Effects of ultrasonic assisted marination on the mass transfer kinetics and quality of low-salt duck breast and thigh meat

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Abstract

The objective of this study was to investigate the effect of ultrasonic-assisted marination on the mass transfer kinetics and quality of duck breast meat and thigh meat. Results showed that the increase of ultrasonic power greatly accelerated the transfer of moisture and NaCl, and the highest yield was obtained by ultrasonic power of 450 W. The values of the mass transfer kinetics parameter (k_2) for weight changes improved as the ultrasonic power increased. The application of ultrasound treatment enhanced the NaCl effective diffusion coefficients (De) of duck breast and thigh meat from $0.7889-0.9472 \times 10^{-9}$ m^2/s to $1.2661-1.3775 \times 10^{-9}$ m^2/s and the highest De was found with 450 W. The treatment of ultrasound can reduce shear force and water loss of duck samples. According to the analysis of water distribution, ultrasound could decrease the T_{22} values which indicated a decrease in water mobility. Thus, ultrasonic-assisted marination could be employed as an emerging technology for various meat-curing processes.

Introduction

Static wet marination and dry curing are conventional treatments for processing Chinese duck meat products, such as Nanjing salted duck, Nanjing pressed salted duck, sauced duck, cured duck, and so on. Traditional marination and curing processes are quite time-consuming, low efficiency and probably enzymatic softening, which also have a negative impact on the shelf-life of meat products[1]. To accelerate these processes, mechanical-assisted methods like tumbling and injecting are applied by many industries to produce traditional duck products. However, these fierce mechanical forces might cause muscle breakdown and deteriorate the appearance as well as the integrity of the final products[2]. Therefore, a more moderate but effective way is urgently needed to shorten the marination time.

In recent years, ultrasonic treatment (UT) has been widely noticed as a feasible and eco-friendly strategy to improve the quality and efficiency of the marination processing[3,4]. There are four major merits of ultrasonic treatment in terms of accelerating the marination process[5–7]: 1) acoustic cavitation: ultrasonic waves generate cavitation bubbles and then propagate in the liquid. These bubbles collapse rapidly and result in internal high acoustic pressure, which facilitates the brine penetration and mass migration; 2) mechanical effects: ultrasound waves also exert mechanical vibration around the meat-brine surface, contributing to muscle tissue rupture and thus muscle tenderness; 3) thermal effects: the acoustic cavitation bubbles vibrate and explode, which leads to obvious temperature rises and potentially protein structure modification; 4) reactive oxygen species generation: ultrasonic treatment could increase H$_2$O$_2$ and ROS production, including hydroxyl radicals (·OH), hydroperoxyl radicals (·HO$_2$), and O$_2$, promoting the moderate oxidation reaction, which might positively affect the functionality of meat protein.

Ultrasonic-assisted treatment is widely applied in poultry meat production, during which the frequency, intensity, treating time are all closely related to the efficiency of mass transfer. Inguglia et al.[8] and Tong et al.[9] have verified that ultrasonic treatment could promote sodium salt, phosphate salt, and water transfer towards chicken breast meat, and under the same amount of time, the ultrasonic marinated chicken breast with increased frequency showed significantly higher sodium uptake compared to low frequency and untreated groups. Combining UT during duck preservation not only inhibits the physicochemical quality deterioration of sauced duck, but also lowers the microbiology growth at 4 °C[10].

It is noticeable that the muscle merit of duck thigh is quite different from duck breast. As reported, the muscle fiber phenotype and composition of poultry thigh meat are distinctive from breast meat, leading to various meat tissue matrices and inter-/intra- muscular fat in muscles. The leg muscle of avians is mainly composed of slow fiber, while the breast muscle is mainly composed of fast fiber. Therefore, the physical properties, tenderness, and water-holding capacity of various duck muscle types are greatly different[11]. Gong et al.[12] have also found that the duck thigh showed a much higher pH value compared to breast meat. Hence, it is easy to predicted that the mass transfer kinetics and physicochemical properties of duck meat are affected by muscle type.

Therefore, the objective of this work is to: 1) investigate the effect of ultrasonic assistant with different powers on the mass transfer of both duck breast and thigh marination process and establish marinating kinetic models; 2) to compare the shear force, drip loss, cooking loss, and water distribution of ultrasonic-marinated duck with static-marinated sample. The results of this study will provide fundamental data for efficiently producing low-salt duck products.

Materials and methods

Materials and samples collection

Samples of duck breast and duck thigh muscle were acquired from Cherry Valley ducks from large meat producers. Samples were placed at −18 °C for less than a week and thawed at 4 °C overnight before using. Before marination, the visually obvious fat and connective tissues were removed, and both breast and thigh counterparts were cut into 4 cm × 2 cm × 1 cm (8 cm³) cubes. These meat samples were treated and analyzed immediately.

Ultrasonic assisted marination

The ultrasonic marination was performed in a non-contact ultrasonic multi-faceted dispersion instrument (LC-1500W, Ningbo, China). All the breast and thigh meat cubes were randomly separated into four groups and then ultrasonic-assisted-marinated under 0, 150, 300, and 400 W power for 20, 40, 80, and 120 min at a frequency of 20 kHz. To inhibit the excessive salt penetration, the meat samples were immersed in a marination brine with 2.5% NaCl, and the ratio between samples and salt solution was set as 1:2 as preliminary determined. The bath temperature was maintained constant at 4 °C during the ultrasonic application.

