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# Genetic selection for growth, wood quality and resin traits of potential Slash pine for multiple industrial uses

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# Abstract

This study aims to understand the genetic basis of key industrial traits in Slash pine (*Pinus elliottii* Engelm. var. *elliottii*) to enhance improvement efficiency. Detailed analyses were conducted on inter-family differences, genetic parameters, correlations, and breeding values (BVs) for growth, wood properties, and resin traits of Slash pine planted in Changle Forest Farm of Hangzhou, leading to the identification of elite families. It indicates that growth traits are primarily influenced by environmental effects, while wood properties exhibit a significant impact of genetic effects. The variation in resin traits arises from both genetic and environmental effects. Notably, Beta-pinene exhibits the highest variability and genetic gains among the traits analyzed. The family heritability ranges for growth, wood properties, and resin traits are 0.543–0.794, 0.870–0.885, and 0.285–0.695, respectively. Significant positive correlations are evident between growth and resin traits, while a negative correlation is observed between growth and wood properties. Elite families identified through single-trait and multi-trait combined selection are 8–126 for growth traits, 2–325 and 0–373 for wood properties, and 8–131 for resin traits. The average genetic gains for these elite families are 7.44%, 7.17%, and 8.84%, respectively. These findings provide valuable insights for high-generation breeding of Slash pine and lay a genetic foundation for formulating effective breeding strategies for conifers.

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# Introduction

Wood, known for its renewable and eco-friendly properties, serves diverse purposes in construction, furniture, and pulp production<sup>[1]</sup>. Key growth parameters like diameter at breast height (DBH) and tree height (Ht), alongside wood properties such as wood density (WD) and modulus of elasticity (MOE), are essential for breeding programs targeting industrial production<sup>[2,3]</sup>. Traditional assessment methods for these traits are costly, time-intensive, and destructive, posing challenges in forestry research<sup>[4]</sup>. However, non-destructive wood quality assessment technologies like Pilodyn and Resistograph offer a promising solution, enabling rapid and accurate target detection in a cost-effective and high-throughput manner<sup>[5,6]</sup>. Resin, a viscous fluid mixture of secondary metabolites comprises turpentine rich in monoterpenes and sesquiterpenes, and rosin predominantly composed of diterpenes<sup>[7]</sup>. Turpentine finds applications in fragrance, pharmaceuticals, and food additives, while rosin is extensively used in agrochemicals and adhesives<sup>[8,9]</sup>. Resin derivatives have diversified applications in biodegradable batteries, green plastics, and petroleum alternatives<sup>[9,10]</sup>. Each resin component serves distinct purposes in industrial applications<sup>[11,12]</sup>. For example, pinene, the predominant turpentine, is utilized in natural insecticides, pharmaceuticals, and food additives<sup>[13-16]</sup>. Among pinene isomers, Alpha-pinene is the most abundant, while Beta-pinene holds significant value in the chemical industry<sup>[17]</sup>. Slash pine (Pinus *elliottii* Engelm. var. *elliottii*) resin, characterized by its high Beta-pinene content is of considerable industrial value<sup>[18,19]</sup>.

Genetic variation is crucial for genetic improvement, providing insights into trait breeding potential<sup>[12]</sup>. In forestry genetic improvement, breeders often aim to enhance multiple traits simultaneously. Therefore, investigating the relationship between growth traits and wood properties is crucial. Studies have indicated a positive genetic correlation between growth traits and wood properties in P. taeda<sup>[20]</sup> and P. massoniana<sup>[21]</sup>. However, the correlation between growth traits and WD in P. contorta<sup>[22]</sup> is not significant, contrasting with other research suggesting a negative correlation between growth trait and wood properties in P. taeda<sup>[23]</sup>, Larix kaempferi<sup>[24]</sup>, and P. radiata<sup>[25]</sup>. Consequently, achieving rapid wood maturation through genetic improvement for growth traits may potentially have adverse effects on wood quality<sup>[26,27]</sup>. Additionally, research on resin traits has revealed significant negative correlations between Alpha- and Beta-pinene. Furthermore, resin yield and resin basic density exhibit a highly significant positive correlation, while turpentine content shows a highly significant negative correlation with resin basic density<sup>[28]</sup>. Understanding these connections is crucial for implementing multitrait combined selection, optimizing breeding strategies, and achieving balanced improvements in growth-wood-resin traits<sup>[18]</sup>.

However, economically valuable traits in tree breeding programs typically exhibit polygenic variation and are strongly influenced by environmental factors. Almost all commercially important tree species have reported significant Genotype-by-Environment interactions (G  $\times$  E), including Scots pine<sup>[29]</sup>, Eucalyptus<sup>[30]</sup>, Douglas fir<sup>[31]</sup>, Norway spruce<sup>[32]</sup>, and Poplar<sup>[33]</sup>. Multi-trait combined selection has the potential to mitigate the impact of the environment on genotype stability and expedite the breeding process<sup>[34]</sup>. Nonetheless, addressing linkage disequilibrium (LD) while simultaneously selecting for multiple desirable traits has led to various methods for multi-trait combined selection<sup>[35]</sup>, such as index selection<sup>[36]</sup>, sequential selection<sup>[37]</sup>, and multi-factor comprehensive evaluation<sup>[38]</sup>. Principal component analysis (PCA) is one such method, revealing the intrinsic characteristics of test materials by extracting factors from the multifactorial relationships between components and performance, which has demonstrated efficacy in Larix kaempferi<sup>[39]</sup> and Camellia chekianaoleosa<sup>[40]</sup>.

Slash pine plays a crucial role in providing wood and resin material, with genetic improvement efforts primarily targeting growth rates, wood quality, and oleresin yield  $(OY)^{[41]}$ . This study aims to: (1) investigate the varied G × E effects on growth, wood properties, and resin traits; (2) estimate the heritability, correlations, and breeding values of these traits; and (3) employ multi-trait combined selection to identify elite families. These findings may enhance the efficiency of genetic improvement for key industrial traits in Slash pine and accelerate its high-generation breeding process.

# **Materials and methods**

# Plant materials and experimental design

The Slash pine clonal seed orchard (located at 30°20' N, 119°50' E) comprises 300 elite clone genotypes selected from Slash pine provenance test forests in seven provinces: Guangdong, Fujian, Jiangxi, Jiangsu, Hunan, Hubei, and Zhejiang. These clones, all exhibiting elite growth performance and unrelated to each other, were propagated in April 1976 by grafting robust branches from the crown of selected elite trees onto 2year-old Slash pine rootstocks using the method of cambial layer grafting.

