

'Honeycrisp': the challenge of the apple crisp revolution

Yosef Al Shoffe^{1*}, Terence Robinson², Gennaro Fazio³, Emily Follett^{1,4}, Matthew Clark⁵, James Luby⁵, David Bedford⁵, Lee Kalcsits⁶ and Gregory Peck¹

¹ Horticulture Section, School of Integrative Plant Science, College of Agriculture and Life Sciences, Cornell University, Ithaca, NY 14853, USA

² Horticulture Section, School of Integrative Plant Sciences, Cornell AgriTech, Geneva, NY 14456, USA

³ USDA-Agricultural Research Service, Plant Genetic Resources Unit, Geneva, NY 14456, USA

⁴ Norwegian Institute of Bioeconomy Research (NIBIO), Ås, Norway

⁵ Department of Horticultural Science, University of Minnesota, St. Paul, MN 55108, USA

⁶ Department of Horticulture, Washington State University, Pullman, WA 99164, USA

* Corresponding author, E-mail: yas24@cornell.edu

Abstract

'Honeycrisp' apples are a crisp cultivar known for their unique texture and flavor. This cultivar is considered revolutionary in the world of crispy apples due to its high value and strong consumer preference. Many new cultivars have recently been developed using 'Honeycrisp' as a parent. However, growing, producing, storing, and marketing 'Honeycrisp' apples present significant challenges. A holistic approach to 'Honeycrisp' production will be discussed, covering aspects such as soil health, rootstocks, orchard management, environmental factors, physiological disorder development, storage protocols, and marketing strategies for sustainable production.

Citation: Al Shoffe Y, Robinson T, Fazio G, Follett E, Clark M, et al. 2025. 'Honeycrisp': the challenge of the apple crisp revolution. *Fruit Research* 5: e039 <https://doi.org/10.48130/frures-0025-0030>

Introduction

'Honeycrisp' apples were developed at the University of Minnesota and released in 1991^[1]. It is a high-value apple cultivar widely planted across the United States apple production areas and worldwide. Its appeal lies in its distinctively crispy texture and sweet flavor, qualities that are highly prized by consumers. Consequently, growers can command a significantly higher wholesale price for 'Honeycrisp' compared to many other apple cultivars. In 2024, 'Honeycrisp' was number four in production in the United States with 9.8% of the total bushels produced behind 'Gala', 'Red Delicious', and 'Granny Smith' (USApple 2024). This is down from the 2023 data, where it was ranked 3rd and estimated at 28 million bushels^[2]. Prices for 'Honeycrisp' were the highest among all cultivars for conventional production (\$2.24/lb) and organic production (\$3.36/lb). For conventionally grown fruit, this is more than \$0.53 higher than the next cultivar and \$0.75 over the median^[3]. According to the USDA-ERS, apples are the highest valued US organic fruit crop. US organic apples sales in 2019 exceeded US\$0.5B and account for nearly 22% of total organic fruit and nut sales, 16% of total apple sales volume, and almost 30% of total apple acreage. On average, organic apples receive a 60% premium over conventional apples. In 2021, there were 2,225 ha of 'Honeycrisp' in the USA, with 95% of the production occurring in Washington State^[4]. Organic 'Honeycrisp' sales in the USA in 2021 accounted for about 32% of the total organic apple sales (US\$165M). Only 'Gala' is produced in greater volume under organic certification in the USA. Producing organically certified 'Honeycrisp' faces many of the same challenges as non-organic production, but is further limited because of the restrictions on the pesticides, plant growth regulators, and fertilizers that can be used.

However, despite its market value, 'Honeycrisp' presents several production challenges that can impact profitability^[5–8].

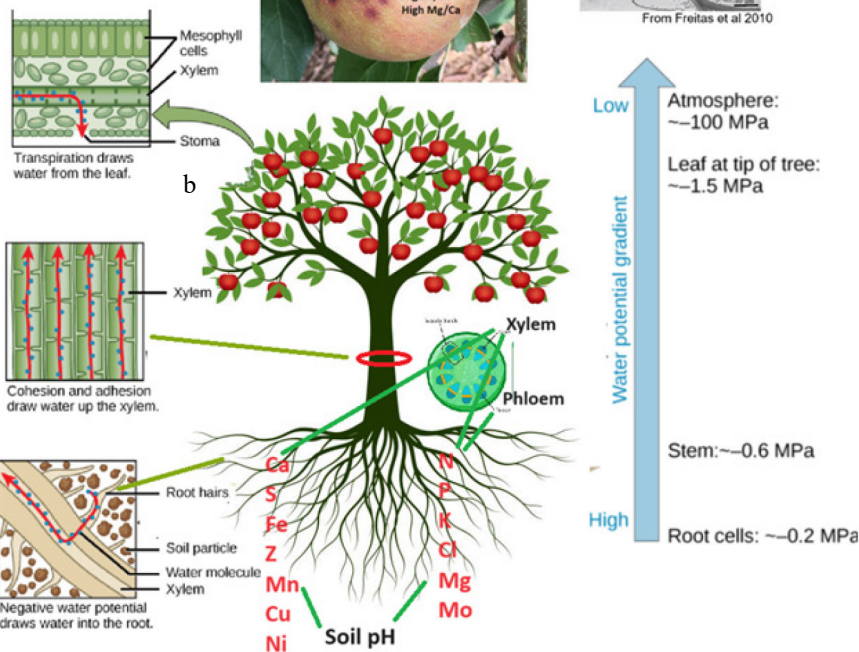
One major issue is its tendency for biennial bearing, which can create cash flow problems for farmers in off-years when the trees

do not flower^[9]. Another significant challenge is its susceptibility to bitter pit, a physiological disorder associated with calcium (Ca) deficiency^[10,11]. Reports from packing houses, industry representatives, and fruit extension agents in New York, Michigan, Pennsylvania, and Washington suggest that growers lose an average of 15%–25% of their 'Honeycrisp' crop to bitter pit annually, with losses rising to 60%–80% in extreme cases^[6].

Rootstock selection is an important step for establishing 'Honeycrisp' orchards. For centuries, the use of rootstocks in apple cultivation was intended to propagate desirable scions through grafting^[12]. Later, the discovery of dwarfing and early-bearing characteristics among certain European apple rootstocks^[13] revolutionized apple growing by enabling the shift from large, inefficient trees to smaller, high-density configurations which greatly boosted orchard productivity^[14]. As the popularity of 'Honeycrisp' rose among North American consumers, researchers and growers sought ways to enhance orchard productivity, disease resilience, and fruit quality for this variety^[15].

'Honeycrisp' is susceptible to a range of physiological disorders during storage and shelf life^[8]. Bitter pit is a physiological disorder related to fruit calcium content that can be seen in the field at harvest but also develops during storage^[16,17]. Different factors affect the development of the disorder, such as growing season, the orchard block, growing region, crop load, rootstock, fruit maturity, and storage temperature^[8,11,18,19]. Cellular partitioning of calcium and cell wall properties have been linked to bitter pit incidence in 'Honeycrisp'^[20]. Intercellular calcium (Fig. 1a) contributes to slower cell wall degradation during the ripening process in apple^[21]. A recent transcriptomic approach identified candidate genes involved in organ abscission and stress response in tissues affected by bitter pit^[22]. Different studies have identified bitter pit QTLs and markers associated with bitter pit^[22,23]. Although bitter pit is related to calcium, excessive nutrients such as potassium or nitrogen can increase bitter pit incidence in fruit. 'Honeycrisp' has higher fruit potassium, magnesium, phosphorus, and nitrogen concentrations

The first report on bitter pit was published in 1840 in Scotland



Modified from: movement of Water and Minerals in the Xylem, LibreTexts libraries

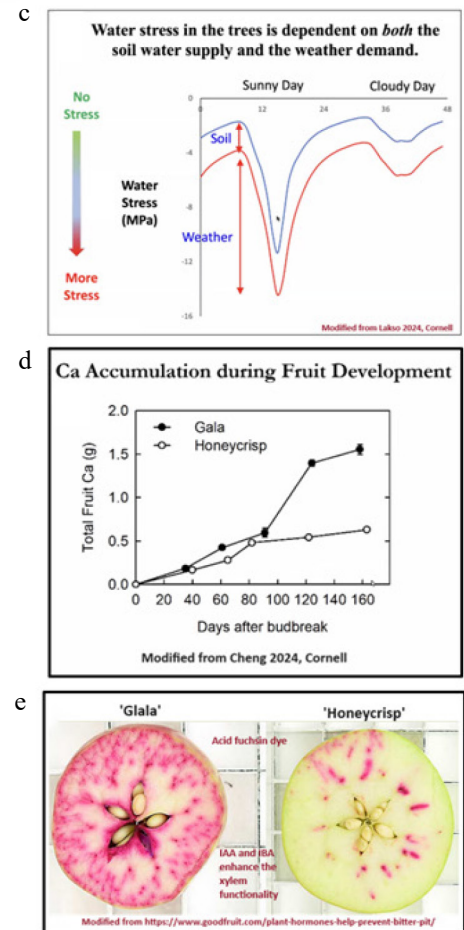


Fig. 1 (a) Bitter pit, calcium deficiency, and calcium in cell membrane modified from de Freitas et al.^[21]. (b) Mineral uptake from soil and movement in tree modified from Raven et al.^[105]. (c) Stress water deficit on sunny and cloudy day. (d) Calcium accumulation in 'Gala' and 'Honeycrisp' apples during fruit development. (e) Xylem functionality in 'Honeycrisp' and 'Gala' apples modified from Griffith and Lake Ontario Fruit Program CU^[106,107].

than other cultivars, which are less susceptible to bitter pit^[24]. Since calcium is not phloem mobile, early xylem dysfunction in developing fruit has been suggested as a primary contributor to low calcium content and nutrient imbalance in developing fruitlets in 'Honeycrisp' apple^[25,26]. However, more research is needed to clarify the underlying physiological reasons behind differences in cellular traits in 'Honeycrisp' compared to other cultivars^[27].