Mass transfer kinetics

Changes in total weight, salt, and water content during marination

The water content of duck meat samples during ultrasonic assisted marination was determined using direct drying methods as described in GB 5009.3-2016. The salt content was carried out according to Zhang et al.[13].

To evaluate the salt content in marinated samples, the meat was minced and ~5 g paste was then removed into 50 ml centrifuge tubes. Adding 3 times deionized water and vortexing for 24 h to let the salt fully extracted into water. The total salt content was then measured by a hand-held salinity meter (PAL-SALT, Atago Co., Ltd., Japan).

Changes in total weight, water weight, and salt weight at t times were calculated using the following Eqns (1−3):

$$\Delta M_t^0 = \frac{M_t^0 - M_0^0}{M_0^0} \times 100 \tag{1}$$

$$\Delta M_t^W = \frac{M_t^0 \times X_t^W - M_0^0 \times X_0^W}{M_0^0} \times 100 \tag{2}$$

$$\Delta M_t^{NaCl} = \frac{M_t^0 \times X_t^{NaCl} - M_0^0 \times X_0^{NaCl}}{M_0^0} \times 100 \tag{3}$$

Where, M_0^0 and M_t^0 are the sample weight at times 0 and t (20, 40, 80, 120 min), respectively. Whereas, X_0^W, X_t^W, X_0^{NaCl} and X_t^{NaCl} represent the water (W) and NaCl content in a given sample at times 0 and t (20, 40, 80, and 120 min), respectively.

Mass transfer modeling and fitting evaluation

(1) To delineate the mass transfer behavior during ultrasonic-assisted marination, the following model (Eqn 4) as previously reported, was used to fit the changes in total weight, moisture content, and NaCl content[14]. Accordingly, the mass changes are linear related to the square root of marination time.

$$\Delta M_t^i = 1 + k_1 + k_2 \times t^{0.5} \tag{4}$$

Where, ΔM_t^i includes total weight changes (ΔM_t^0), water content changes (ΔM_t^w) and NaCl content changes (ΔM_t^{NaCl}). The k_1 represent the initial state at the beginning of mass transfer, and k_2 is related to diffusion kinetics, which is dependent on brine composition.

(2) The salt equilibrium equation (Eqn 5) was also applied to describe the mass transfer process[15]. The NaCl contents in the brine (y_e^{NaCl}) and aqueous phase (Z_e^{NaCl}) of the duck muscles are theoretically equal.

$$Z_e^{NaCl} = y_e^{NaCl} = \frac{\frac{M_0^{SD}}{M_0^{SS}} \times X_0^{NaCl} + y_0^{NaCl}}{\frac{M_0^{SD}}{M_0^{SS}} \times (X_0^w + X_0^{NaCl}) + (y_0^w + y_0^{NaCl})} \tag{5}$$

Where, X_0^w and X_0^{NaCl} represent the water content and NaCl content in duck muscle at 0 marination time, y_0^w and y_0^{NaCl} represent the water content and NaCl content in brine solution, and M_0^{SD}/M_0^{SS} is the ratio of weight between duck meat and brine solution.

(3) As reported by Gallart-Jornet et al.[14], to correct the effect of hydrodynamic mechanisms on the deviation of the adjusted equation from the coordinate's origin and diminish mass transfer phenomena occurring at the very beginning of the marination, the integrated solution of Fick's equation for a semi-infinite slab was introduced with an independent term K. The changes in the Z^{NaCl} and y^{NaCl} values with marination time were used to determine the effective diffusion coefficient of the duck breast and thigh samples as fitted with Eqn (6).

$$1 - Y_t^{NaCl} = 1 - \left[\frac{Z_t^{NaCl} - y_t^{NaCl}}{Z_0^{NaCl} - y_e^{NaCl}} \right] = 2 \times \left(\frac{De \times t}{\pi \times l^2} \right)^{0.5} + K \tag{6}$$

Where, $1 - Y_t^{NaCl}$ represents the reduced driving force between meat liquid phase and brine solution, Z_0^{NaCl}, Z_t^{NaCl} and Z_e^{NaCl} represent the NaCl content in a given sample at 0, t, and balance salting time and l is half of the samples' thickness.

Shear forces

After marination, the raw meat cubes were placed in a digital meat tenderness meter (C-LM3B, Northeast Agricultural University, Harbin, China) under room temperature. Each samples were shear at three locations and the averages were calculated as the shear forces.

Water loss

Drip loss

The pieces of marinated duck meat were weighed and the weight was recorded as m_1. A hook was used under the lid to hang the meat slice, which was then put in a PVA plastic bag, tightly sealed, and stored at 4 °C for 24 h. After storage, the slice of meat was carefully dabbed and weighed (recorded as m_2). The drip loss was calculated using the following equation:

$$Drip\ loss = \frac{m_1 - m_2}{m_1} \times 100\%$$

Cooking loss

The marinated duck meat samples were placed in a plastic bag after carefully weighed (recorded as m_3). The sealed samples were heated in a 80 °C water bath for 15 min until the core temperature of meat reached > 75 °C. The surface of these thermally treated meat was dabbed and weighed again (recorded as m_4). The cooking loss was calculated using the following equation:

$$Cooking\ loss = \frac{m_3 - m_4}{m_3} \times 100\%$$

Low-field nuclear magnetic resonance (LF-NMR)

The marinated duck meat sample was trimmed to 4 cm × 1 cm × 1 cm (4 cm³) cubes, wrapped with plastic film and then placed in a cylindrical glass tube (d = 15 mm) with a resonant frequency of 21.0 MHz by an LF-NMR analyzer (MesoMR23, Newsmy, Suzhou). The transverse relaxation time (T_2) was measured using a Carr-Purcell-Meiboom-Gill (CPMG) with the following parameters: temeprature = 32 °C, testing time = 200 ms, interval time = 4,000 ms, and NECH = 4,000. The resulting attenuation curve was subjected to an inversion operation with MultiExp Inv Analysis software (Niumag Electric Corporation, Suzhou, China).

