature storage. *Transactions of the Chinese Society of Agricultural Engineering* 31:367–72
52. Hu H, Shen W, Li P. 2014. Effects of hydrogen sulphide on quality and antioxidant capacity of mulberry fruit. *International Journal of Food Science & Technology* 49:399–409
53. Liu Z, Li Y, Cao C, Liang S, Ma Y, et al. 2019. The role of H₂S in low temperature-induced cucurbitacin C increases in cucumber. *Plant Molecular Biology* 99:535–44
54. Sun Y, Zhang W, Zeng T, Nie Q, Zhang F, Zhu L. 2015. Hydrogen sulfide inhibits enzymatic browning of fresh-cut lotus root slices by regulating phenolic metabolism. *Food Chemistry* 177:376–81

55. Aghdam MS, Jannatizadeh A, Luo Z, Paliyath G. 2018. Ensuring sufficient intracellular ATP supplying and friendly extracellular ATP signaling attenuates stresses, delays senescence and maintains quality in horticultural crops during postharvest life. *Trends in Food Science & Technology* 76:67–81
56. Li D, Limwachiranon J, Li L, Du R, Luo Z. 2016. Involvement of energy metabolism to chilling tolerance induced by hydrogen sulfide in cold-stored banana fruit. *Food Chemistry* 208:272–78
57. Michaeli S, Fromm H. 2015. Closing the loop on the GABA shunt in plants: are GABA metabolism and signaling entwined? *Frontiers in Plant Science* 6:419
58. Xu B, Long Y, Feng X, Zhu X, Sai N, et al. 2021. GABA signalling modulates stomatal opening to enhance plant water use efficiency and drought resilience. *Nature Communications* 12:1952
59. Wang Y, Luo Z, Mao L, Ying T. 2016. Contribution of polyamines metabolism and GABA shunt to chilling tolerance induced by nitric oxide in cold-stored banana fruit. *Food Chemistry* 197:333–39
60. Xu D, Lam SM, Zuo J, Yuan S, Lv J, et al. 2021. Lipidomics reveals the difference of membrane lipid catabolism between chilling injury sensitive and non-sensitive green bell pepper in response to chilling. *Postharvest Biology and Technology* 182:111714
61. Moukhtari AE, Cabassa-Hourton C, Farissi M, Savouré A. 2020. How does proline treatment promote salt stress tolerance during crop plant development? *Frontiers in Plant Science* 11:1127
62. Szabados L, Savouré A. 2010. Proline: a multifunctional amino acid. *Trends in Plant Science* 15:89–97
63. Tang J, Hu K, Hu L, Li Y, Liu Y, et al. 2014. Hydrogen sulfide acts as a fungicide to alleviate senescence and decay in fresh-cut sweetpotato. *HortScience* 49:938–43
64. Fu L, Hu K, Hu L, Li Y, Hu L, et al. 2014. An antifungal role of hydrogen sulfide on the postharvest pathogens *Aspergillus niger* and *Penicillium italicum*. *PLoS One* 9:e104206
65. Gadalla MM, Snyder SH. 2010. Hydrogen sulfide as a gas transmitter. *Journal of Neurochemistry* 113:14–26
66. Murphy B, Bhattacharya R, Mukherjee P. 2019. Hydrogen sulfide signaling in mitochondria and disease. *The FASEB Journal* 33:13098–125
67. Guo S, Yao G, Ye H, Tang J, Huang Z, et al. 2019. Functional characterization of a cystathionine beta-synthase gene in sulfur metabolism and pathogenicity of *Aspergillus niger* in pear fruit. *Journal of Agricultural and Food Chemistry* 67:4435–43
68. Saleem M, Fariduddin Q, Janda T. 2021. Multifaceted role of salicylic acid in combating cold stress in plants: A review. *Journal of Plant Growth Regulation* 40:464–85