Extensive work has been done to manage pre and post-harvest bitter pit development, starting from soil and foliar mineral nutrient applications in early and late stages of fruit development^[28]. Also, chemical^[29] and non-chemical^[16] prediction models have been developed to manage the disorder at harvest and during storage and help storage operators and marketers make appropriate marketing decisions^[11].

Other disorders of 'Honeycrisp' include soft scald and soggy breakdown, which are low-temperature storage disorders^[8]. The mechanism of soft scald development has been studied in relation to pre- and post-harvest factors^[30–32]. In addition, protocols have been established to manage soft scald development during storage by conditioning the fruit for 1 week at 10 °C before transferring to 3 °C^[33]. Adjustments to storage temperatures can be used to manage warm and chilling temperature disorders that have been investigated^[34,35]. While a few studies have targeted soft scald development^[30,32], the association between bitter pit and soft scald in relation to environment, rootstocks, mineral nutrients, and hormonal balance has not been studied yet. Omics approaches have

been used to understand the etiology of different abiotic stresses, providing potential future gene targets. There have been recent omics approaches to develop predictive tools to improve storage management decisions with the goal of reducing crop losses^[36].

The aim of this review article is to examine the challenges and successes associated with planting 'Honeycrisp' apples over the past 34 years. It also explores the cultivar's role as a parent in the development of many emerging apple varieties and its broader impact on sustainable apple production both in the USA and globally.

Breeding history, current, and offspring of 'Honeycrisp' apples

History

The apple cultivar 'Honeycrisp' has emerged as one of the top-produced and consumer-preferred apples. The apple market (wholesale commodity and direct-to-consumer) can be divided into two eras, with the key event being the release of 'Honeycrisp' in 1991^[1]. The genotype was first selected as a seedling in a family row and identified as MN1711 at the University of Minnesota Horticultural Research Center (Excelsior, MN) from a cross made circa 1960 and planted in 1962. At that time, the recorded parentage was 'Honeygold' x 'Macoun', although it took decades to disprove this notation. Although apple trees are highly heterozygous and seedlings often vary phenotypically from their parents, key indicators have been called into question regarding the recorded

parents^[37]. SSR markers were used to identify 'Keepsake' (mother; 'Frostbite' x 'Northern Spy') as it was extant in the breeding program^[38]. Later, SNP markers and associated haplotypes through pedigree analysis fully elucidated the parents as MN1627 (extinct father) and 'Keepsake'^[39].

A complete history of 'Honeycrisp' acknowledges that this selection was slated for discard and removal due to poor horticultural performance, specifically winter injury. Fortuitously, David Bedford, who had been newly installed to breed apples, recognized the unfavorable site conditions and limited data, which led him to determine that the selection warranted a second chance. After another decade of evaluation, Luby & Bedford^[1] sought and were awarded the plant patent for the variety. Described by Bedford as 'explosively crisp' in a press release in 1991^[40], this distinctive characteristic truly revolutionized consumer expectations for a premium apple eating experience. Per capita apple consumption rose from the early 1990s until 2016, which aligned with the growth and expanded production of 'Honeycrisp' (and other branded varieties). However, consumers continued to demand 'Honeycrisp', as evidenced by the price received in the retail market, where it remained the ceiling through 2024, when pricing collapsed due to oversupply in the apple market. Production for 'Honeycrisp' globally includes Australia, Canada, China, Chile, France, Germany, Italy, New Zealand, and South Africa, with additional territories being explored. In Europe, the fruit is licensed under the trademark Honeycrunch®. 'Honeycrisp', like many red-skinned varieties, has produced bud sport mutations with greater red skin coloration favored by commercial producers who prefer a uniform appearance and by consumers who expect the higher red coloration more typical of its production in Minnesota. These novel sports have continued to create opportunities for 'Honeycrisp'.

Breeding goals at the University of Minnesota, and elsewhere, have focused on crisp texture, as this is a differentiating feature in the market and a hallmark of quality, and more importantly, a heritable trait from 'Honeycrisp'. As such, many offspring have become cultivars or important breeding parents because of their ability to retain crispness alleles with flavor, storage, disease resistance, and appearance traits. Despite concerted research to map a 'Honeycrisp' 'crispness' locus^[41] or to define the phenotype succinctly (sensory evaluation, cell size, cell wall structure and fraction, crispness retention)^[42] or the investigation of related candidate genes^[43,44], no DNA test has been developed for routine screening of seedlings with this characteristic.

Sports and offspring

Apple producers have long desired improved, red-colored sport mutations for many cultivars (e.g., 'Red Delicious' or 'Gala'), as they are appealing to customers and solve harvest, packing, and marketing challenges. Red color is frequently a visual cue for ripeness to customers. Uniform fruit coloration on individual fruit reduces the complexity of product sorting and display. A uniform red color can

lead to homogeneity across distribution territories, which supports brand recognition. In some club varieties, color parameters are dictated in the brand standards. So far, at least seven 'Honeycrisp' sports have been identified and registered, which have demonstrated an improvement in red coloration, such as 'LJ1000' (Royal Red Honeycrisp®), 'BAB 2000' Firestorm™, 'SO 7' ('Roseland Red™'), and 'MinnB42' (Table 1). These color sports are important in areas where environmental conditions can limit anthocyanin accumulation due to warm nights in autumn. One sport has a distinctly earlier harvest date, allowing producers to deliver 'Honeycrisp' to markets sooner, and is marketed as 'DAS-10' (Premier Honeycrisp). All sport mutations are sold under the 'Honeycrisp' price look-up code (PLU 3283) at retail. The sports may be licensed by the intellectual property owner.

A review of the Fruit and Nut Registry^[45–51] identified 35 F1 offspring of 'Honeycrisp' that are in the market (Table 2). This includes targeted, controlled crosses from breeding programs and those using open-pollinated (i.e., 'Honeycrisp') seed. Of course, the University of Minnesota's breeding program has been using 'Honeycrisp' as a parent for the longest (with records confirming its use beginning in 1986). They have advanced to the third generation of seedlings and are using complex pseudo backcross/inbreeding strategies to incorporate 'Honeycrisp' alleles for texture and storage attributes. Notable varieties which have been selected as seedlings include 'CN121' (SugarBee®), 'MAIA1' (EverCrisp®), 'Minneiska' (SweeTango®), 'NY1' (SnapDragon®), and 'WA38' (Cosmic Crisp®).

European offspring

The success of 'Honeycrisp' in the USA has spread internationally, including to Europe, where it has been grown and sold as HoneyCrunch® since the early 2000s. As in the USA, 'Honeycrisp' has become frequently used in breeding programs throughout Europe to develop cultivars with crunchy, juicy flesh and unique tastes for various markets. In addition, many 'Honeycrisp' offspring bred in the USA are now being grown in Europe as well, such as 'Minnieska' (SweeTango®), 'NY1' (SnapDragon®), and 'WA-38' (Cosmic Crisp®), under various licensing programs. During the 2023 season, 62.3% of the total apple trees planted were 'Honeycrisp' offspring in a region of Northern Germany, Niederelbe, up from 52% the year before^[52]. Many of these cultivars have been rapidly adopted by industry and introduced as club varieties with significant marketing behind them, including cultivar-specific websites translated into multiple languages. However, research appears to be limited, or very location-specific, with most literature found in regional industry journals and focusing on pre-harvest factors. As such, published scientific literature on postharvest characteristics is lacking for most of these cultivars.

From gatherable knowledge, 'Wurtwinning' (Bloss®, 'Honeycrisp' x 'SQ 159'/MagicStar®/Natyra®) has been planted in Switzerland and green spots have been observed on the fruit. Another new cultivar, 'GS66' (Fraülein®, parents are unknown: suspected to be

Table 1. 'Honeycrisp' sports.

Variety (trademark)	Source	Date	Origin
'Cameron Select'®	Various	2001	WA/USA
'Walden'	Whole tree mutation of Honeycrisp	2005	NY/USA
'LJ-1000' (Royal Red Honeycrisp®)	Whole tree mutation of Honeycrisp	2011	WA/USA
'BAB2000' (Firestorm®)	Whole tree mutation of Honeycrisp	2014	WA/USA
'DAS-10' (Premier Honeycrisp®)	Whole tree mutation of Honeycrisp	2014	PA/USA
'Lewis'	Limb mutation of Honeycrisp	2015	WA/USA
'MinnB42'	Limb mutation of Honeycrisp	2016	MN/USA
'SO 7' (Roseland Red™)	Whole tree mutation of Honeycrisp	2021	VA/USA

Table 2. 'Honeycrisp' progeny. Modified from the Fruit and Nut registry^[45–51].