In 1992, seeds from the clonal families of the orchard were collected, and seedlings were raised in 1993. In 1994, a clonal stand test forest was established at coordinates 30°27' N, 119°49' E. The test forest comprises 33 families, arranged in a completely randomized design with six offspring in single-row plots and six replications (blocks), with trees spaced 2 m apart within rows and rows spaced 3 m apart. The local annual average precipitation is 1,480 mm, and the annual average temperature is 17.0 °C. The planting site is characterized by gently sloping hills with yellow-red soil of moderate fertility. Thinning was carried out in the test forest after 10 years of establishment. The test forest had never been tapped for resin before the commencement of this experiment.

In July 2019, phenotypic surveys were conducted on four blocks of the test forest. The three trees with the largest DBH were measured in each plot, and trees with obvious damage were excluded. This resulted in a total of 240 individual phenotype data collected (Supplemental Table S1).

#### Growth traits phenotyping

In this study, the phenotypic traits and survey methods for the growth traits of Slash pine are as follows: (1) DBH: Measured with a caliper at a height of 1.3 m from the ground on the trunk.

(2) Ht and height under the crown (Huc): Measured using a height measuring pole.

(3) Crown width: Diameter of the crown measured in both east-west and north-south directions, with the average result taken and the area calculated.

(4) Average annual ring width (ARW): ARW was measured using the Resistograph-IML-RESIPD500 (IML-RESI GmbH, Taufkirchen, Germany). The drill was inserted vertically at the tree's DBH, recording depth and amplitude for each 0.1 mm. Peaks and valleys correspond to annual growth cycles. Data selection excluded bark-induced irregularities, focusing on distinct resistance peaks. The ARW and resistance value (Mountain peak, MP) were calculated. Due to needle height, the first-year ring is often unclear, starting ring counting from the outermost. Each tree's data covered 24 years for RW and yearly MP. ARW and AMP were recorded, followed by data analysis using PDToolsPro software. The detailed operational method has been previously described in our earlier research<sup>[42]</sup>.

#### Wood properties phenotyping

(1) WD: Using a growth increment borer with a diameter of 5 mm, cores were extracted from the trees at breast height in the north-south direction, ensuring that the borehole penetrated through the pith of the tree. Basic density ( $\rho$ ) was determined using the saturated moisture content method<sup>[43]</sup>:

$$\rho = \frac{1}{\frac{M_{\rm w}}{M_{\rm s}} - 0.3464} \tag{1}$$

In Eqn (1),  $\rho$  is the basic density of wood (g-cm<sup>-3</sup>),  $M_w$  is the mass of the wood core when saturated with water,  $M_d$  is the mass of wood core when completely dry.

(2) Determination of stress wave velocity v and *MOE*: The Hitman ST300 (Fibre-gen, Christchurch, New Zealand) was utilized to measure the v of the samples. The specific method is detailed in the research findings of Zhang et al.<sup>[44]</sup>. Subsequently, the *MOE* of the samples was calculated using Eqn (2) based on the  $\rho$  and v.

$$MOE = \rho v^2 \tag{2}$$

(3) Pilodyn (Pd) detection: Using Pilodyn (6J, PROCEQ, Switzerland), cores were extracted from the lower 2 cm of the stem of individual trees in both the south and north directions. Two Pd resistance values (Ps and Pn) were measured, and only values with a difference of no more than 2 mm were considered valid. Otherwise, the two values with the smallest difference were selected. The average resistance values in the south and north directions were denoted as Pd.

(4) The AMP data acquisition method is as described in the section 'Growth traits phenotyping'.

#### **Resin traits phenotyping**

(1) OY: A specialized plastic tube with a diameter of 1.8 mm and a volume of 15 mL is fixed in the borehole on the sunny side of the trunk. After waiting for 24 h, the resin is collected, and then its yield is measured using a balance. The detailed installation method is described in Li et al.<sup>[45]</sup>.

(2) Relative amount of resin components: Gas chromatography experiments were carried out with a GC 6890 gas chromatograph (Agilent 5975B, Santa Clara, CA, USA) equipped with a DB-5MS capillary column cross-linked with 5% Ph Me

silicone (60 m/0.25 mm/0.25 mm) and coupled with a Hewlett Packard GC 5975B mass spectrometer. The qualitative and quantitative analysis of resin composition with the chromatographic conditions was as follows: GC: 0.05 g of pine resin was dissolved in 0.5 mL of ethyl alcohol containing 50  $\mu$ L tetramethylammonium hydroxide. The initial column temperature was 60 °C, held for 2 min, increased at 8 °C·min<sup>-1</sup> to 80 °C, and reached a maximum of 280 °C at a rate of 2 °C·min<sup>-1</sup> for 5 min. The helium gas flow was set at 1 mL·min<sup>-1</sup>. The temperature of the injector was 260 °C, and the volume was 1  $\mu$ L with a 1/50 split ratio. Mass spectra were recorded under electron impact ionization at an electron energy of 70 eV in the range from m/z 30 to 600 along with solvent delay for 3 min.

Resin compositions were identified by matching experimental fragmentation patterns in mass spectra with the NIST08 database through the data processing system of Agilent Chem Station and then compared with the relevant literature. Monoterpene content was determined by isobutylbenzene content, and diterpene content was determined by heptadecanoic acid content. The resin component contents were calculated by comparing their peak areas. We then selected four components believed to have broad industrial utility for subsequent analysis, including two monoterpenes (Alpha- and Beta-pinene) and two diterpenes (Abietic and Levoprimaric acid).

#### Statistical analysis method

The linear model for each observation of the resin component trait  $y_{ijk}$  in each tree was shown as follows:

$$y_{ijk} = \mu + f_i + b_j + f b_{ij} + e_{ijk} \tag{3}$$

Where the observed values  $(y_{ijk})$  for individual plants within each family (*i*), block (*j*), and specific plant (*k*) are considered, The  $\mu$  represents the average value across all observations. Family effects are denoted by  $f_i$ . Similarly,  $b_j$  refers to block effects. The interaction effect between family and block is represented by  $f_{bij}$ . Environmental error effects ( $e_{ijk}$ ) encapsulate random variations and unforeseen factors affecting individual plant observations.