Statistical analysis

Statistical analysis were carried out using one-way ANOVA with SPSS software (version 26.0, IBM Co., USA). The significant difference between treatments was determined using Duncan's multiple range test. The fitting process of mass transfer kinetics was performed using a simple linear regression (least square) using the Origin program. A statistical significance was defined as $p < 0.05$.

Results and discussion

Mass transfer kinetics

Mass changes of total, water, and NaCl weight

During the marination process, the mass transfer mainly occurred between meat samples and salt solution, and was manifested by the diffusion of small molecules (i.e. moisture,
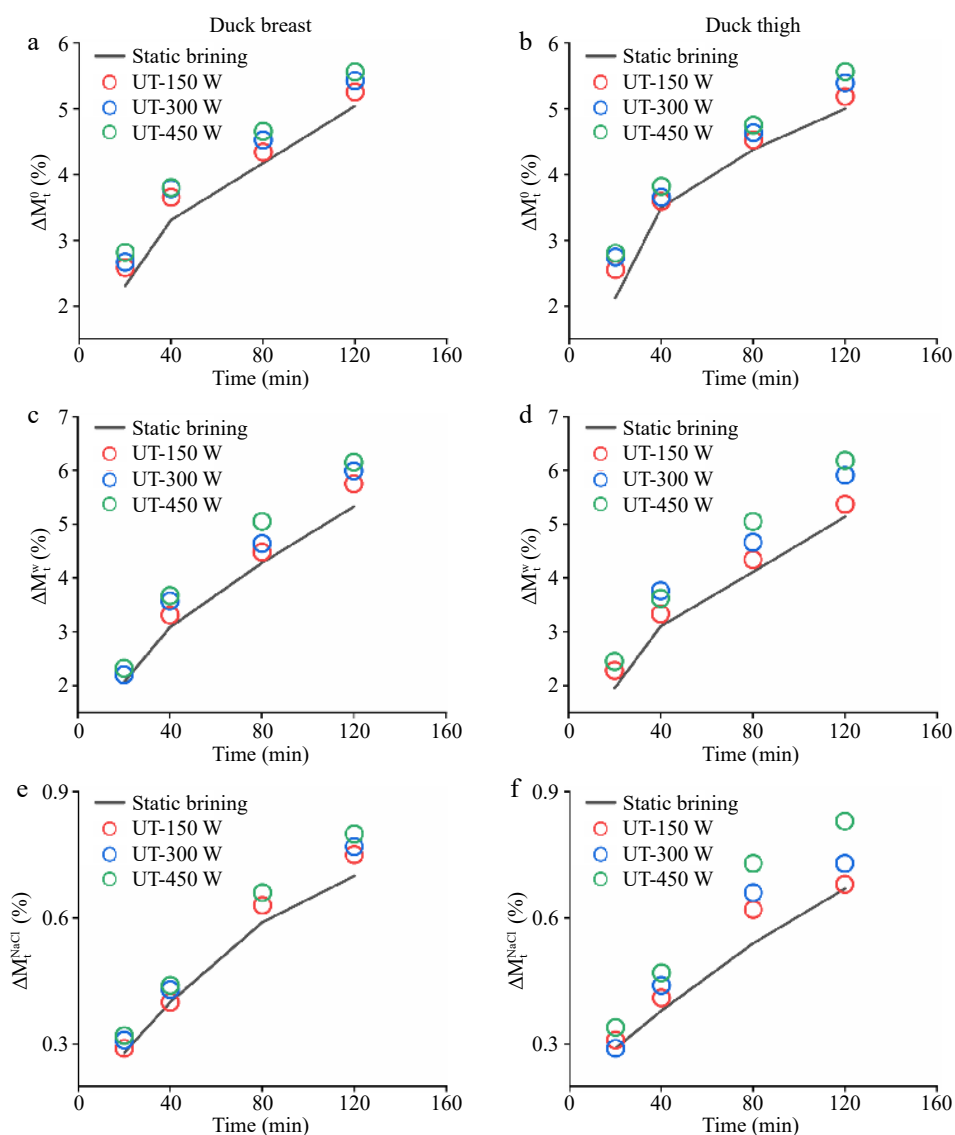

Fig. 1 (a), (b) Total weight changes (ΔM_t^0), (c), (d) water weight changes (ΔM_t^w), and (e), (f) NaCl weight changes (ΔM_t^{NaCl}) in duck breast and thigh samples with different ultrasound treatments.