Variety (trademark)	Parentage	Date	Origin
'Minneiska' (SweetTango®)	'Honeycrisp' × 'Minnewashta'	2008	MN/USA
'New York 1' (SnapDragon®)	'Honeycrisp' × 'NY752'	2011	NY/USA
'WA38' (Cosmic Crisp®)	'Enterprise' × 'Honeycrisp'	2012	WA/USA
'DS 3' (Pazazz®)	'Honeycrisp' O.P.	2013	WI/USA
'DS 22' (Riverbelle®)	'Honeycrisp' O.P.	2013	WI/USA
'CN 121' (SugarBee®)	'Honeycrisp' O.P.	2013	MN/USA
'CN B60'	'Honeycrisp' O.P.	2013	MN/USA
'Regal 13-82' (Juici®)	'Braeburn' × 'Honeycrisp'	2014	WA/USA
'MAIA1' (Evercrisp®)	'Honeycrisp' × 'Fuji'	2014	IN/USA
'DS-41'	'Honeycrisp' O.P.	2014	WI/USA
'CN B110'	'Honeycrisp' O.P.	2014	MN/USA
'MN55' (Rave®)	'Honeycrisp' × 'MonArk'	2016	MN/USA
'R10-45'	'Honeycrisp' × 'Cripps Pink'	2017	WA/USA
'Orléans'	'Empire' × 'Honeycrisp'	2017	Quebec/Canada
'MAIA12' (Summerset®)	'Honeycrisp' × 'Fuji'	2018	OH/USA
'MAIA11' (Rosalee®)	'Honeycrisp' × 'Fuji'	2018	OH/USA
'MAIA7' (Crunch-A-Bunch®)	'Honeycrisp' O.P.	2018	OH/USA
'Howell TC5 WF'	'Honeycrisp' O.P.	2018	WA/USA
'Howell TC4 WF'	'Cripps Pink' × 'Honeycrisp'	2018	WA/USA
'NY56' (Cordera®)	'Honeycrisp' × 'NY65707-19'	2019	NY/USA
'MAIA-T' (Scruffy®)	'Honeycrisp' O.P.	2019	OH/USA
'MAIA-L' (Ludacrisp®)	'Honeycrisp' O.P.	2019	OH/USA
'Honeysuckle Rose #1-6'	'Honeycrisp' × 'Simmons Gala'	2019	IL/USA
'NY73' (Pink Luster®).	'Imperial Gala' × 'Honeycrisp'	2020	NY/USA
'Regal D5-100' (Karma®).	'Huaguan' × 'Honeycrisp'	2020	WA/USA
'Howell TC7' (Lucy™Gem)	'Airlie Red Flesh' × 'Honeycrisp'	2020	WA/USA
'Regal D17-121'	'Honeycrisp' × Co-op 39	2021	WA/USA
'DS 102'	'Honeycrisp' O.P.	2021	WI/USA
'PremA003' (Posh®)	'Honeycrisp' × 'Sciros'	2021	New Zealand
'MN80' (Triumph®).	'Honeycrisp' × 'Liberty'	2021	MN/USA
'R204' (Kissabel®)	'Honeycrisp' × CR35-1	2022	France
'MAIA-SM' (Sweet Maia®).	'Honeycrisp' × 'Co-op 31'	2022	OH/USA
'D27-16' (Aura®)	'8S6923' × 'Honeycrisp'	2022	WA/USA
'Wurtwinning' (Bloss®)	'Honeycrisp' × 'SQ 159'	2022	Netherlands
'WA 64' (Sunflare™)	'Honeycrisp' × 'Cripps Pink'	2023	WA/USA

O.P. refers to open-pollinated.

'Honeycrisp' × 'Braeburn'), is susceptible to bitter pit and another unique skin pitting disorder named GS66-spot when grown in northern Germany, but initial storage experiments suggested otherwise good storability^[53].

Although many new 'Honeycrisp' offspring in Europe show promising storability, the available observations remain preliminary, despite substantial plantings and industry investment. In some cases, physiological disorders and challenging storage have only been discovered after industry investment has begun in earnest. In Norway, production of two cultivars, 'Wursixo' (Eden®) and 'Wuranda' (Fryd®) with 'SQ-159'/MagicStar®/Natyra® × 'Honeycrisp' parentage has started and rapidly increased in recent years, with commercial planting occurring before thorough postharvest studies were conducted^[54].

Honeycrisp rootstock interactions

Rootstocks are considered an important factor in controlling vegetative growth, fruit development, and fruit mineral nutrient content^[7,55,56].

Early evaluations suggested that apple rootstocks might play a more significant role in fruit quality and productivity than initially

believed. Certain rootstocks, such as Geneva 935 (G.935) and G.969, demonstrated the ability to produce balanced, productive trees with fewer apples affected by bitter pit^[57]. Building on this early data, a multi-state field trial featuring 'Honeycrisp' as the scion was conducted using a diverse set of rootstocks. This trial aimed to identify rootstocks that could mitigate 'Honeycrisp's' tendency for weak growth while maintaining an optimal crop load, an essential factor for fruit quality surpassing nitrogen's role^[58].

The trials revealed that diverse rootstocks significantly influence leaf and fruit nutrient concentrations, particularly critical ratios like potassium-calcium (K/Ca), magnesium-calcium (Mg/Ca), and nitrogen-calcium (N/Ca), which are closely tied to bitter pit development. This nutrient effect, observed during the establishment phase^[59], was confirmed in subsequent multi-year, multi-location studies^[60–62]. Inherently low calcium transport capacity in 'Honeycrisp' amplifies rootstocks' impact, emphasizing the importance of nutrient uptake and transport as key objectives in breeding programs^[60,63].

Furthermore, rootstocks affect soil pH interactions, phytohormone levels, stress tolerance, tree architecture, and graft union strength. For example, G.41 produces weaker unions while G.214 forms stronger ones^[64]. Overall, rootstock choice profoundly

influences productivity, disease resilience, fruit quality, and orchard profitability. Options include G.214, G.935, G.969, B.10, and G.890 optimal for vigor, productivity, and nutrient balance.

Donahue et al.^[65] suggested that bitter pit performance of a rootstock should be a major consideration when choosing a rootstock for a new 'Honeycrisp' orchard. Islam et al.^[20] reported that B.10 rootstock had lower bitter pit incidence at harvest and during storage compared with other rootstocks, and was associated with higher Ca and lower Mg concentrations as well as the water-insoluble pectin fractions in 'Honeycrisp' fruit compared with those from trees grafted on G.41 and V.6 (Vineland series) rootstocks. Others have found that rootstocks have a significant impact on the Ca, K, and K/Ca ratio in the scion tissues, and the K/Ca in the fruit was tightly correlated with bitter pit incidence^[66].

Previous research^[66–69] investigated the influence of rootstocks on the mineral nutrient profile and bitter pit incidence in 'Honeycrisp' apples. Certain rootstocks result in higher potassium (K) and nitrogen (N) levels in fruit, while the ratio of K/Ca or N/Ca is elevated in some rootstocks. Rootstocks vary in their efficiency in taking up K and N, which influences fruit quality and bitter pit occurrence, with some rootstocks leading to higher incidences of bitter pit in the fruit (Fig. 2)^[6,70]. Four long-term field trials conducted in Geneva, NY, measured total yield, bitter pit incidence, and crop value across various rootstocks. Rootstocks such as B.9 showed lower K uptake and had lower bitter pit incidence but lower yields. Rootstocks like G.41 and G.11 exhibited higher K uptake and greater vigor, resulting in higher yields but more bitter pit in certain years^[71]. G.214, with similar vigor to G.41 but lower K uptake, resulted in low bitter pit risk. Results showed that rootstocks like B.9, B.10, G.969, and G.214 consistently had lower bitter pit incidences. A negative correlation was observed between calcium concentration and bitter pit, while a positive correlation existed between K/Ca ratio and bitter pit. Rootstocks that consistently had low bitter pit also had higher calcium levels and/or lower K/Ca ratios^[72]. The cause of differences in K and N uptake efficiency remains unclear but is likely related to rootstock vigor and root system size. In selecting a rootstock for 'Honeycrisp', the incidence of bitter pit is only part of the equation; total cumulative yield and crop value must also be considered^[55,73]. Robinson et al.^[6] found that B.9 had low bitter pit incidence but also lower cumulative yield, making other rootstocks more economically viable. Matching rootstocks to scion variety in a 'designer rootstock' approach can help optimize fruit size and reduce bitter pit incidence. Rootstocks with high K uptake, like G.11 and G.41, are ideal for small-fruit cultivars such as 'Gala', while rootstocks with low K

uptake, like G.214 and G.969, are better for bitter pit-sensitive cultivars like 'Honeycrisp'^[6].

'Honeycrisp' is known to have low vigor, which, when grown in organic systems, requires additional attention. If 'Honeycrisp' is grown on a dwarfing rootstock under organic production, poor weed control can lead to reduced growth and low crop yields^[74]. Several Geneva® series rootstocks, especially G.890, have been shown to have greater productivity under organic management, and should be used for organic 'Honeycrisp' plantings^[75].