The genetic variation analysis employed the Restricted Maximum Likelihood (REML) method to fit the Generalized Linear Mixed Model (GLMM). Cuevas et al. provided a detailed description of this approach, wherein the model equations are derived by stacking the vectors for all individual plants<sup>[46]</sup>:

$$y = Xm + Z_1f + Z_2b + Z_3fb + e$$
 (4)

Where the vector y represents the observed values of the overall phenotype. The vector m denotes the population mean values, while b, f, and e represent the vectors of block effects, family effects, family and block interaction effects, and random error effects, respectively. The design matrices X,  $Z_1$ ,  $Z_2$  and  $Z_3$  are corresponding correlation matrices used to link the observed values with their respective effects. We define the expected value vector (E) and the discrete matrix (Var) as:

$$E[\mathbf{y}] = \mathbf{X}\mathbf{m} \tag{5}$$

$$Var[\mathbf{f}] = \mathbf{Z}_1 \otimes \mathbf{F}_0 \tag{6}$$

$$Var[\mathbf{b}] = \mathbf{Z}_2 \otimes \mathbf{B}_0 \tag{7}$$

$$Var[fb] = Z_1 Z_2 \otimes F_0 B_0 \tag{8}$$

$$Var[e] = Z \oplus R_0 \tag{9}$$

and

$$\mathbf{F}_{0} = \begin{bmatrix} \sigma_{f_{1}}^{2} & \sigma_{f_{1}f_{2}} \\ \sigma_{f_{2}f_{1}} & \sigma_{f_{2}}^{2} \end{bmatrix}$$
(10)

$$\mathbf{B}_0 = \begin{bmatrix} \sigma_{b_1}^2 & \sigma_{b_1 b_2} \\ \sigma_{b_2 b_1} & \sigma_{b_2}^2 \end{bmatrix}$$
(11)

$$F_{0}B_{0} = \begin{bmatrix} \sigma_{f_{1}b_{2}}^{2} & \sigma_{f_{1}b_{2}} \\ \sigma_{f_{2}b_{1}} & \sigma_{f_{2}b_{2}}^{2} \end{bmatrix}$$
(12)

$$\mathbf{R}_{0} = \begin{bmatrix} \sigma_{e_{1}}^{2} & \sigma_{e_{1}e_{2}} \\ \sigma_{e_{2}e_{1}} & \sigma_{e_{2}}^{2} \end{bmatrix}$$
(13)

Where the  $\otimes$  and  $\oplus$  are vector product and vector addition respectively;  $\sigma_{f_i}^2$ ,  $\sigma_{b_i}^2$ ,  $\sigma_{f_ib_i}^2$  and  $\sigma_{e_i}^2$  represent the variances of the block effect, family effect, family × block interaction effect, and environmental error effect for trait *i*, respectively;  $\sigma_{b_ib_j}$ ,  $\sigma_{f_if_j}$  and  $\sigma_{e_ie_j}$  are the covariances effects of block, family and environmental errors for traits *i* and *j*. The variance component of the model was used to calculate the family mean heritability (*H*<sup>2</sup>) and individual narrow-sense heritability (*h*<sup>2</sup>):

$$H_{i}^{2} = \frac{\sigma_{f_{i}}^{2} + \sigma_{b_{i}}^{2}}{\sigma_{f_{i}}^{2} + \sigma_{b_{i}}^{2} + \sigma_{f_{i}b_{i}}^{2}/n_{b} + \sigma_{e_{i}}^{2}/n_{b}n_{k}}$$
(14)

$$h_i^2 = \frac{4(\sigma_{f_i}^2 + \sigma_{b_i}^2)}{\sigma_{f_i}^2 + \sigma_{b_i}^2 + \sigma_{f_i b_i}^2 + \sigma_{e_i}^2}$$
(15)

Phenotypic correlation  $r_{p_{ij}}$  and genetic correlation  $r_{g_{ij}}$  of traits *i* and *j*:

$$r_{p_{ij}} = \frac{\sigma_{f_{ij}} + \sigma_{b_{ij}} + \sigma_{e_{ij}}}{\sqrt{\left(\sigma_{f_i}^2 + \sigma_{b_i}^2 + \sigma_{f_i b_i}^2 + \sigma_{e_i}^2\right)\left(\sigma_{f_j}^2 + \sigma_{b_j}^2 + \sigma_{f_j b_j}^2 + \sigma_{e_j}^2\right)}}$$
(16)

$$\sigma_{g_{ij}} = \frac{\sigma_{f_{ij}} + \sigma_{b_{ij}}}{\sqrt{\left(\sigma_{f_i}^2 + \sigma_{b_i}^2\right)\left(\sigma_{f_j}^2 + \sigma_{b_j}^2\right)}}$$
(17)

Among them,  $\sigma_{f_{ij}}$ ,  $\sigma_{b_{ij}}$ ,  $\sigma_{e_{ij}}$  represent the covariance of family effect, block effect and environmental errors effect for traits *i* and *j*;  $\sigma_{f_i}^2$  and  $\sigma_{f_j}^2$  represent the family variance estimate of traits *i* and *j*;  $\sigma_{b_i}^2$  and  $\sigma_{b_j}^2$  represent the block variance estimate of traits *i* and *j*;  $\sigma_{e_i}^2$  and  $\sigma_{e_j}^2$  represent the environmental errors variance estimate of traits *i* and *j*;  $\sigma_{e_i}^2$  and  $\sigma_{e_j}^2$  represent the environmental errors variance estimate of traits *i* and *j*;  $\sigma_{e_i}^2$  and  $\sigma_{e_j}^2$  represent the environmental errors variance estimate of traits *i* and *j*.  $\sigma_{b_i f_i}^2$  and  $\sigma_{b_j f_j}^2$  represent the variances of family × block interaction effect; The  $n_b$  and  $n_k$  are block numbers and tree numbers per family, respectively.

Coefficient of variation (CV/%):

$$V = 100\overline{x}/\sigma \tag{18}$$

Coefficient of phenotypic variation  $(l_p)$  and Coefficient of genetic Variation  $(l_q)$ :

CV

$$I_p = \sqrt{\sigma_p^2 / \bar{x}} \tag{19}$$

$$I_g = \sqrt{\sigma_g^2 / \overline{x}} \tag{20}$$

Among them,  $\sqrt{\sigma_p^2}$  is the square root of the phenotypic variance component;  $\sqrt{\sigma_g^2}$  is the square root of the genetic variance component;  $\overline{x}$  is the trait mean value.

The genetic gain ( $\Delta G$ ) is estimated by equation (17) :

$$\Delta G = i \cdot H \cdot \sigma_g / t \tag{21}$$

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Among them, *i* is the selection intensity; *H* is represented by the square root of the heritability;  $\sigma_g$  represents the square root of the additive genetic variance; *t* represents the breeding period. In this study, we employed varying selection intensities, specifically 1.40, 1.75, and 2.06, to simulate diverse selective pressures.

Breeding value (BV) is estimated by the best linear unbiased prediction (BLUP) model.