69. van Loon LC, Rep M, Pieterse CMJ. 2006. Significance of inducible defense-related proteins in infected plants. *Annual Review of Phytopathology* 44:135–62
70. Wu W, Zhang C, Chen L, Li G, Wang Q, et al. 2018. Inhibition of hydrogen sulfide and hypotaurine on *Monilinia fructicoladisease* in peach fruit. In *ISHS Acta Horticulturae 1194: VIII International Postharvest Symposium: Enhancing Supply Chain and Consumer Benefits - Ethical and Technological Issues*, eds. Artés-Hernández F, Gómez PA, Aguayo E, Artés F. Cartagena, Murcia, Spain: Acta Horticulturae. pp. 257–66 <http://doi.org/10.17660/ActaHortic.2018.1194.38>
71. Mohammadi M, Kazemi H. 2002. Changes in peroxidase and polyphenol oxidase activities in susceptible and resistant wheat heads inoculated with *Fusarium graminearum* and induced resistance. *Plant Science* 162:491–98
72. Liu Q, Luo L, Zheng L. 2018. Lignins: Biosynthesis and biological functions in plants. *International Journal of Molecular Science* 19:335
73. Siddiqui MW, Deshi V, Homa F, Aftab MA, Aftab T. 2021. Inhibitory effects of hydrogen sulfide on oxidative damage and pericarp browning in harvested litchi. *Journal of Plant Growth Regulation* 40:2560–69
74. Filipovic MR, Zivanovic J, Alvarez B, Banerjee R. 2018. Chemical biology of H₂S signaling through persulfidation. *Chemical Reviews* 118:1253–337
75. Corpas FJ, Palma JM. 2020. H₂S signaling in plants and applications in agriculture. *Journal of Advanced Research* 24:131–37
76. Shang Y, Ma Y, Zhou Y, Zhang H, Duan L, et al. 2014. Plant science. Biosynthesis, regulation, and domestication of bitterness in cucumber. *Science* 346:1084–88
77. Du X, Jin Z, Liu Z, Liu D, Zhang L, et al. 2021. H₂S persulfidated and increased kinase activity of MPK4 to response cold stress in *Arabidopsis*. *Frontiers in Molecular Biosciences* 8:635470
78. Jia H, Chen S, Liu D, Liesche J, Shi C, et al. 2018. Ethylene-induced hydrogen sulfide negatively regulates ethylene biosynthesis by persulfidation of ACO in tomato under osmotic stress. *Frontiers in Plant Science* 9:1517
79. Chu-Puga Á, González-Gordo S, Rodríguez-Ruiz M, Palma JM, Corpas FJ. 2019. NADPH Oxidase (Rboh) activity is up regulated during sweet pepper (*Capsicum annuum* L.) fruit ripening. *Antioxidants* 8:9
80. Aghdam MS, Palma JM, Corpas FJ. 2020. NADPH as a quality footprinting in horticultural crops marketability. *Trends in Food Science & Technology* 103:152–61
81. Corpas FJ, González-Gordo S, Palma JM. 2021. Nitric oxide and hydrogen sulfide modulate the NADPH-generating enzymatic system in higher plants. *Journal of Experimental Botany* 72:830–47
82. Muñoz-Vargas MA, González-Gordo S, Palma JM, Corpas FJ. 2020. Inhibition of NADP-malic enzyme activity by H₂S and NO in sweet pepper (*Capsicum annuum* L.) fruits. *Physiologia Plantarum* 168:278–88
83. Palma JM, Mateos RM, López-Jaramillo J, Rodríguez-Ruiz M, González-Gordo S, et al. 2020. Plant catalases as NO and H₂S targets. *Redox Biology* 34:101525
84. Corpas FJ, González-Gordo S, Cañas A, Palma JM. 2019. Nitric oxide and hydrogen sulfide in plants: which comes first? *Journal of Experimental Botany* 70:4391–404
85. Mishra V, Singh P, Tripathi DK, Corpas FJ, Singh VP. 2021. Nitric oxide and hydrogen sulfide: an indispensable combination for plant functioning. *Trends in Plant Science* 26:1270–85



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