Orchard management of 'Honeycrisp'

Soil preparation

Soil management challenges have been the subject of extensive research over many years, with the findings meticulously documented and shared^[76,77]. Soil acidity, with pH below 5.5, impacts 'Honeycrisp' apple productivity, resulting from precipitation-driven leaching, replacing essential cations like potassium and calcium with hydrogen and aluminum, especially in regions where rainfall exceeds evapotranspiration^[78]. Preplant soil preparation for 'Honeycrisp' apple orchards focuses on soil pH^[69,79], calcium (Ca) content, potassium (K) content, and organic matter. Optimal soil targets are a pH of 7.2, 6,000 kg/ha of Ca, 300 kg/ha of K, and 2%–3% organic matter^[6]. Soils high in K or organic matter, > 500 kg/ha K and > 4% organic matter, are difficult to manage for low bitter pit incidence^[6]. In the Western US, soils are often high in K, while some Eastern US soils are too rich in organic matter, creating challenges for successful 'Honeycrisp' growth. Differences in solubility and mobility of soil potassium and their availability to plants are explained^[80]. To address low Ca and pH in Eastern US soils, intensive liming before planting is recommended, followed by bi-annual lime applications post-planting. Honeycrisp requires higher Ca levels than other cultivars, with at least 6,000 kg/ha of Ca in the top 30 cm. Soil pH should be raised to 7.2, a level shown to promote low bitter pit incidence and strong tree growth^[69]. For soil amendments, calcitic lime (CaCO₃) is preferred over dolomitic lime or gypsum, as dolomitic lime adds magnesium^[81], which can increase bitter pit, while gypsum is a source of less soluble Ca^[6].

Annual fertilization standards

Current annual fertilization standards of 'Honeycrisp' in NY State are based on leaf nutrient values^[82]. Leaf samples should be collected earlier than for other varieties, as 'Honeycrisp' stops shoot growth earlier, affecting leaf nutrient concentrations^[83]. The recommended leaf N concentration is 2.0% of dry weight, similar to

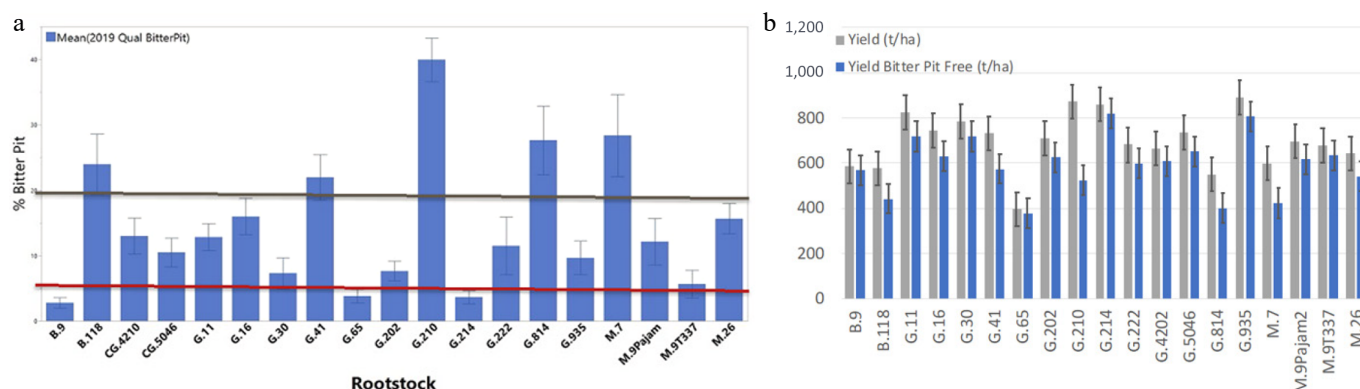


Fig. 2 (a) Bitter pit incidence of various rootstocks on mature 'Honeycrisp' trees at Geneva, NY, in 2019. B.9, G.65, and G.214 had low bitter pit incidence (< 5%) while B.118, G.41, G.210, G.814, and M.7 had high bitter pit incidence (> 20%). (b) Total yield and yield of bitter pit-free fruit of 'Honeycrisp' on 19 rootstocks over 16 years at Geneva, NY, USA. Modified from Robinson et al.^[6].

'McIntosh'. For blocks with less than 2.0% N, apply 40–50 kg of N/ha, while blocks with 2.0%–2.25% should receive 20 kg/ha. No N should be applied if leaf N exceeds 2.25%^[6]. Fertilization should occur early in the growing season, with split applications at bud break and petal fall^[84]. Post-harvest foliar N can be applied for low vigor trees, particularly for those with leaf N under 2%. Potassium fertilization for 'Honeycrisp' is more cautious than for other varieties due to its tendency to develop bitter pit at high K levels. Cheng and Miranda Sazo^[82] found that 'Honeycrisp' has a lower K requirement than 'Gala'. Thus, the optimal leaf K range for 'Honeycrisp' is 1.0%–1.1%, lower than varieties like 'Gala' or 'Empire'. If leaf K is below 1.0%, apply 60–80 kg K₂O/ha; for levels between 1.0% and 1.3%, apply 30–50 kg K₂O/ha. No K should be applied if leaf K exceeds 1.3%^[6].

Calcium is managed through preplant calcitic lime applications and bi-annual lime applications post-planting. 'Honeycrisp' requires higher leaf Ca (1.5%–2.0%) than other cultivars, so soil and leaf Ca levels should be monitored regularly. Fruit peel sap analysis in early July is also recommended to assess nutrient balance, particularly the K/Ca ratio. If the K/Ca ratio exceeds 25, growers should reduce K applications and increase foliar Ca sprays. If the ratio is below 25, the recommendation is to apply 40–50 kg K₂O/ha for optimal fruit size and quality^[6].

Crop load management

Crop load is crucial for determining yield, fruit size, and the incidence of bitter pit in 'Honeycrisp' apples^[85]. It is also linked to biennial bearing, making its management critical for consistent annual production^[86]. Low crop load can increase bitter pit, while high crop load often leads to biennial bearing. During an 'on-year', when there is often a heavy crop load, without thinning, fruit size is lower, ripening is delayed, and fruit color development is poor^[62]. Flower bud formation occurs during the previous growing season^[87]. When fruit is thinned early during fruit development, it has a substantial effect on competing fruits and shoots. Thus, achieving an optimal crop load each year is essential for minimizing bitter pit and ensuring annual yields^[88]. Crop load is managed through pruning, chemical thinning, and hand thinning^[89].

Precision pruning is the first step in crop load management^[90]. Research indicates that when flower bud load is too high, chemical thinning alone may not reduce crop load sufficiently^[91]. Pruning should focus on removing enough flower buds to allow chemical thinning to achieve the target fruit number. It is suggested to count floral buds and leave around 80% more buds than the target as insurance against frost or poor set^[92].

Precision chemical thinning is essential for preventing biennial bearing and ensuring fruit quality^[88]. Early thinning at bloom and petal fall is critical because fertilized flowers produce gibberellin (GA), which inhibits flower bud formation for the next year^[93]. However, Elsasy & Hirst^[94] show that seeds do not play a direct role in regulating flower formation. By thinning blossoms and fruitlets early, seed number and GA concentration are reduced, promoting flower formation for the next season. Two bloom sprays of ammonium thiosulfate or lime sulfur plus oil, followed by naphthaleneacetic acid (NAA) + carbaryl spray at petal fall, help control crop load. If pruning has been effective, this thinning strategy should yield a crop close to the target, requiring minimal hand thinning. Overcropped trees can suffer from poor fruit quality, including reduced color, delayed maturity, and decreased fruit dry matter^[58,88]. If biennial bearing occurs due to improper thinning or frost, strategies like deficit irrigation and delayed hand thinning can help manage oversized fruit.

The biennial fruit-bearing nature of 'Honeycrisp' is particularly difficult for organic producers because the main plant bioregulators

used for fruitlet thinning are unavailable due to being synthetically manufactured^[95]. This leaves growers with fewer sprayable options, most of which rely on caustic materials to either disrupt fertilization during flowering or cause a transient but severe phytotoxic stress on the entire tree that results in fruitlet abortion^[96]. Liquid lime sulfur (calcium polysulfide) is the most common material used for thinning under organic production, but it can result in fruit peel russetting that devalues the fruit in wholesale markets^[97]. Other caustic materials, such as salts, biofungicides, and solvents, have been trialed with limited success^[98,99]. A gibberellin product (ProVide; Valent BioSciences, LLC) has been formulated for use in organic production. Some research has shown that gibberellins can be applied in the off-year of a biennial bearing cycle to encourage flower bud formation and minimize low yields during the off-year^[100]. However, a heavy crop load will likely overwhelm the effects of exogenously applied gibberellins, so it must be done in coordination with other crop load management approaches^[101].

Return bloom management

To overcome biennial flowering, growers have traditionally applied four summer sprays of either 10 µL/L NAA or 473 mL Ethephon (Ethrel®), typically applied starting on June 21 (under NY conditions), when fruits are 35 mm in diameter. While this program has worked well for other biennial varieties, it has proven less reliable for 'Honeycrisp'^[102]. Recent research suggests two key reasons for this inconsistency:

1. Timing of flower initiation: Flower initiation for 'Honeycrisp' occurs much earlier than other cultivars, beginning around 30 d after bloom, with peak initiation between 45 and 55 d. The traditional spray timing often misses this critical period. Thus, initiating the summer sprays earlier, closer to the flower initiation period in mid to late June, could improve consistency (Francescato et al., unpublished data)^[9].

2. Gibberellin (GA) load: When the number of fruit and seeds is high, the resulting high gibberellin production can inhibit flower initiation^[103], counteracting the flower-promoting effects of NAA or Ethrel®. In years with fewer fruit or seeds per fruit, the sprays are more effective, as the GA load is lower.