PCA was employed in the process of conducting multi-trait combined selection of elite families. The principal components (PCs) for growth traits, wood properties, and resin traits were derived from the breeding values associated with each trait. Subsequently, we computed the loadings of each trait across various PCs, along with eigenvalues, eigenvectors, contribution rates, and cumulative contribution rates for each PC. Elite families were selected based on the comprehensive scores obtained from the PC factors ( $F_{total}$ ) of each family.

# **Statistical software**

All data analyses were based on R software<sup>[47]</sup>. The analysis of variance was calculated by the function aov, the pedigree relationship matrix was calculated by the R language 'pedigreemm' software package<sup>[48]</sup>, and the genetic parameters and breeding values were estimated using the R language 'sommer' software package<sup>[49]</sup>. PCA analysis was performed by the function prcomp and the software package 'psych'<sup>[50]</sup>. All result visualizations were implemented using the R language 'ggplot2' software package<sup>[51]</sup>.

# Results

# Descriptive statistics and variance analysis of various traits

Table 1 presents statistical results for growth, wood properties, and resin traits in the progeny test stand of Slash pine halfsib families. Growth traits, including DBH, Ht, Huc, Crown, and ARW, had average values of 19.766 cm, 16.773 m, 9.439 m, 3.469 m<sup>2</sup>, and 4.254 mm, respectively, with CV ranging from 15.724% to 26.931%. For wood properties, Pd, MOE, and AMP had average values of 19.836 mm, 3.006 GPa, and 0.415%, with CVs ranging from 10.335% to 29.488%. Regarding resin traits, average values for OY, Alpha-pinene, Beta-pinene, Abietic acid,



and Levopimaric acid were 1.838 g, 14.790  $\mu$ g·g<sup>-1</sup>, 10.661  $\mu$ g·g<sup>-1</sup>, 8.183  $\mu$ g·g<sup>-1</sup>, and 21.631  $\mu$ g·g<sup>-1</sup>, respectively. Betapinene exhibited the highest CV at 42.126%, while levopimaric acid had the lowest at 14.868%. Significant differences (p < 0.05) were observed in all traits among families. Moreover, except for DBH and ARW, all other traits were significantly influenced by the block effect. Additionally, Ht, Crown, AMP, Alphapinene, and Beta-pinene exhibited effects from family, block, and their interaction.

The variation in growth traits primarily stems from the interaction between family and block effects (Fig. 1). Notably, the largest effect influenced by environmental factors is the Crown variance at 66.28%, indicating a predominant influence of the environment on Slash pine growth traits. Conversely, variation in wood properties is primarily attributed to family effects, with a proportion of 78.08% for AMP, suggesting a lesser impact of environmental factors. For resin traits, environmental effects emerge as the primary source of variation, while the OY trait shows nearly equal contributions from genetic and environmental factors at 47.40% and 48.33%, respectively.

# Estimation of genetic parameters of various traits

Based on the variance components, genetic parameters for various traits were estimated (Table 2). Among the three types



**Fig. 1** The proportion of variance components of different effects for each trait.

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Traits	Mean	S D	<b>C</b> 1/10/	F-statistic			
		50	CV/ 70	Family	Block	Block × family	
DBH (cm)	19.766	4.026	20.370	1.518*	0.616	0.894	
Ht (m)	16.773	2.637	15.724	2.901**	7.738**	1.902*	
Huc (m)	9.439	2.334	24.730	1.393*	5.102**	0.875	
Crown (m <sup>2</sup> )	3.469	0.934	26.931	1.529*	3.362*	1.463*	
ARW (mm)	4.254	0.818	19.228	1.477*	0.427	0.622	
Pd (mm)	19.836	2.050	10.335	2.637**	4.351**	1.007	
MOE (Gpa)	3.006	0.368	12.244	1.750*	3.842*	1.239	
AMP (%)	0.415	0.122	29.488	2.731**	2.902**	1.446*	
OY (g)	1.838	0.710	38.622	1.618*	1.213*	0.972	
Alpha_pinene (µg·g <sup>−1</sup> )	14.790	4.301	29.078	1.184*	0.649*	0.771*	
Beta_pinene (μg·g <sup>−1</sup> )	10.661	4.491	42.126	1.274*	1.241*	0.929*	
Abietic_acid (µg·g <sup>−1</sup> )	8.183	2.994	36.596	0.887*	1.221*	1.094	
Levopimaric_acid ( $\mu g \cdot g^{-1}$ )	21.631	3.216	14.868	1.341*	2.703*	1.212	

\*\* denotes the significance level of p < 0.01, while \* indicates p < 0.05.

of traits, Crown, AMP, and Beta-pinene exhibited relatively greater phenotypic and genetic variability, while Ht, Pd, and Levopimaric acid showed lower variability. Ht, Pd, and Betapinene demonstrated relatively high family heritabilities of 0.794, 0.855, and 0.695, respectively, indicating significant genetic control. Notably, all three wood properties exhibited relatively high genetic control. The individual heritability of all traits aligned with the family heritability. Particularly, Ht, Pd, and Beta-pinene achieved the highest genetic gain, reaching 11.4%, 11.1%, and 14.8%, respectively, at a 5% selection rate. These findings underscore the exceptional breeding potential of Ht, Pd, and Beta-pinene among all traits in this study.

# **Correlation analysis of various traits**

Further phenotypic and genetic correlation analyses were conducted among the 13 traits (Fig. 2). Positive correlations were observed within growth traits and wood properties, while some negative correlations were noted within resin traits. Notably, Beta-pinene showed the highest phenotypic and genetic correlations with OY ( $r_p = 0.62$ ;  $r_g = 0.67$ ), while Alphapinene exhibited negative phenotypic ( $r_p = -0.03$ ) and genetic ( $r_g = -0.02$ ) correlations with Beta-pinene and Abietic acid. Among wood properties and resin traits, the lowest phenotypic correlation was between MOE and abietic acid ( $r_p = -0.28$ ). Among growth and resin traits, the highest phenotypic correlation was between DBH and Beta-pinene ( $r_p = 0.79$ ), with a higher genetic correlation ( $r_g = 0.45$ ). In addition, negative phenotypic ( $r_p = -0.32$ ) and genetic ( $r_g = -0.09$ ) correlations were observed with DBH and MOE.

#### Estimation of breeding value of Slash pine

The BVs for growth traits are outlined in Supplemental Table S2. Family 8-126 showed the highest BV (0.719) for DBH, whereas family 11-26 had the lowest (BV = -0.821). Elite families for DBH, selected at a 20% rate, included 8-126, 8-131, 4-49, 7-258, 0-53, 2-296, and 0-1339. Similarly, elite families for Ht comprised 0-1027, 0-53, 8-126, 0-1339, 7-258, 10-105, and 11-6. For Huc, selected elite families were 3-1, 2-90, 0-636, 8-126, 7-258, 10-73, and 5-12. For ARW, the top 20% selected families were 2-296, 8-13, 11-6, 8-49, 3-1, 0-636, and 10-73. Supplemental Table S3 presents the BVs for wood properties. For MOE, selected elite families were 2-325, 0-464, 0-53, 0-373, 5-12, 4-9, and 11-26. Selected elite families for AMP included 2-325,

Table 2. Genetic parameter estimation results for various traits.