salt, etc.). The rate of total weight change, water change, and NaCl change with different ultrasound conditions is shown in Fig. 1. The changes in total weight (ΔM_t^0) of samples were affected by meat types and ultrasound conditions. With the increase of ultrasonic power from 0 W to 450 W, an increment in changes of total weight was observed, and the largest change was obtained in the power of 450 W. At the end of marination (120 min), the values of ΔM_t^0 in duck breast and duck thigh were 5.56% and 5.57%, respectively. This phenomenon may be caused by the swelling of muscle fibers and the damage to tissue structure[16]. As for the content change of water (ΔM_t^w), it gradually increased with the processing of marination. Compared with the control group (0 W), the UT group with higher ultrasonic power gained more water. When the marination time was 120 min, the highest change was observed in the UT group with 450 W (duck breast: 6.15%; duck thigh: 6.18%). When the ultrasound was propagated, extreme pressures were produced, resulting in the disruption of sample

structure and the absorption of water[17]. The changes of NaCl (ΔM_t^{NaCl}) showed a similar trend with ΔM_t^w. With the increase in marination time, the concentration gradient between duck samples and the salt solution decreased. The higher the ultrasonic power, the faster the rate of NaCl penetration into the samples. The largest changes in NaCl weight were obtained in the group of 450 W. When the marination time was 80 min, compared with the control group (0 W), the value of ΔM_t^{NaCl} in the 450 W group of duck breast and thigh meat increased by 11.86% and 35.18%, respectively. Deumier et al.[18] and Ozuna et al.[19] reported similar findings that the changes in total, water, and salt weight with UT were higher than those of the non-treated samples because the cavitation effect of ultrasound could promote the penetration of NaCl.

Mass transfer kinetics

The linear relationship between the weight changes and the square root of time ($t^{0.5}$) of duck samples under different ultrasonic conditions is shown in Fig. 2. Table 1 displays the fitted
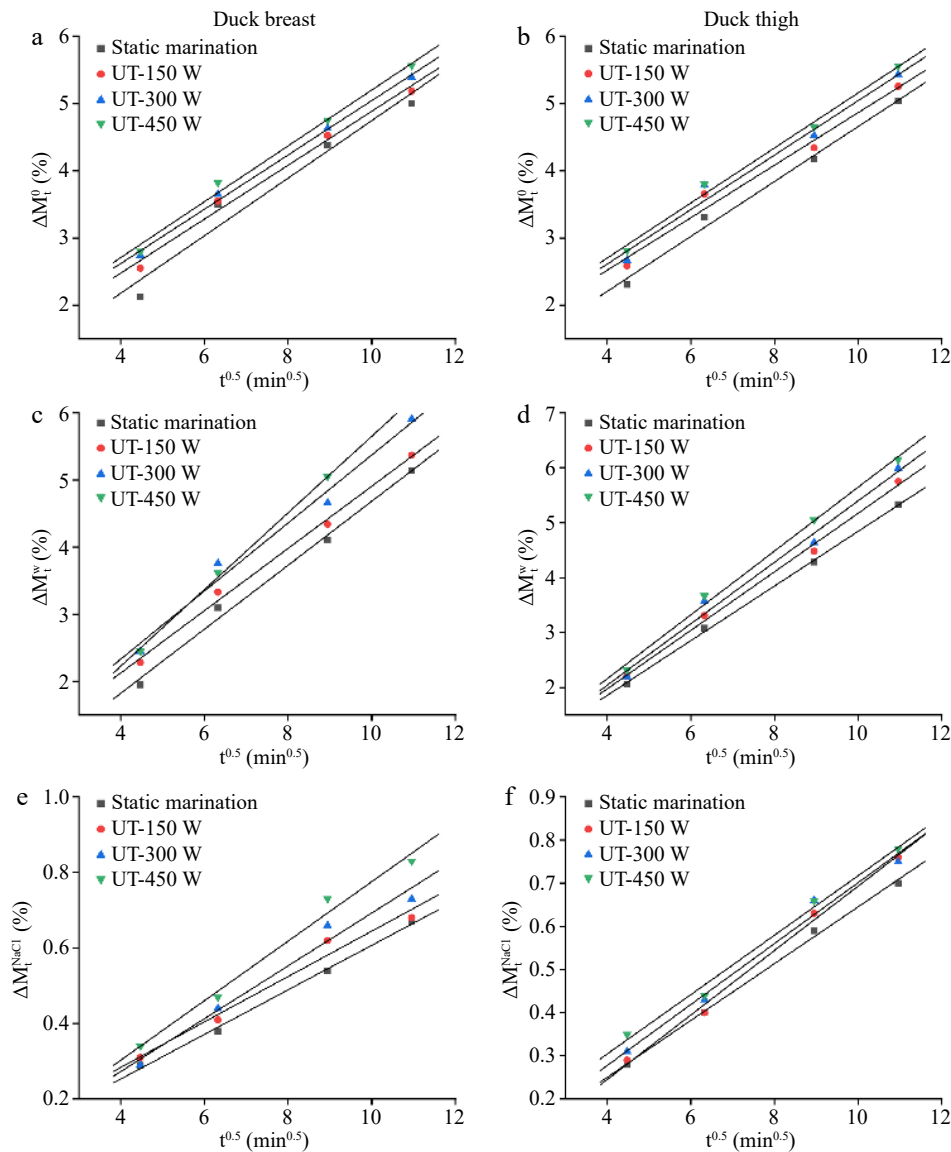

Fig. 2 Plot of (a), (b) total weight (ΔM_t^0), (c), (d) water weight (ΔM_t^w), and (e), (f) NaCl weight changes (ΔM_t^{NaCl}) vs the square root of time ($t^{0.5}$) in duck breast and thigh samples with different ultrasound treatments.

equations obtained from the liner relationship (Fig. 2) and the values of mass transfer kinetics parameters (k_1 and k_2). The coefficient of determination (R^2) of the experimental kinetic model achieved percentages of explained variance for the total, moisture, and NaCl weight changes, ranging from 95.6% to 99.8%. These results suggested that this model could be used to well fit the relationship of substances between the mass transfer process and marination time.