To address these issues, researchers recommend precision pruning to limit the initial flower bud load to no more than 1.8 times the target fruit number. This helps avoid excessive GA levels and supports flower bud initiation. Additionally, bloom thinning and petal fall thinning should be done early to reduce fruit numbers, ensuring that flower initiation is not hindered by high GA levels. In recent trials, starting Ethrel® sprays earlier, at the 16 mm fruit size stage, has shown better results for return bloom than the traditional timing of starting the sprays at 35 mm fruit size. Four sprays at 10-d intervals, starting at 150 µL/L and escalating to 300 µL/L, have been effective in promoting bloom in the 'on' year. However, this approach requires careful control of fruit numbers and seed load to avoid excessive GA. Ethrel® should only be applied when temperatures are below 26 °C to minimize the risk of unintentional thinning^[6].

Irrigation management to manage growth and nutrition partitioning

The uptake of calcium (Ca), potassium (K), and magnesium (Mg) by apple trees is influenced by soil moisture levels. In dry soils, the uptake of these cations is limited, leading to deficiencies, such as K leaf deficiency in August, even if soil K levels are adequate. Conversely, excessively wet, waterlogged soils hinder root function and nutrient uptake. K can be transported through both the xylem and phloem, while Ca is only transported in the xylem, requiring water movement into the fruit to prevent issues like bitter pit

(Fig. 1b). Early in the season, fruitlets transpire through stomates, facilitating greater water and Ca movement. However, as stomata become non-functional later in the season, this transport is reduced. Research indicates that increased water supply during the season can improve fruit size and red color but negatively affect fruit firmness, soluble solids, and increase bitter pit and rot incidence^[104]. Bitter pit is particularly influenced by water stress during early season (near petal fall) and mid-season (early July) (Fig. 1c), with more stress leading to higher bitter pit after storage. The condition is worsened by low crop loads. Water deficits from July 3–17 result in high bitter pit incidence, while adequate water supply and higher crop loads reduce it. Excessive water, on the other hand, decreases root function and Ca uptake, worsening bitter pit.

Deficit irrigation (DI) can be used as a tool to limit fruit growth and control bitter pit in 'Honeycrisp'^[108]. Deficit irrigation imposes predetermined periods of soil or plant water deficit to improve fruit quality, decrease vigor, or conserve water^[109]. With 'Honeycrisp', reduced water supply in the last half of the season helps reduce K uptake and reduces bitter pit incidence. Dry climates with little to no rainfall are more successful at maintaining good DI because growing environments with higher rainfall lose their ability to control water supply.

To prevent bitter pit in 'Honeycrisp', irrigation should begin early in the season (May–June) and continue through July to ensure adequate Ca uptake, and only through June in hotter, irrigated environments in the Western US. However, irrigation should be suspended after August 1 to prevent excessive K uptake, which worsens the K/Ca ratio and leads to bitter pit. Under conditions that will promote large fruit and in irrigated, hot environments, deficit irrigation should start earlier, closer to July 1^[108]. Deficit irrigation during fruit cell expansion in the later season can reduce bitter pit incidence, although conditions such as rainfall remain uncontrollable. Proper early nutrient uptake is essential for reducing bitter pit. Deficit irrigation is a valuable tool to use when crop loads are low or when bitter pit risk is high in orchards with higher vigor.

Foliar Ca sprays to manage bitter pit

Foliar calcium (Ca) sprays can slightly increase fruit Ca levels, typically by around 10%, but this is usually insufficient to fully address Ca deficiencies, since calcium is immobile in the phloem. The effectiveness of calcium sprays relies on fruit contact to increase overall fruit calcium content^[110]. When proper soil Ca levels and adequate irrigation are maintained, foliar sprays can significantly reduce bitter pit. The most effective Ca sprays are applied during the second half of the season, after xylem dysfunction in the fruit limits Ca movement into the fruit (Fig. 1e). There are many different commercial calcium sprays. More commonly, calcium can be sprayed as calcium nitrate ($\text{Ca}[\text{NO}_3]_2$)^[111], calcium carbonate (CaCO_3)^[112], or calcium chloride (CaCl_2)^[28,113]. Calcium chloride is the most widely used calcium spray because it is the least expensive and most effective spray available^[114,115]. Although calcium sprays have been shown to be effective, the amount of calcium absorbed and the effect of weather or development on the absorption efficiency are less understood. Kalcits et al.^[110] reported that calcium spray adhesion was greater during early development for 'Honeycrisp'. The decrease in spray adhesion and increase in fruit surface area as fruit development progressed provided the conditions for a consistent amount of calcium spray to adhere to each fruit throughout the growing season. Their results demonstrated that frequent calcium sprays are required to increase fruit calcium concentrations. The current recommendations for 'Honeycrisp' include eight to ten cover sprays of calcium chloride (CaCl_2), compared to five to six sprays for other bitter pit-prone varieties. Various Ca products, such

as STOPIT[™] (calcium chloride), Sysstem©-CAL (calcium phosphite), Cell Power© Calcium Platinum (calcium nitrate), and Ele-Max© Calcium FL (calcium carbonate), were tested over three years^[6]. STOPIT[™] showed the best results in reducing bitter pit, both at harvest and after storage, outperforming other products. Based on these trials and other studies, CaCl_2 is considered the most effective spray form^[28,116]. A total of 10–13 kg/ha of Ca per season is recommended in high-density orchards; this may also vary depending on soil composition and leaf mineral analysis. However, CaCl_2 can cause fruit damage if sprayed too early (before mid-June), so it should only be applied once fruit reaches 30 mm in size. Sysstem©-CAL, on the other hand, has been found safe for early-season sprays without causing fruit injury.

Prediction methods of bitter pit in 'Honeycrisp' apples

The link between low calcium content in fruit and an increased risk of bitter pit has long been established, and foliar applications of calcium products have been shown to reduce the development of bitter pit in susceptible cultivars (Fig. 1d)^[117], including 'Honeycrisp'^[28]. These relationships have also led to the development of prediction models for various cultivars. While some models focus solely on calcium, most incorporate other minerals, such as magnesium, potassium, and nitrogen, into their predictions^[118,119]. Bitter pit incidence is related to high ratios of Mg/Ca in 'Fuji' apples^[120], Mg + K + N/Ca in 'Honeycrisp' apples^[16], Mg + K/Ca in 'Cox Orange Pippin'^[121] and 'Granny Smith' apples^[21], and Mg/Ca and K/Ca in 'Catarina' apples^[122]. However, regression coefficients vary across studies, ranging from as low as 0.4 to as high as 0.9^[16,123,124]. Furthermore, it has been suggested that hormone concentrations and their interactions play a pivotal role in regulating xylem dysfunction, which is closely associated with the nutrient imbalances commonly observed in bitter pit^[26].

In addition to mineral-based methods, several non-mineral techniques for predicting bitter pit have been developed. These include treatments with magnesium salts, such as vacuum infiltration with MgCl_2 in 'Braeburn' apples, which predicts bitter pit incidence after storage^[125]. Ethylene treatments accelerate ripening through methods such as ethephon dipping or acetylene gas fumigation^[126,127]. A passive method, where fruit is stored at warm temperatures to allow bitter pit development before harvest, has also been shown to reliably predict bitter pit in 'Golden Smoothee' apple^[124].

A variety of methods have been developed to predict and manage pre- and post-harvest bitter pit in 'Honeycrisp' apples, including:

Shoot length and N/Ca ratios

The method relies on correlating the average of current season terminal shoot length, with moderate branch angles, measured using five terminal shoots per tree from 20 trees per orchard after terminal bud set, with the fruit peel N/Ca ratio taken 3 weeks before the anticipated harvest from the same 20 trees (60 apples per orchard in total) to predict bitter pit development during cold storage. The results show that bitter pit development consistently increases with higher N/Ca ratios and longer shoot lengths^[16]. However, in a different study, the model underestimated the incidence of bitter pit^[19].

Peel sap analysis method

Assessing bitter pit risk early in the growing season is crucial for timely mitigation, which reduces the fruit disorder at harvest and during storage. Current methods predict bitter pit risk shortly before harvest, limiting the ability to implement preventive measures. The

peel sap analysis method measures K/Ca and N/Ca ratios from fruit-let peel sap. This method, conducted around 2 months after bloom, involves freezing the fruit peel at -80°C to lyse the cells and then squeezing it to extract the juice. It is quicker, simpler, and more environmentally friendly compared to others. The K/Ca ratio helps categorize orchards into low, medium, or high-risk groups, while the N/Ca ratio provides similar categorizations. These results are available by mid-July, allowing growers to take action to reduce bitter pit risk. Mitigation strategies include increasing calcium sprays, avoiding certain growth regulators, reducing nitrogen and potassium applications, and increasing soil pH with lime^[6]. This early prediction system offers a valuable tool for effectively managing bitter pit risk.

Passive method

The passive method, which involves collecting 100 fruits 3 weeks before the anticipated harvest and storing them at room temperature, has consistently produced reliable results across different orchards, regions, and successive years under New York and Pennsylvania conditions. Its simplicity makes it a powerful and dependable tool that growers and storage operators can easily use to predict the occurrence of bitter pit in 'Honeycrisp' apples during storage^[11].