10-105, 8-126, 0-465, 0-1339, 4-49, and 0-1077. Supplemental Table S4 provides BVs for resin traits. The top 20% selected families for OY were 0-53, 7-77, 3-1, 0-1339, 8-131, 2-325, and 3-1. Alpha-pinene had the highest BV (1.314) in family 7-77 and the lowest BV (–1.226) in family 2-101. Selected families for Alpha-pinene were 7-77, 8-131, 7-258, 0-373, 0-636, 0-510, and 0-1027. Families selected for Beta-pinene matched those for Alpha-pinene.

### Multi-trait combined selection elite families of Slash pine

The results of PCA for growth traits are summarized in Table 3. Following the principle of achieving a cumulative contribution rate of over 85%, the study selected the top three PCs, which accounted for 91.5% and represented the main features of five growth traits. The eigenvalues for these three PCs were 1.689, 1.088, and 0.944, with contributions of 66.4%, 17.3%, and



**Fig. 2** The analysis of phenotypic and genetic correlation within and among various traits. The upper triangular matrix displays phenotypic correlation, while the lower triangular matrix represents the genetic correlation results. AP, BP, AA, and LA represent Alpha-pinene, Beta-pinene, Abietic acid, and Levopimaric acid, respectively.

Traits					$\Delta G$			
	$I_p$	$I_g$	$h_i^2$	$H_i^2$	r = 0.05 i = 2.06	r = 0.10 i = 1.75	r = 0.20 i = 1.40	
DBH	0.205	0.041	0.172 (0.017)	0.593 (0.296)	0.107	0.091	0.073	
Ht	0.155	0.092	0.355 (0.021)	0.794 (0.151)	0.114	0.097	0.077	
Huc	0.240	0.091	0.145 (0.017)	0.543 (0.033)	0.052	0.045	0.036	
Crown	0.268	0.115	0.183 (0.017)	0.610 (0.284)	0.026	0.022	0.017	
ARW	0.194	0.113	0.338 (0.024)	0.781 (0.181)	0.035	0.030	0.024	
Pd	0.099	0.072	0.524 (0.024)	0.885 (0.100)	0.111	0.094	0.075	
MOE	0.121	0.063	0.273 (0.019)	0.870 (0.219)	0.015	0.012	0.010	
AMP	0.297	0.170	0.281 (0.020)	0.874 (0.214)	0.005	0.005	0.004	
OY	0.379	0.150	0.157 (0.023)	0.566 (0.042)	0.017	0.015	0.012	
Alpha_pinene	0.290	0.143	0.244 (0.020)	0.694 (0.234)	0.146	0.124	0.099	
Beta_pinene	0.421	0.202	0.245 (0.020)	0.695 (0.251)	0.148	0.126	0.101	
Abietic_acid	0.366	0.061	0.173 (0.013)	0.285 (0.049)	0.022	0.019	0.015	
Levopimaric_acid	0.147	0.061	0.172 (0.010)	0.592 (0.031)	0.084	0.071	0.057	

r represents the selection rate, and i represents the selection intensity.

Table 3. PCA of growth traits of P. elliottii half-sib families.

Summany	Traite	PCs				
Summary	maits	PC1	PC2	PC3		
Feature vector	DBH	-0.457	0.654	0.000		
	Ht	0.740	0.322	-0.300		
	Huc	0.425	0.695	0.177		
	Crown	-0.769	0.246	0.249		
	ARW	0.400	-0.116	0.872		
Eigenvalue		1.689	1.088	0.944		
Contribution rate (%)		0.664	0.173	0.077		
Cummulative contribution rate (%)		0.664	0.837	0.915		

7.7%, respectively. The weighted sum of individual PC scores, calculated based on the proportion of their eigenvalues to the total selected eigenvalues was utilized to determine the comprehensive scores for growth traits. The comprehensive evaluation model was defined as:  $F_{Total} = 0.454F_1 + 0.292F_2 + 0.254F_3$ . Utilizing this model, comprehensive scores for each family were computed (Fig. 3), ranging from -4.032 to 3.431. The top seven families, identified as elite for growth traits based on a 20% selection rate, were 3-1, 8-126, 2-90, 0-636, 0-1027, 10-73, and 0-53.

For wood properties, PCA results are presented in Table 4. The cumulative contribution rate of the top three principal components reached 100%, representing all features of the three wood properties. Eigenvalues for these PCs were 1.182,

0.984, and 0.833, contributing 39.4%, 32.8%, and 27.8%, respectively. The comprehensive evaluation model for wood properties was defined as:  $F_{Total} = 0.394F_1 + 0.328F_2 + 0.278F_3$ . Comprehensive scores ranged from -4.167 to 2.730 (Fig. 4), with elite families identified as 2-325, 0-1077, 0-373, 0-465, 10-105, 5-12, and 0-510.

The PCA results for resin traits are presented in Table 5, where the cumulative contribution rate of the top three PCs reached 91.5%. Eigenvalues for these PCs were 2.290, 1.291, and 0.993, contributing 45.8%, 25.8%, and 19.9%, respectively. The comprehensive evaluation model for resin traits was defined as:  $F_{Total} = 0.501F_1 + 0.282F_2 + 0.217F_3$ . Comprehensive score for each family ranged from -3.243 to 3.816 (Fig. 5), with elite families identified as 0-1077, 8-131, 7-258, 7-77, 0-1027, 0-510, and 3-1, based on a 20% selection rate.

# Discussion

# Enormous genetic improvement potential in breeding populations of Slash pine

Genetic variations are pivotal, directly influencing genetic gains and guiding breeding strategies in forests<sup>[12]</sup>. The study revealed significant differences among different family lines in traits. Growth traits were notably more influenced by environmental factors, whereas timber traits were primarily shaped by familial effects and resin traits were impacted by both family and environment in comparable proportions. These findings



Fig. 3 PC factor scores of growth traits in *P. elliottii* 33 half-sib families. The color of the squares transitions from blue to red, indicating increasing PC scores.

Table 4. PCA of wood properties of *P. elliottii* half-sib families.