The k_1 value describes the behavior at the beginning of the mass transfer, which is related to the salt concentration and hydrodynamic mechanism[14]. The k_2 value is associated with the kinetics of the diffusion mechanism and the product yield and reflects the increase of total, water and NaCl weight and mass transfer diffusion efficiency[20]. As shown in Table 1, the k_1 values of ΔM_t^0 improved along with ultrasonic power increasing. When the ultrasonic power reached 450 W, k_1 exhibited the maximum values (duck breast: 0.0640; duck thigh: 0.0449). But for ΔM_t^w and ΔM_t^{NaCl}, the k_1 values of duck thigh showed an increasing and then decreasing trend with increasing ultrasonic power and the maximum values were obtained by 150 W (ΔM_t^w: −0.6942; ΔM_t^{NaCl}: −0.9573). As for k_2, the values of duck breast meat were gradually improved with the increase of ultrasonic power. The UT group with 450 W possessed maximum values of ΔM_t^0 (0.4098), ΔM_t^w (0.5813) and ΔM_t^{NaCl} (0.0755), demonstrating that ultrasound treatment could improve the diffusion efficiency of mass transfer. These results were in accordance with Zhao et al.[21]. The k_2 values of duck thigh samples showed a similar trend to breast meat. When the ultrasonic power was 450 W, the k_2 values of ΔM_t^0 (0.4157) and ΔM_t^{NaCl} (0.0789) were maximum and higher than that of the breast samples, indicating that meat type could influence the marination efficiency.

Salt diffusion coefficients (De) and calculation of other kinetic parameters

The NaCl content of duck breast and thigh meat at the equilibrium of marination could be calculated by the above-mentioned Eqn (5) as 2.633% and 2.629%, respectively. According to Fick's second law equation, the $1 - Y_t^{NaCl}$ values plotted versus $t^{0.5}$ and the De and K values are shown in Fig. 3 and Table 2, respectively. The coefficient of determination (R^2) of the NaCl transport model achieved percentages of explained variance for De, ranging from 95.0% to 99.6%. A time-independent constant K was introduced to adjust the deviation from the coordinate origin in case of any effect of mass transfer phenomena occurring at the beginning of marination[15,22]. During marination, salt diffusion efficiency (De) of meat products is linked to mass transfer resistance, which is mainly affected by the structural changes of muscle bundle[6]. As shown in Table 2, the De values of the control group in duck breast and thigh were 0.9472×10^{-9} m²/s and 0.7889×10^{-9} m²/s, respectively. The value of De improved with the increasing ultrasonic power, and the highest De value was obtained under 450 W treatment. Compared with the control group, the values of duck breast (1.2337×10^{-9} m²/s) and thigh (1.3775×10^{-9} m²/s) increased by 30.25% and 74.61%, respectively. These results suggested that ultrasonic treatment could significantly improve the De values of duck breast and thigh samples[23].

Shear forces changes

Compared to duck breast meat, duck thigh meat had lower shear stress, indicating a tender meat structure (Fig. 4). Hudaet al.[24] found the same results in that the texture of the breast part was tougher than the thigh in both Peking and Muscovy duck. The shear stress of duck breast meat was significantly decreased ($p < 0.05$) from 12.90 to 10.03 N along with ultrasonic power increasing from 0 to 450 W. While for duck thigh

Table 1. Kinetic parameters for weight changes (ΔM_t^0), water weight changes (ΔM_t^w), and NaCl weight changes (ΔM_t^{NaCl}) in duck samples with different ultrasound treatments.