Pre- and post-harvest application of PGRs to control fruit drop and postharvest fruit quality

Sprayable formulas of 1-methylcyclopropene (1-MCP) commercialized as Harvista™ and aminoethoxyvinylglycine (AVG) commercialized as ReTain® are plant growth regulators (PGRs) that inhibit ethylene action or biosynthesis to delay fruit drop and maturity. While effective for harvest management, these PGRs can worsen bitter pit development in susceptible orchards. Research shows that applying 1-MCP before harvest increases bitter pit, leather blotch, core browning, and CO_2 injury. However, preharvest 1-MCP reduces senescent breakdown and wrinkled skin, signs of advanced maturity, and helps prevent soft scald during storage at 0.5°C ^[18]. For fruit with a high K/Ca ratio from the peel sap analysis method, it is recommended to avoid using preharvest AVG or 1-MCP to prevent aggravating bitter pit^[6].

Postharvest application of 1-MCP at the commercial rate can sometimes reduce greasiness. Still, it significantly increases the risk of leather blotch in fruit susceptible to bitter pit and that has also been treated with preharvest 1-MCP. Although the leather blotch is not fully understood, the effects of 1-MCP suggest that it may be linked to inhibited fruit ripening. Additionally, postharvest 1-MCP treatment has been shown to increase the incidence of internal carbon dioxide (CO_2) injury in treated fruit. However, 1-MCP has no effect on fruit firmness in 'Honeycrisp'. Various studies have shown that changes in fruit firmness in 'Honeycrisp' apples are minimal, likely due to low polygalacturonase activity^[128] associated with high turgor and cell wall integrity^[27].

Maturity indices in 'Honeycrisp' apples for short- and long-term storage

Color is a key factor influencing the harvest timing of 'Honeycrisp' apples, and anthocyanin accumulation in fruit peel follows the blush and stripe patterns^[129]. Spot picking is performed two to four times during the harvest window, depending on the orchard, regional conditions, and preharvest treatments. Internal ethylene concentrations (IEC), starch indices (measured on a 1–8 scale), firmness, and soluble solids content (SSC) did not exhibit consistent changes over the 3-week harvest period. Additionally, 'Honeycrisp' apples

are susceptible to stem punctures, with incidences reaching up to 18.5%. This susceptibility is unaffected by the timing of the harvest^[130]. Harvest time has a strong impact on fruit susceptibility to physiological disorder development during storage^[130]. Early harvest of 'Honeycrisp' apples may lead to the development of bitter pit, superficial scald, core browning, and internal CO_2 injury^[34,130,131]. Conversely, late harvest increases the fruit's susceptibility to soft scald, soggy breakdown, and flesh browning^[8].

Storage recommendations for short- and long-term storage

The increased production of 'Honeycrisp' apples has created a demand for extending their storability beyond 9 months^[132]. Although fruit firmness decline can be low in air storage^[128], various reports have indicated that fruit flavor often declines under commercial air storage conditions^[133].

'Honeycrisp' apples are prone to numerous physiological disorders (Fig. 3), and several factors impact fruit quality during storage, including storage temperature, growing region, harvest timing, and storage regime^[8,34,131]. Traditionally, 'Honeycrisp' apples have been stored at 3°C following 1 week of conditioning at 10°C to mitigate the risk of low-temperature storage disorders, such as soft scald and soggy breakdown. However, this method can increase the likelihood of bitter pit development in fruit in susceptible orchards^[8]. Utilizing prediction models for bitter pit incidence provides valuable guidance for managing storage and marketing strategies^[11]. It is not currently recommended to store 'Honeycrisp' apples at temperatures close to 0°C ^[34]. Various studies have explored storage strategies based on the fruit's susceptibility to temperature-related disorders, such as bitter pit and soft scald. Bitter pit typically develops during the first month of storage, while soft scald begins after 1 month and continues throughout the storage period. A negative correlation between the development of soft scald and bitter pit has been observed over several years and across different regions^[8]. However, attempts to manage storage disorders based on these interactions have not been successful^[34]. According to preharvest bitter pit prediction models, fruit with a high likelihood of developing bitter pit was stored at 0.5°C for varying durations before being transferred to 3°C to address both disorders. The results showed inconsistent outcomes across years and orchard blocks. In one year, a linear increase in vascular browning susceptibility was observed with prolonged storage at 0.5°C before transferring fruit to 3°C ^[34].

Controlled atmosphere (CA) and dynamic controlled atmosphere (DCA) storage systems have recently been employed to store 'Honeycrisp' apples for over 9 months while maintaining fruit quality, flavor, and consumer preference^[132–135]. However, the cultivar remains susceptible to internal CO_2 injury^[131,133]. In North America, the apple industry has historically mitigated CO_2 injury through treatments such as dipping, drenching, or fumigation with the antioxidant diphenylamine (DPA)^[136]. Since DPA is banned in Europe and for organically grown apples, and may eventually be restricted for conventionally grown apples in the USA, the industry must prepare alternative strategies^[137].

One approach for mitigating internal CO_2 injury involves delaying the start of CA and DCA storage by 4 weeks. During this period, fruit may be conditioned at 10°C for 1 week, depending on the preharvest bitter pit prediction model. If the predicted bitter pit risk is high, conditioning is not recommended. After 1 week, the fruit is transferred to 3°C , and $1\ \mu\text{L/L}$ of 1-MCP is applied 24 h later. The fruit remains at 3°C for 3 weeks before transitioning to CA or DCA storage^[131,132].

For 'Honeycrisp' offspring and sports, there appears to be a recurring trend of releasing new cultivars before completing large-scale

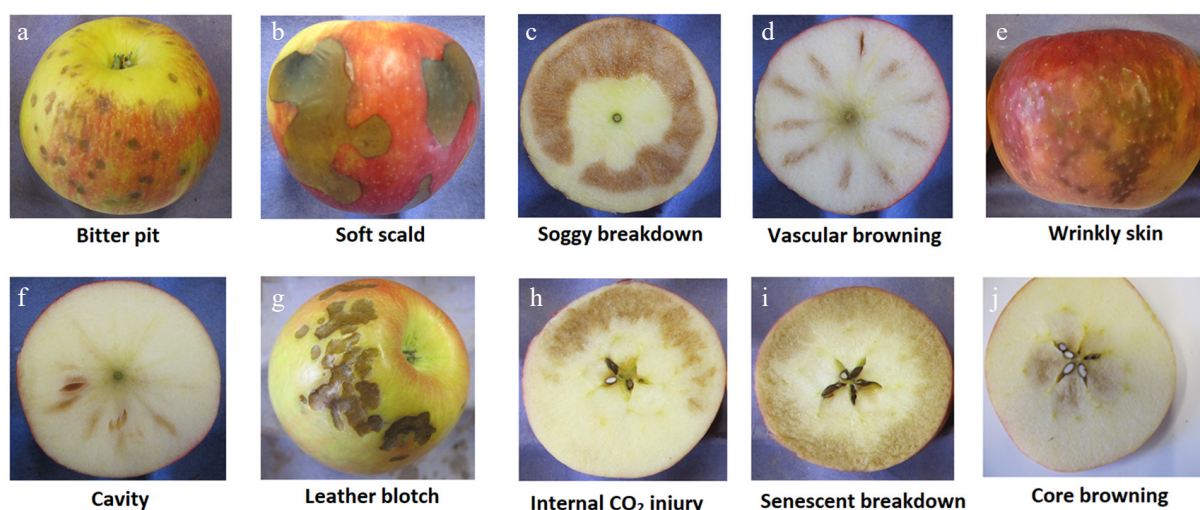


Fig. 3 Physiological disorder development in 'Honeycrisp' apples during storage. (a) Bitter pit, (b) soft scald, (c) soggy breakdown, (d) vascular browning, (e) wrinkly skin, (f) cavity, (g) leather blotch, (h) internal CO₂ injury, (i) senescent breakdown, and (j) core browning. Modified from Al Shoffe et al.^[18].

postharvest evaluations. This is concerning given the need to understand optimal storage practices, particularly when the parent cultivar, like 'Honeycrisp', is prone to physiological disorders. To ensure a comprehensive understanding of both growth and storage characteristics, multi-year storage trials should be incorporated into the final stages of cultivar testing before commercialization.

Conclusion

To optimize 'Honeycrisp' apple production, the apple industry should implement strategies to mitigate biennial bearing, bitter pit, and other physiological disorder losses, including careful nutrient management and storage protocols. Selecting the right rootstocks and implementing high-density planting configurations can enhance orchard productivity. Ongoing research into physiological traits and omics approaches may further refine best practices. Since the release of the 'Honeycrisp' apple in 1991, an analysis of the Google Scholar database indicates that approximately 34% of published papers on *Malus domestica* specifically reference 'Honeycrisp' compared with other cultivars. Given the high market value of 'Honeycrisp' and its offspring as high-value emerging cultivars, in both conventional and organic production, continued investment in predictive models and sustainable management practices will be key to maintaining profitability. Addressing production challenges through integrated techniques will ensure long-term success for growers while supporting the cultivar's role in shaping the future of apple cultivation.

Author contributions

The authors confirm their contributions to the paper as follows: study conception and design: Al Shoffe Y; data organization: Al Shoffe Y; literature search: Al Shoffe Y, Robinson T, Fazio G, Follett E, Clark M, Luby J, Bedford D, Kalcsits L, Peck G; manuscript drafting/revision: Al Shoffe Y, Robinson T, Fazio G, Follett E, Clark M, Luby J, Bedford D, Kalcsits L, Peck G. All authors reviewed the results and approved the final version of the manuscript.

Data availability

All data generated or analyzed during this study are included in this published article.