Summany	Traite	PCs			
Summary	TTAILS	PC1	PC2	PC3	
Feature vector	Pd	0.750	0.000	0.659	
	MOE	0.673	0.464	-0.576	
	AMP	-0.409	0.875	0.260	
Eigenvalue		1.182	0.984	0.833	
Contribution rate (%)		0.394	0.328	0.278	
Cummulative contribution rate (%)		0.394	0.722	1.000	

align with previous research on Slash pine<sup>[18]</sup> and Japanese black pine<sup>[52,53]</sup>. Crown, AMP, and Beta-pinene display higher variability in this study when contrasted to Korean pine, whereas DBH and Ht show comparatively lower variations<sup>[54]</sup>.

Additionally, the heritability of growth traits in Slash pine surpassed previous findings<sup>[18]</sup>, while that of wood properties was slightly higher than those related to radiata pine<sup>[55]</sup>, suggesting a stronger genetic influence on wood properties compared to growth traits. Family heritability estimates for monoterpenes were akin to the study of Lei et al.<sup>[56]</sup> but slightly lower than the result of Li et al.<sup>[57]</sup>, likely influenced by locationspecific factors. Moreover, Ht, Pd, and Beta-pinene exhibit higher genetic gains at varying selection rates and intensities, slightly exceeding those reported in previous studies for growth and wood properties<sup>[18,58]</sup>, but demonstrating lower gains for resin traits<sup>[12]</sup>.

# Correlations analysis informs breeding strategy for Slash pine

In practical breeding, considering multiple traits simultaneously is essential for selecting improved genes, yet past studies indicate a trade-off between growth and wood guality<sup>[59]</sup>, underscoring the need for a comprehensive understanding of trait interrelationships to devise rational breeding strategies. This study reveals either non-significant or negative correlations between growth and wood properties, consistent with findings in other tree species like Norway spruce<sup>[60]</sup> and poplar<sup>[61]</sup>, posing a significant challenge to genetic improvement in forestry<sup>[62]</sup>. Additionally, selecting trees at different growth stages may yield varying results<sup>[63]</sup>. Correlative research on Japanese black pine identified significant genetic correlations between growth, morphological, and OY, such as a correlation of 0.73 between DBH and OY<sup>[53]</sup>. Similarly, this study shows comparable results, with a correlation of 0.79 between DBH and Beta-pinene content, and 0.77 between Ht and resin yield, possibly due to vigorous growth promoting resin duct formation<sup>[64,65]</sup>. Furthermore, weak positive or negative correlation between wood properties and resin traits, uncommon in other tree species, were observed. Overall, breeders must meticulously consider trait correlations when selecting for improved genes, and the observed correlations in Slash pine offer valuable insights for effective breeding strategies.



**Fig. 4** PC factor scores of wood quality traits in *P. elliottii* 33 half-sib families. The color of the squares transitions from blue to red, indicating increasing PC scores.

Summany	Traite	PCs			
Summary	ITAILS	PC1	PC2	PC3	
Feature vector	OY	-0.334	0.717	0.494	
	Alpha_pinene	0.934	0.330	0.000	
	Beta_pinene	0.933	0.331	0.000	
	Abietic_acid	-0.660	0.572	0.000	
	Levopimaric_acid	0.000	-0.481	0.856	
Eigenvalue		2.290	1.291	0.993	
Contribution rate (%)		0.458	0.258	0.199	
Cummulative contribution (%)		0.458	0.716	0.915	

# Substantial achievable genetic gains in elite families of Slash pine

For a considerable duration, combined selection of multiple traits has been central in both animal and plant breeding<sup>[66,67]</sup>, with models addressing the impact of LD between traits. PCA, a multivariate selection technique, facilitates the exploration of relationships between explanatory variables and correlated traits<sup>[66]</sup>. In this study, a combined approach using single-trait and PCA-based selection identified elite families for growth, wood properties, and resin traits. Achievable average genetic gains for the three trait categories were 7.4%, 7.2%, and 8.8%, respectively, consistent with a prior study on Slash pine<sup>[18]</sup>. Genetic gains for growth traits exceeded the 10% for Scots pine, while those for wood properties were comparable to its WD related traits<sup>[69]</sup>.

However, genetic gains for resin traits were lower than those reported for Slash pine<sup>[45]</sup>. Quantitative traits are subject to varying degrees of gene-environment interactions, with diverse outcomes influenced by factors such as experimental design, the number of tested families, and different growth periods.

# Conclusions

This study conducted a comprehensive assessment of genetic variation among 33 half-sib families of Slash pine, aiming to identify elite families suitable for industrial use through a multi-trait combined selection approach. Significant differences were noted among families across 13 traits, with growth traits primarily influenced by block × family interaction, wood properties mainly affected by family effects, and resin traits showing variation attributed to both family effects and block × family interaction. Strong genetic control was evident for several traits, notably Beta-pinene, which exhibited the highest variations, and genetic gains, indicating significant breeding potential. Negative correlations were observed between growth and wood properties, while positive correlations were found between growth and resin traits. The multitrait combined selection successfully identified elite families for growth, wood properties, and resin traits. This study provides important references for the long-term breeding strategies of Slash pine, offering rich genetic resources for genomic breeding and molecular breeding.



**Fig. 5** PC factor scores of resin traits in *P. elliottii* 33 half-sib families. The color of the squares transitions from blue to red, indicating increasing PC scores.

# **Author contributions**

The authors confirm contribution to the paper as follows: data curation: Ding X, Zhang Y, Sun J; writing - original draft: Ding X; resources: Luan Q, Jiang J; formal analysis: Ding X; investigation: Tan Z, Huang Q, Diao S, Wu Y; writing - reviewing and editing: Luan Q, Jiang J. All authors reviewed the results and approved the final version of the manuscript.

# **Data availability**

The datasets generated during and/or analyzed during the current study are available from the corresponding author on reasonable request.

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# **Conflict of interest**

The authors declare that they have no conflict of interest.