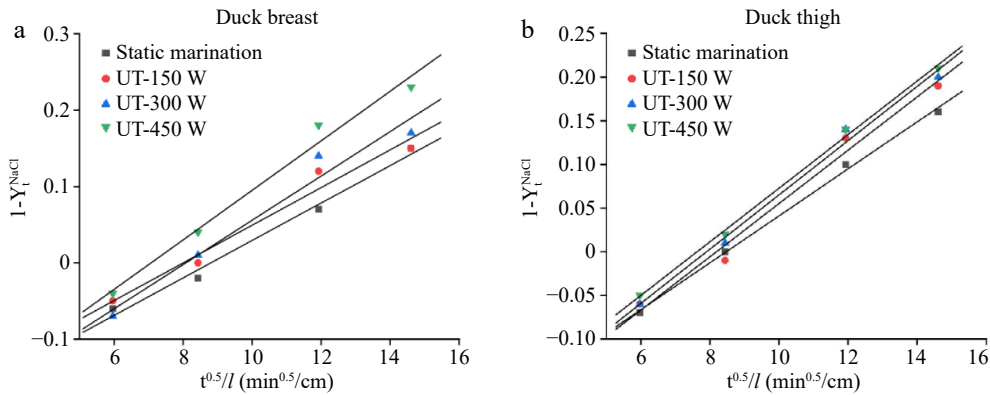| Parameters	Meat type	Treatments	Fitted equation	k_1	k_2	R^2
ΔM_t^0	Duck breast	Static marination	y = 0.3912x + 0.5712	−0.4288	0.3912	0.987
		150 W	y = 0.3910x + 0.9575	−0.0425	0.3910	0.969
		300 W	y = 0.4054x + 0.9914	−0.0086	0.4054	0.971
		450 W	y = 0.4098x + 1.064	0.0640	0.4098	0.988
	Duck thigh	Static marination	y = 0.3959x + 0.4646	−0.5354	0.3959	0.956
		150 W	y = 0.4014x + 0.8719	−0.1281	0.4014	0.984
		300 W	y = 0.4040x + 1.0041	0.0041	0.4040	0.994
		450 W	y = 0.4147x + 1.0449	0.0449	0.4157	0.988
ΔM_t^w	Duck breast	Static marination	y = 0.4979x − 0.1329	−1.1329	0.4979	0.998
		150 W	y = 0.5336x − 0.1595	−1.1595	0.5336	0.993
		300 W	y = 0.5598x − 0.1953	−1.1953	0.5598	0.982
		450 W	y = 0.5813x − 0.1632	−1.1632	0.5813	0.993
	Duck thigh	Static marination	y = 0.4620x − 0.0840	−1.0840	0.4620	0.988
		150 W	y = 0.4638x + 0.2714	−0.7286	0.4638	0.992
		300 W	y = 0.5068x + 0.3058	−0.6942	0.5068	0.974
		450 W	y = 0.5711x − 0.057	−1.0571	0.5711	0.999
ΔM_t^{NaCl}	Duck breast	Static marination	y = 0.0659x − 0.0128	−1.0128	0.0659	0.996
		150 W	y = 0.0733x − 0.0450	−1.0530	0.0733	0.991
		300 W	y = 0.0733x − 0.0199	−1.0043	0.0733	0.989
		450 W	y = 0.0755x − 0.0240	−0.9708	0.0755	0.997
	Duck thigh	Static marination	y = 0.0590x + 0.0171	−0.9829	0.0590	0.996
		150 W	y = 0.0602x + 0.0427	−0.9573	0.0602	0.964
		300 W	y = 0.0700x − 0.0071	−1.0071	0.0700	0.965
		450 W	y = 0.0789x − 0.0125	−1.0125	0.0789	0.979

Fig. 3 Reduced driving force ($1 - Y_t^{NaCl}$) vs $t^{0.5}/l$ in (a) duck breast, and (b) thigh samples with different ultrasound treatments.

Table 2. Modeling of NaCl transport in duck breast and thigh with different ultrasound treatments.

Parameters	Meat type	Treatments	De ($\times 10^{-9}$ m^2/s)	K	R^2
Duck breast	Static marination	y = 0.0269x − 0.2275	0.9472	−0.2275	0.996
	150 W	y = 0.0305x − 0.2495	1.2177	−0.2495	0.976
	300 W	y = 0.0307x − 0.2454	1.2337	−0.2454	0.988
	450 W	y = 0.0311x − 0.2339	1.2661	−0.2339	0.996
Duck thigh	Static marination	y = 0.02455x − 0.2162	0.7889	−0.2162	0.982
	150 W	y = 0.02469x − 0.1976	0.7979	−0.1976	0.955
	300 W	y = 0.02903x − 0.2345	1.1031	−0.2345	0.950
	450 W	y = 0.03244x − 0.2294	1.3775	−0.2294	0.976

meat, the 300 W and 450 W ultrasonic-treated samples exhibited similar ($p > 0.05$) shear stress (8.93 and 8.49 N, respectively), indicating a similar tenderness. It is also observed that the 300 W (10.71 N) and higher power ultrasonic-treated breast samples exhibit even lower shear stress than static marinated thigh meat (10.96 N). This suggested that the ultrasonic-treatment could not only accelerate the marination rate but also show a tenderization effect, which is in accordance with Zou et al.[7]. Previous studies have pointed out that the meat tenderness increased with power enhancement due to two main reasons: 1) helping to release tenderizing enzymes by disrupting lysosomes, and thus undermining aligned muscle structure; 2) expanding spaces between myofibrils and accelerating brine solution permeation[25,26].

Water loss changes

Drip loss

Drip loss, as one of the most important factors, is closely related to poultry texture and storage quality. The effect of ultrasonic power on the drip loss of marinated duck breast and thigh is shown in Fig. 4. It is suggested that for the breast meat, the drip loss gradually decreased along with the power increasing. The lowest drip loss (2.79%) is in the 450 W sonicated samples. This is because the ultrasound could enhance NaCl to transfer into the muscle bundle (as shown in the salt mass transfer results), thus leading more salt-soluble protein to dissolve into extracellular space, trapping more free water in the system. Pan et al.[27] also found the reduced drip loss of porcine muscle was attributed to protein hydration generated by ultrasonic-induced mild oxidation.

For the thigh meat, although the drip loss decreased from 3.21% to 2.70% when the power increases from 0 to 450 W, the 150 W and 300 W treated samples exhibited similar values ($p > 0.05$), suggesting a comparable water holding capacity during storage. A previous study has mentioned that the ultrasonic treatment could promote the salt-soluble protein to be released from muscle fiber[26] . The drip loss presented in this work is higher (~3%) than other studies reported in ultrasonic
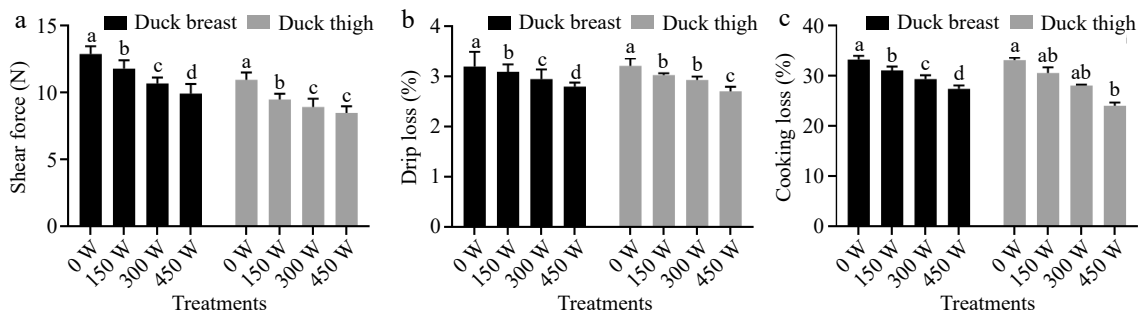