Acknowledgements

The authors gratefully acknowledge the *Journal of Fruit Research* for covering the Article Processing Charges.

Conflict of interest

The authors declare that they have no conflict of interest.

Dates

Received 20 June 2025; Revised 2 September 2025; Accepted 5 September 2025; Published online 1 November 2025

References

1. Luby J, Bedford D. 1990. Apple tree: Honeycrisp. *US Patent No. 7197*. United States Patent and Trademark Office. <https://patents.google.com/patent/USPP7197P/en>
2. USApple Association. 2024. *2024-25 Apple Production Will Reach Nearly 260 Million Bushels*. <https://usapple.org/news-resources/2024-25-apple-production-will-reach-nearly-260-million-bushels>
3. USApple Association. 2023. *Industry Outlook 2023*. <https://usaa.memberclicks.net/assets/2023/Outlook2023/USAPPLE-OutlookReport-2023.pdf>
4. United States Department of Agriculture-National Agricultural Statistics Service (USDA-NASS). 2022. *Certified Organic Survey 2021 Summary*. https://data.nass.usda.gov/Publications/Highlights/2022/2022_Organic_Highlights.pdf
5. Gallardo RK, Hanrahan I, Hong YA, Luby JJ. 2015. Crop load management and the market profitability of 'Honeycrisp' apples. *HortTechnology* 25:575–84
6. Robinson T, Cheng L, Fazio G, Watkins C, Sazo MM, et al. 2024. Management of 'Honeycrisp' apple trees for maximum sustained yield and minimal bitter pit. *Fruit Quarterly* 32:4–13
7. Valverdi NA, Kalcsits L. 2021. Rootstock affects scion nutrition and fruit quality during establishment and early production of 'Honeycrisp' apple. *HortScience* 56:261–69
8. Al Shoffe Y, Nock JF, Baugher TA, Marini RP, Watkins CB. 2020. Bitter pit and soft scald development during storage of unconditioned and conditioned 'Honeycrisp' apples in relation to mineral contents and harvest indices. *Postharvest Biology and Technology* 160:111044
9. Campbell T, Kalcsits L. 2024. Strategies to overcome biennial bearing in apple – a review. *European Journal of Agronomy* 158:127213

10. Sun C, Zhang W, Qu H, Yan L, Li L, et al. 2022. Comparative physiological and transcriptomic analysis reveal MdWRKY75 associated with sucrose accumulation in postharvest 'Honeycrisp' apples with bitter pit. *BMC Plant Biology* 22:71
11. Al Shoffe Y, Nock JF, Zhang Y, Zhu L, Watkins CB. 2019. Comparisons of mineral and non-mineral prediction methods for bitter pit in 'Honeycrisp' apples. *Scientia Horticulturae* 254:116–23
12. Marini RP, Fazio G. 2018. Apple rootstocks: history, physiology, management, and breeding. *Horticultural Reviews* 45:197–312
13. Fazio G, Wan Y, Kviklys D, Romero L, Adams R, et al. 2014. Dw2, a new dwarfing locus in apple rootstocks and its relationship to induction of early bearing in apple scions. *Journal of the American Society for Horticultural Science* 139:87–98
14. Gonzalez Nieto L, Reig G, Lordan J, Miranda Sazo M, Hoying SA, et al. 2023. Long-term effects of rootstock and tree type on the economic profitability of 'Gala', 'Fuji' and 'Honeycrisp' orchards performance. *Scientia Horticulturae* 318:112129
15. Khan A, Carey SB, Serrano A, Zhang H, Hargarten H, et al. 2022. A phased, chromosome-scale genome of 'Honeycrisp' apple (*Malus domestica*). *Gigabyte* 2022:1–15
16. Baugher TA, Marini R, Schupp JR, Watkins CB. 2017. Prediction of bitter pit in 'Honeycrisp' apples and best management implications. *HortScience* 52:1368–74
17. Bangerth F. 1979. Calcium-related physiological disorders of plants. *Annual Review of Phytopathology* 17:97–122
18. Al Shoffe Y, Nock JF, Zhang Y, Watkins CB. 2021. Physiological disorder development of 'Honeycrisp' apples after pre-and post-harvest 1-methycyclopropene (1-MCP) treatments. *Postharvest Biology and Technology* 182:111703
19. Marini RP, Baugher TA, Muehlbauer M, Sherif S, Crassweller R, et al. 2020. Verification and modification of a model to predict bitter pit for 'Honeycrisp' apples. *HortScience* 55:1882–87
20. Islam MT, Liu J, Das PR, Singh A, Sherif SM. 2022. Rootstock effects on bitter pit incidence in 'Honeycrisp' apples are associated with changes in fruit's cell wall chemical properties. *Frontiers in Plant Science* 13:1034664
21. de Freitas ST, do Amarante CVT, Labavitch JM, Mitcham EJ. 2010. Cellular approach to understand bitter pit development in apple fruit. *Postharvest Biology and Technology* 57:6–13
22. Orcheski B, Meng D, Bai Y, Fei Z, Cheng L. 2021. The transcriptomes of healthy and bitter pit-affected 'Honeycrisp' fruit reveal genes associated with disorder development and progression. *Tree Genetics & Genomes* 17:37
23. Buti M, Poles L, Caset D, Magnago P, Fernandez Fernandez F, et al. 2015. Identification and validation of a QTL influencing bitter pit symptoms in apple (*Malus × domestica*). *Molecular Breeding* 35:29
24. Cheng L, Miranda Sazo M. 2018. Why is 'Honeycrisp' so susceptible to bitter pit? *Fruit Quarterly* 26:19–23
25. Gomez R, Kalcsits L. 2020. Physiological factors affecting nutrient uptake and distribution and fruit quality in 'Honeycrisp' and 'WA 38' apple (*Malus × domestica* Borkh.). *HortScience* 55:1327–36
26. Griffith C, Einhorn TC. 2023. The effect of plant growth regulators on xylem differentiation, water and nutrient transport, and bitter pit susceptibility of apple. *Scientia Horticulturae* 310:111709
27. Tong C, Krueger D, Vickers Z, Bedford D, Luby J, et al. 1999. Comparison of softening-related changes during storage of 'Honeycrisp' apple, its parents, and 'Delicious'. *Journal of the American Society for Horticultural Science* 124:407–15
28. Biggs AR, Peck GM. 2015. Managing bitter pit in 'Honeycrisp' apples grown in the Mid-Atlantic United States with foliar-applied calcium chloride and some alternatives. *HortTechnology* 25:385–91
29. do Amarante CVT, Steffens CA, Ernani PR. 2010. Preharvest identification of bitter pit risk in 'Gala' apples by fruit infiltration with magnesium and analysis of fruit contents of calcium and nitrogen. *Revista Brasileira de Fruticultura* 32:27–34 (in Portuguese)
30. Leisso R, Hanrahan I, Mattheis J. 2019. Assessing preharvest field temperature and at-harvest fruit quality for prediction of soft scald risk of 'Honeycrisp' apple fruit during cold storage. *HortScience* 54:910–15
31. Leisso RS, Buchanan DA, Lee J, Mattheis JP, Sater C, et al. 2015. Chilling-related cell damage of apple (*Malus × domestica* Borkh.) fruit cortical tissue impacts antioxidant, lipid and phenolic metabolism. *Physiologia Plantarum* 153:204–20
32. Leisso RS, Gapper NE, Mattheis JP, Sullivan NL, Watkins CB, et al. 2016. Gene expression and metabolism preceding soft scald, a chilling injury of 'Honeycrisp' apple fruit. *BMC Genomics* 17:798
33. Watkins CB, Nock JF, Weis SA, Jayanty S, Beaudry RM. 2004. Storage temperature, diphenylamine, and pre-storage delay effects on soft scald, soggy breakdown and bitter pit of 'Honeycrisp' apples. *Postharvest Biology and Technology* 32:213–21
34. Al Shoffe Y, Park D, Algul BE, Watkins CB. 2023. Short-term storage of 'Honeycrisp' apples at 0.5 °C does not improve control of bitter pit without increasing low temperature-induced physiological disorders. *Scientia Horticulturae* 321:112364
35. Al Shoffe Y, Watkins CB. 2018. Initial short-term storage at 33 °F reduces physiological disorder development in 'Honeycrisp' apples. *HortTechnology* 28:481–84
36. Gapper NE, Bowen JK, Brummell DA. 2023. Biotechnological approaches for predicting and controlling apple storage disorders. *Current Opinion in Biotechnology* 79:102851
37. Mann H, Bedford D, Luby J, Vickers Z, Tong C. 2005. Relationship of instrumental and sensory texture measurements of fresh and stored apples to cell number and size. *HortScience* 40:1815–20
38. Cabe PR, Baumgarten A, Onan K, Luby JJ, Bedford DS. 2005. Using microsatellite analysis to verify breeding records: a study of 'Honeycrisp' and other cold-hardy apple cultivars. *HortScience* 40:15–17
39. Howard NP, van de Weg E, Bedford DS, Peace CP, Vanderzande S, et al. 2017. Elucidation of the 'Honeycrisp' pedigree through haplotype analysis with a multi-family integrated SNP linkage map and a large apple (*Malus × domestica*) pedigree-connected SNP data set. *Horticulture Research* 4:17003
40. Bedford D. 1991. University of Minnesota's new apple is explosively crisp. *Minnesota Magazine* 91(2)
41. Schmitz CA, Clark MD, Luby JJ, Bradeen JM, Guan Y, et al. 2013. Fruit texture phenotypes of the JRSBRED U.S. apple reference germplasm set. *HortScience* 48:296–303
42. McKay SJ, Bradeen JM, Luby JJ. 2011. Prediction of genotypic values for apple fruit texture traits in a breeding population derived from 'Honeycrisp'. *Journal of the American Society for Horticultural Science* 136:408–14
43. Chang HY, Tong CBS. 2020. Identification of candidate genes involved in fruit ripening and crispness retention through transcriptome analyses of a 'Honeycrisp' population. *Plants* 9:1335
44. Trujillo DI, Mann HS, Tong CBS. 2012. Examination of expansin genes as related to apple fruit crispness. *Tree Genetics & Genomes* 8:27–38
45. Clark JR, Finn CE. 2010. Register of new fruit and nut cultivars list 45. *HortScience* 45:716–56
46. Finn CE, Clark JR. 2008. Register of new fruit and nut cultivars list 44. *HortScience* 43:1321–43
47. Gasic K, Preece JE. 2014. Register of new fruit and nut cultivars list 47. *HortScience* 49:396–421
48. Gasic K, Preece JE, Karp D. 2016. Register of new fruit and nut cultivars list 48. *HortScience* 51:620–52
49. Gasic K, Preece JE, Karp D. 2018. Register of new fruit and nut cultivars list 49. *HortScience* 53:748–76
50. Gasic K, Preece JE, Karp D. 2020. Register of new fruit and nut cultivars list 50. *HortScience* 55:1164–201
51. Karp D, Gasic K. 2022. Register of new fruit and nut cultivars list 51. *HortScience* 57:1174–233
52. Hermann H, Holthusen F, Paap M. 2024. Trends in fruit tree sales in the Lower Elbe region 2022/23. https://www.researchgate.net/publication/380095810_Entwicklung_der_Obstbaumverkaufe_in_der_Niederelberegion_202223
53. Brüggewirth M, Klein N, Harms H. 2021. Fräulein®: Eine deutsche Erfolgsgeschichte? *Mitteilungen des Obstbauversuchsrings* 76:7–16 (in German)
54. Follett EBS. 2023. Effect of Crop Load, Conditioning, Gradual Cooling, and Storage Temperature on the Development of Multiple Physiological