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# References

- 1. Fengel D, Wegener G. 1984. *Wood: chemistry, ultrastructure, reactions*. Berlin, Germany: Walter de Gruyter. xiii, 613 pp
- Hamilton MG, Freeman JS, Blackburn DP, Downes GM, Pilbeam DJ, et al. 2017. Independent lines of evidence of a genetic relationship between acoustic wave velocity and kraft pulp yield in *Eucalyptus globulus. Annals of Forest Science* 74:17
- 3. Silva JC, Borralho NMG, Araújo JA, Vaillancourt RE, Potts BM. 2009. Genetic parameters for growth, wood density and pulp yield in *Eucalyptus globulus*. *Tree Genetics & Genomes* 5:291–305
- Santos GA, Nunes ACP, Resende MDV, Silva LD, Higa A, et al. 2016. Genetic control and genotype-by-environment interaction of wood weight in *Eucalyptus* clones in the state of Rio Grande do Sul, Brazil. *Revista Árvore* 40:867–76
- Gao S, Wang X, Wiemann MC, Brashaw BK, Ross RJ, et al. 2017. A critical analysis of methods for rapid and nondestructive determination of wood density in standing trees. *Annals of Forest Science* 74:27
- Viana LC, Trugilho PF, Hein PRG, Moreira da Silva JR, Lima JT. 2010. Modelos de calibração e a espectroscopia no infravermelho próximo para predição das propriedades químicas e da densidade básica da madeira de *Eucalyptus. Ciência Florestal* 20:367–76

- Zulak KG, Bohlmann J. 2010. Terpenoid biosynthesis and specialized vascular cells of conifer defense. *Journal of Integrative Plant Biology* 52:86–97
- Kelkar VM, Geils BW, Becker DR, Overby ST, Neary DG. 2006. How to recover more value from small pine trees: essential oils and resins. *Biomass and Bioenergy* 30:316–20

7.

- Neis FA, de Costa F, de Araújo AT Jr, Fett JP, Fett-Neto AG. 2019. Multiple industrial uses of non-wood pine products. *Industrial Crops and Products* 130:248–58
- Luan Q, Tao X, Diao S, Ding X, Jiang J. 2021. Methods, characteristics, variance, and genetics of pine oleoresin components, and their potential for renewable and sustainable energy. *Studies in Natural Products Chemistry* 68:221–53.
- Luan Q, Diao S, Sun H, Ding X, Jiang J. 2022. Prediction and comparisons of turpentine content in Slash pine at different slope positions using near-infrared spectroscopy. *Plants* 11:914
- 12. Ding X, Li Y, Zhang Y, Diao S, Luan Q, et al. 2023. Genetic analysis and elite tree selection of the main resin components of *slash pine*. *Frontiers in Plant Science* 14:1079952
- Diao S, Zhang Y, Luan Q, Ding X, Sun J, et al. 2022. Identification of *TPS-d* subfamily genes and functional characterization of three monoterpene synthases in Slash pine. *Industrial Crops and Products* 188:115609
- 14. Serra S, Fuganti C, Brenna E. 2005. Biocatalytic preparation of natural flavours and fragrances. *Trends in Biotechnology* 23:193–98
- Cunnighan A. 2012. Pine resin: biology, chemistry and applications. Pine Tapping Recent Advances. Bosto: PCA. pp. 1–8
- Peralta-Yahya PP, Zhang F, Del Cardayre SB, Keasling JD. 2012. Microbial engineering for the production of advanced biofuels. *Nature* 488:320–28
- da Silva Rodrigues-Corrêa KC, de Lima JC, Fett-Neto AG. 2013.Oleoresins from pine: production and industrial uses. In *Natural Products*, eds Ramawat K, Mérillon JM. Heidelberg: Springer, Berlin. pp. 4037–60. https://doi.org/10.1007/978-3-642-22144-6\_175
- Lai M, Dong L, Yi M, Sun S, Zhang Y, et al. 2017. Genetic variation, heritability and genotype × environment interactions of resin yield, growth traits and morphologic traits for *Pinus elliottii* at three progeny trials. *Forests* 8:409
- Lai M, Zhang L, Lei L, Liu S, Jia T, et al. 2020. Inheritance of resin yield and main resin components in *Pinus elliottii* Engelm. at three locations in southern China. *Industrial Crops and Products* 144:112065
- Williams CG, Megraw RA. 1994. Juvenile-mature relationships for wood density in *Pinustaeda*. *Canadian Journal of Forest Research* 24:714–22
- 21. Mao T. 2007. Genetic analysis and combined selection of the growth and material of superior tree progeny of Masson pine. Thesis. Nanjing Forestry University, China.
- 22. Fries A. 1986. Volume growth and wood density of plus tree progenies of *Pinus contorta* in two Swedish field trials. *Scandinavian Journal of Forest Research* 1:403–19
- Belonger PJ, McKeand SE, Jett JB. 1996. Genetic and environmental effects on biomass production and wood density in loblolly pine. In *Tree Improvement for Sustainable Tropical Forestry*, Proceedings of the QFRI-IUFRO Conference, Caloundra, Queensland, Australia. pp. 307–10.
- 24. Sun X, Zhang S, Li S, Hou Y. 2005. Combined selection of fine families with multiple traits for pulp wood of *Larch japonica*. *Scientia Silvae Sinicae* 41:48–54
- Wu HX, Powell MB, Yang JL, Ivković M, McRae TA. 2007. Efficiency of early selection for rotation-aged wood quality traits in radiata pine. *Annals of Forest Science* 64:1–9
- Apiolaza L, Chauhan S, Hayes M, Nakada R, Sharma M, et al. 2013. Selection and breeding for wood quality: a new approach. *New Zealand Journal of Forestry* 58:33–37
- 27. O'hehir JF, Nambiar EKS. 2010. Productivity of three successive rotations of *Pinus radiata* plantations in South Australia over a century. *Forest Ecology and Management* 259:1857–69
- Li Y, Luan Q, Shen D, Chen B, Jiang J, et al. 2012. Study on genetic variation of resin components among open-pollinated families of slash pine. *Forest Research* 25:773–79