Fig. 4 Effects of different ultrasound treatments on (a) shear force, (b) drip loss and, (c) cooking loss of duck breast and thigh samples. Note: the letters a-d above the bars indicate the significant difference between samples within same muscle type ($p < 0.05$).

marinated pork and chicken meat (~1% to 2%)[26,27], which is probably induced by different meat species and brine solution composition.

Cooking loss

The cooking loss of meat can reflect the yield of final meat product, which is generally considered to decide their economic values. In addition, the meat samples exhibited greater water holding capacity during heating often showed better juiciness, higher tenderness, and improved acceptance for consumers[28]. The results of the cooking loss of duck breast and thigh meat are shown in Fig. 4. As shown, the trends of cooking loss affected by ultrasonic power are pretty similar to drip loss. For duck breast meat, the cooking loss greatly reduced from 33.21% to 27.37% under 120 min marination time while increasing ultrasonic power from 0 to 450 W, indicating a much higher production yield (raising 5.84%). This is possibly because the ultrasonic wave destroyed the muscle structure of the breast meat, promoting the dissolution of salt-soluble protein which enriched on the surface and prevented water extrusion from the system[29].

For duck thigh meat, the 0, 150, and 300 W sonicated meat samples exhibited cooking loss with no significant difference ($p > 0.05$). The 450 W treated duck thigh meat showed lowest cooking loss (24.01%) compared to all other groups. As reported previously, the breast meat of poultry obtained thicker muscle fiber and larger cross-sectional area compared to leg meat[30]. Therefore, it is assumed that the ultrasonic wave would show a more significant effect on breast meat, while the salt-soluble protein could more easily be dissolved from thigh meat fiber, thus less affected by ultrasonic treatment.

LF-NMR

Transverse relaxation time (T_2)

The insights of ultrasonic-assisted marination on the water distribution of duck breast and thigh meat were analyzed by using LF-NMR. Through observing the exchange behavior between water proteins and protein-contained protons, the mobility of water within poultry meat muscle could be evaluated[6,31]. The proton transverse relaxation time T_2 can be normalized into three populations: 1) T_{21}: ranging from 1.4 to 2.2 ms in this study, representing the bound water, which is closely restrained by proteins though hydrogen bonds or other molecular forces; 2) T_{22}: ranging from 52.2 to 67.1 ms, which refers to immobilized water that existed around epimysium, perimysium, and endomysium; and 3) T_{23}: referring to free water ranging from 428.4 to 635.2 ms, which is freely expelled from muscle cell and distributed around the meat surface.

According to Figs 5 & 6, the T_2 of duck breast was more significantly affected by ultrasonic marination compared to duck thigh, which is consistent with the results of drip loss and cooking loss. For duck breast, the T_{21} and T_{22} were shortened by ultrasonic treatment, especially 450 W treated samples. It indicated that the mobility of bound water in muscle was lowered by higher ultrasonic power. A study in pork tenderloin has shown a similar reduction trend of T_{21} when cured under ultrasound and glycerol mediation[32]. The T_{23} was prolonged in 300 W and 450 W sonicated samples, which indicated the freedom and flow ability of the free water was enhanced. This could partially be due to more salt penetration leading to osmotic pressure.

It could be seen that the static marinated thigh meat obtained shorter T_{22} than breast meat, indicating the immobilized water could be more tightly bound by the muscle system in the duck leg. This could be attributed to the wider distance between muscle fibers in breast meat. The T_{21} of duck thigh was not affected by ultrasonic treatment. Compared to 0 W and 150 W treatment samples, the 300 W and 450 W sonicated duck thigh exhibited significantly ($p < 0.05$) longer T_{22}, which is consistent with the change trends of duck breast. Guo et al.[33] have also verified that the free water relaxation time T_{22} of pork
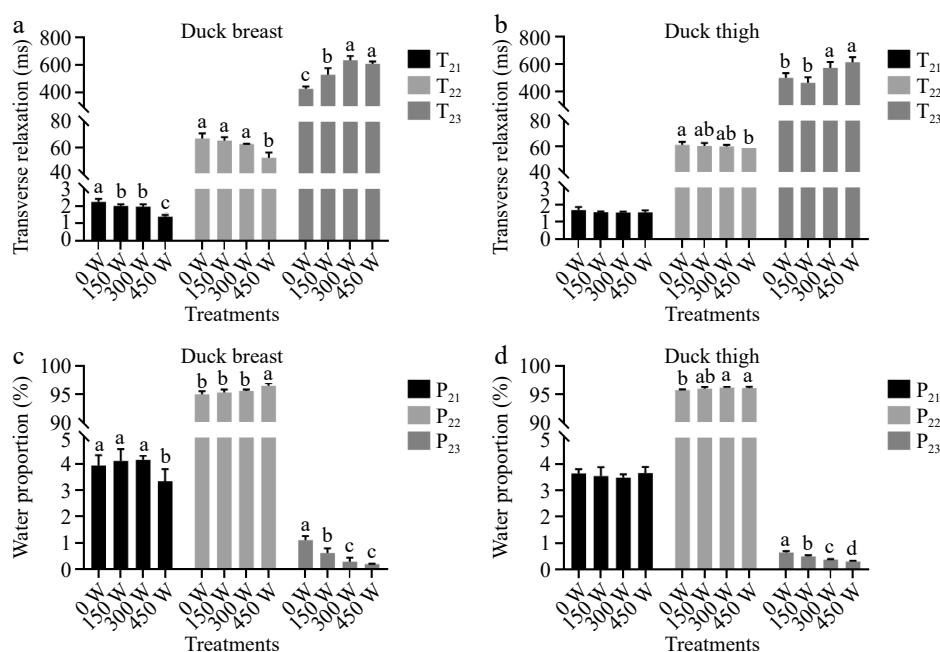