- 29. Haapanen M. 1996. Impact of family-by-trial interaction on the utility of progeny testing methods for Scots pine. *Silvae Genetica* 45:130–35
- Costa e Silva J, Potts BM, Dutkowski GW. 2006. Genotype by environment interaction for growth of *Eucalyptus globulus* in Australia. *Tree Genetics & Genomes* 2:61–75
- Campbell RK. 1992. Genotype × environment interaction: a case study for Douglas-fir in western Oregon. *Research Paper. Res. Pap. PNW-RP-455*. U.S. Department of Agriculture, Forest Service, Pacific Northwest Research Station, Portland, OR. 21 pp. https://doi.org/10.2737/PNW-RP-455
- 32. Lundströmer J, Karlsson B, Berlin M. 2020. Strategies for deployment of reproductive material under supply limitations - a case study of Norway spruce seed sources in Sweden. *Scandinavian Journal of Forest Research* 35:495–505
- Sixto H, Salvia J, Barrio M, Ciria MP, Cañellas I. 2011. Genetic variation and genotype-environment interactions in short rotation *Populus* plantations in southern Europe. *New Forests* 42:163–77
- Alexandru AM, Mihai G, Stoica E, Curtu AL. 2023. Multi-trait selection and stability in Norway Spruce (*Picea abies*) provenance trials in Romania. *Forests* 14:456
- 35. Yan W, Frégeau-Reid J. 2008. Breeding line selection based on multiple traits. *Crop Science* 48:417–23
- Smith HF. 1936. A discriminant function for plant selection. Annals of Eugenics 7:240–50
- BuenoJ, Vencovsky R. 2000. Efficiency of combined selection over sequential selection in forest tree progeny trials. *Silvae Genetica* 49:169–73
- Dong P, Chang J, Huang L, Li C. 2015. Application of multi-factor comprehensive appraisal in maize breeding. *Agricultural Science & Technology* 16:1614–1616, 1638
- Jia Q, Zhang H, Zhang L. 2016. Variation analysis of hybrid larch families and superior families selection. *Journal of Northeast Forestry University* 44:1–7
- 40. Dong L, Li T, Huang W, Wang B, Xu L, et al. 2021. Screening and comprehensive evaluation of superior strains of Zhejiang safflower camellia oleifera. *Journal of Central South Forestry University* 41:35–45
- Ding X, Diao S, Luan Q, Wu HX, Zhang Y, et al. 2022. A transcriptome-based association study of growth, wood quality, and oleoresin traits in a *slash pine* breeding population. *PLoS Genetics* 18:e1010017
- 42. Ding X, Tao X, Diao S, Luan Q, Jiang J. 2020. Estimation of wood basic density in a *Pinus elliottii* stand using Pilodyn and Resistograph measurements. *Journal of Nanjing Forestry University (Natural Sciences Edition)* 44:142–48
- 43. Tham VTH, Inagaki T, Tsuchikawa S. 2018. A novel combined application of capacitive method and near-infrared spectroscopy for predicting the density and moisture content of solid wood. *Wood Science and Technology* 52:115–29
- 44. Zhang S, Jiang J, Xu Y, Luan Q. 2017. Study on the modulus of elasticity non-destructive evaluation technique of slash pine standing tree. *Forestry Scientific Research* 30:75–80
- 45. Li Y, Jiang J, Luan Q. 2012. Determination and genetic analysis of lipid-producing power, resin density and oleresin content of slash pine. *Journal of Beijing Forestry University* 34:48–51
- 46. Cuevas J, Montesinos-López O, Juliana P, Guzmán C, Pérez-Rodríguez P, et al. 2019. Deep kernel for genomic and near infrared predictions in multi-environment breeding trials. G3 Genes Genomes Genetics 9:2913–24
- 47. R Core Team. 2014. R: a language and environment for statistical computing.
- 48. Vazquez Al, Bates DM, Rosa GJM, Gianola D, Weigel KA. 2013. Technical note: an R package for fitting generalized linear mixed models in animal breeding. *Journal of Animal Science* 88:497–504
- 49. Covarrubias-Pazaran G. 2016. Genome-assisted prediction of quantitative traits using the R package *sommer. PLoS One* 11:e0156744

- Revelle W. 2020. How to: use the psych package for factor analysis and data reduction. Northwestern University, Department of Psychology, Evanston, IL, USA. 95 pp. https://personalityproject.org/r/psych/HowTo/factor.pdf
- 51. Wickham H. 2011. ggplot2. WIREs Computational Statistics 3:180-85
- 52. Yuan C, Zhang Z, Jin G, Zheng Y, Zhou Z, et al. 2021. Genetic parameters and genotype by environment interactions influencing growth and productivity in Masson pine in east and central China. *Forest Ecology and Management* 487:118991
- 53. Liu Q, Zhou Z, Fan H, Liu Y. 2013. Genetic variation and correlation among resin yield, growth, and morphologic traits of *Pinus massoniana*. *Silvae Genetica* 62:38–43
- Jia Q, Liu G, Zhao J, Li K, Sun W. 2022. Variation analyses of growth traits in half-sib families of Korean pine and superior families selection. *Journal of Nanjing Forestry University (Natural Sciences Edition)* 46:109–16
- 55. Baltunis BS, Wu HX, Powell MB. 2007. Inheritance of density, microfibril angle, and modulus of elasticity in juvenile wood of *Pinus radiata* at two locations in Australia. *Canadian Journal of Forest Research* 37:2164–74
- Lei L, Pan X, Zhang L, Ai Q, Li B, et al. 2015. Genetic variation and comprehensive selection of turpentine composition in high-yielding slash pine (*Pinus elliottii*). *Forest Research* 28:804–09
- 57. Li Y, Luan Q, Jiang J. 2012. Study on genetic variation of rosin components in open-pollinated slash pine families. *Forestry Science Research* 25:773–79
- Li Y, Ding X, Jiang J, Luan Q. 2020. Inheritance and correlation analysis of pulpwood properties, wood density, and growth traits of slash pine. *Forests* 11:493
- Lenz PRN, Nadeau S, Mottet MJ, Perron M, Isabel N, et al. 2020. Multi-trait genomic selection for weevil resistance, growth, and wood quality in Norway spruce. *Evolutionary Applications* 13:76–94
- 60. Hannrup B, Cahalan C, Chantre G, Grabner M, Karlsson B, et al. 2004. Genetic parameters of growth and wood quality traits in *Picea abies. Scandinavian Journal of Forest Research* 19:14–29
- Johnson LPV. 1942. Studies on the relation of growth rate to wood quality in *Populus* hybrids. *Canadian Journal of Research* 20:28–40
- 62. Park YS, Weng Y, Mansfield SD. 2012. Genetic effects on wood quality traits of plantation-grown white spruce (*Picea glauca*) and their relationships with growth. *Tree Genetics & Genomes* 8:303–11
- Gaspar MJ, Lousada JL, Rodrigues JC, Aguiar A, Almeida MH. 2009. Does selecting for improved growth affect wood quality of *Pinus pinaster* in Portugal. *Forest Ecology and Management* 258:115–21
- Hood S, Sala A. 2015. Ponderosa pine resin defenses and growth: metrics matter. *Tree Physiology* 35:1223–35
- 65. López-Álvarez Ó, Zas R, Marey-Perez M. 2023. Resin tapping: a review of the main factors modulating pine resin yield. *Industrial Crops and Products* 202:117105
- 66. Hazel LN. 1943. The genetic basis for constructing selection indexes. *Genetics* 28:476–90
- Walsh B, Lynch M. 2018. Evolution and selection of quantitative traits. Oxford: Oxford University Press. https://doi.org/10.1093/ oso/9780198830870.001.0001
- Amaya A, Martínez R, Cerón-Muñoz M. 2021. Selection indexes using principal component analysis for reproductive, beef and milk traits in *Simmental cattle. Tropical Animal Health and Production* 53:378
- 69. Fries A. 2012. Genetic parameters, genetic gain and correlated responses in growth, fibre dimensions and wood density in a Scots pine breeding population. *Annals of Forest Science* 69:783–94

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