Fig. 5 (a), (b) Transverse relaxation time of water, and (c), (d) water proportion in three states of duck breast and thigh samples with different ultrasound treatments. Note: the letters a–d above the bars indicate the significant difference between samples within same muscle type ($p < 0.05$).
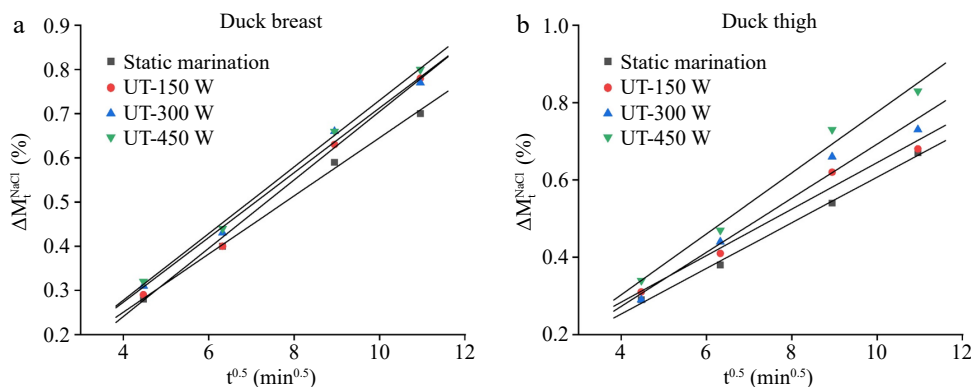
Fig. 6 The dynamic changes of transverse relaxation time (T_2) of (a) duck breast, and (b) thigh samples with different ultrasound treatments.

would significantly increase during marination, which was attributed to the infiltration of marinade into the surface layer of muscle cells.

Population of water (P_2)

The content of different kinds of water proportion is shown in Fig. 5 as P_{21} (bound water), P_{22} (immobilized water) and P_{23} (free water). After marination, the major component of water in the muscle system is immobilized water. For duck breast meat, it is seen that only 450 W ultrasonic treatment would lead to significant lower P_{21} and higher P_{22} ($p > 0.05$), and other groups exhibited similar values ($p < 0.05$).

Combined with T_2 changes trends, it is indicated that the bound water in chicken breast meat could be transferred to immobilized water with great binding ability by ultrasound, which is then well distributed in extra-myofibril space and potentially restrained by capillary force. The 450 W treated group also exhibited the lowest P_{22}, indicating a reduction of free water and thus a better water retention capacity. A previous study in pork meatball reached a similar conclusion, that the 450 W assisted cooking could significantly lower the proportion of free water[34]. The ultrasonic treatment could lower the content of free water, which indicated a better water holding capacity. This result is consistent with changes in drip loss and cooking loss.

Conclusions

The present study revealed how ultrasonic-assisted marination affects the quality of duck breast and thigh meat. The results showed that ultrasonic treatment significantly influenced the mass transfer kinetics during the marination process. With the increase of ultrasonic power, the changes of total weight, moisture content, and NaCl content continuously increased. The maximum effective diffusion coefficient (De) of duck breast and thigh meat was both obtained with the power of 450 W. Compared with static marination, UT could accelerate the marination process and shorten the marination time from 120 min (0 W) to 80 min (450 W) for achieving similar salt content. The ultrasonic treatment could not only accelerate marination rate, but also showed a tenderizing effect. The reduction of drip loss and cooking loss represented the improvement of water holding capacity of duck samples and corresponded with the weight changes of NaCl. The distribution of water analysis confirmed the reduction of water loss

with ultrasonic treatment, indicated by the decrease in relaxation times T_{22}. Overall, ultrasonic-assisted marination is a potential alternative for accelerating the efficiency of brining and improving meat quality.

Author contributions

The authors confirm contribution to the paper as follows: methodology, data curation: Shao J, Ding R, Zhao X; formal analysis, writing - original draft: Shao J; supervision, validation: Sheng C; funding acquisition: Sheng C, Zhao X; conceptualization, supervision: Xu X; project administration, writing - review & editing: Zhao X. All authors reviewed the results and approved the final version of the manuscript.

Data availability

The datasets generated during and/or analyzed during the current study are available from the corresponding author on reasonable request.

Acknowledgments

Conflict of interest

The authors declare that they have no conflict of interest.

Dates

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