- Disorders in three Norwegian Apple Cultivars*. Master Thesis. Norwegian University of Life Sciences, Ås, Norway
55. Autio W, Robinson T, Blatt S, Cochran D, Francescato P, et al. 2020. Budagovsky, Geneva, Pillnitz, and Malling apple rootstocks affect 'Honeycrisp' performance over eight years in the 2010 NC-140 'Honeycrisp' apple rootstock trial. *Journal of the American Pomological Society* 74:261–69
 56. Lordan J, Fazio G, Francescato P, Robinson T. 2017. Effects of apple (*Malus × domestica*) rootstocks on scion performance and hormone concentration. *Scientia Horticulturae* 225:96–105
 57. Robinson TL, Hoying SA, Fazio G. 2011. Performance of Geneva® rootstocks in on-farm trials in New York State. *Acta Horticulturae* 903:249–55
 58. Robinson T, Lopez S. 2012. Crop load affects 'Honeycrisp' fruit quality more than nitrogen, potassium, or irrigation. *Acta Horticulturae* 940:529–37
 59. Neilsen G, Hampson C. 2014. 'Honeycrisp' apple leaf and fruit nutrient concentration is affected by rootstock during establishment. *Journal of the American Pomological Society* 68:178–89
 60. Fazio G, Lordan J, Francescato P, Robinson T. 2018. Breeding apple rootstocks to match cultural and nutrient requirements of scion varieties. *Fruit Quarterly* 26:25–30
 61. Lordan J, Wallis A, Francescato P, Robinson TL. 2018. Long-term effects of training systems and rootstocks on 'McIntosh' and 'Honeycrisp' performance, a 15-year study in a northern cold climate – part 1: agronomic analysis. *HortScience* 53:968–77
 62. Serra S, Leisso R, Giordani L, Kalcits L, Musacchi S. 2016. Crop load influences fruit quality, nutritional balance, and return bloom in 'Honeycrisp' apple. *HortScience* 51:236–44
 63. Fazio G. 2021. Genetics, breeding, and genomics of apple rootstocks. In *The Apple Genome. Compendium of Plant Genomes*. ed. Korban SS. Cham: Springer International Publishing. pp. 105–30 doi: [10.1007/978-3-030-74682-7_6](https://doi.org/10.1007/978-3-030-74682-7_6)
 64. Fazio G, Adams S, Lordan J, Roberts N, Robinson TL, et al. 2020. Rootstock, scion, and graft type influence graft union flexural strength of apple trees. *Journal of the American Pomological Society* 74:24–44
 65. Donahue DJ, Reig Córdoba G, Elone SE, Wallis AE, Basedow MR. 2021. 'Honeycrisp' bitter pit response to rootstock and region under Eastern New York climatic conditions. *Plants* 10:983
 66. Fazio G, Lordan J, Grusak MA, Francescato P, Robinson TL. 2020. I. Mineral nutrient profiles and relationships of 'Honeycrisp' grown on a genetically diverse set of rootstocks under Western New York climatic conditions. *Scientia Horticulturae* 266:108477
 67. Fazio G, Lordan J, Francescato P, Cheng L, Wallis A, et al. 2018. 'Honeycrisp' apple fruit nutrient concentration affected by apple rootstocks. *Acta Horticulturae* 1228:223–28
 68. Fazio G, Robinson TL. 2022. Time analysis of rootstock mediated nutrient transport in 'Honeycrisp'. *Acta Horticulturae* 1333:405–12
 69. Al Farqani A, Fazio G, Cheng L, Robinson TL. 2025. Effects of soil pH on growth, early fruiting and mineral nutrient profile of 'Honeycrisp' apple trees grafted on eight rootstocks. *Scientia Horticulturae* 342:114029
 70. Lordan J, Francescato P, Fazio G, Robinson TL. 2020. Effects of apple rootstocks on nutrient concentration in 'Honeycrisp' scions in the early orchard life. *Acta Horticulturae* 1281:97–104
 71. Robinson T, Fazio G, Hoying S, Miranda M, Iungerman K. 2011. Geneva® rootstocks for weak growing scion cultivars like 'Honeycrisp'. *New York Fruit Quarterly* 19:10–16
 72. Ferguson IB, Watkins CB. 1989. Bitter pit in apple fruit. *Horticultural Reviews* 11:289–355
 73. Cline J, Autio W, Clements J, Cowgill W, Crassweller R, et al. 2021. Early performance of 'Honeycrisp' apple trees on several size-controlling rootstocks in the 2014 NC-140 rootstock trial. *Journal of the American Pomological Society* 75:189–202
 74. Brown K, Zakalik D, Brown MG, Peck GM. 2025. Integrated approaches are needed to manage weeds in organic apple orchards. *HortScience* 60:519–29
 75. Bradshaw T, Autio W, Blatt S, Clements J, Einhorn T, et al. 2023. Performance of 'Modi®' apple trees on several Geneva rootstocks managed organically: five-year results from the 2015 NC-140 Organic Apple Rootstock Trial. *Journal of the American Pomological Society* 77:14–27
 76. Wooldridge J, Fourie J, Joubert ME. 2013. Effects of soil surface management practices on soil and tree parameters in a'Cripps Pink'/M7 apple orchard-1. Mineral nutrition. *South African Journal of Plant and Soil* 30:163–70
 77. Goh KM, Pearson DR, Daly MJ. 2001. Effects of apple orchard production systems on some important soil physical, chemical and biological quality parameters. *Biological Agriculture & Horticulture* 18:269–92
 78. Arrobas M, Conceição N, Pereira E, Martins S, Raimundo S, et al. 2023. Dolomitic limestone was more effective than calcitic limestone in increasing soil pH in an untilled olive orchard. *Soil Use and Management* 39:1437–52
 79. Farqani AA, Cheng L, Robinson TL, Fazio G. 2024. Effect of solution pH on root architecture of four apple rootstocks grown in an aeroponics nutrient misting system. *Frontiers in Plant Science* 15:1351679
 80. Reitemeier RF. 1951. Soil potassium. *Advances in Agronomy* 3:113–64
 81. Pavan MA, Bingham FT, Peryea FJ. 1987. Influence of calcium and magnesium salts on acid soil chemistry and calcium nutrition of apple. *Soil Science Society of America Journal* 51:1526–30
 82. Cheng L, Miranda Sazo M. 2021. Honeycrisp' requires a lower K level for fruit growth and leaf 810 photosynthesis than 'Gala'. *Fruit Quarterly* 29:4–9
 83. Cheng L, Robinson TL. 2006. Zonal chlorosis of Honeycrisp leaves: Causes and implications. *Compact Fruit Tree* 39:29
 84. Licina V, Krogstad T, Simić A, Akšić MF, Meland M. 2021. Nutrition and fertilizer application to apple trees – a review. *NIBIO Report*. Norwegian Institute of Bioeconomy Research <https://nibio.brage.unit.no/nibio-xmliui/handle/11250/2735389>
 85. Robinson TL, Watkins CB. 2003. Cropload of Honeycrisp affects not only fruit size but many quality attributes. *New York Fruit Quarterly* 11:7–10
 86. Embree CG, Myra MTD, Nichols DS, Wright AH. 2007. Effect of blossom density and crop load on growth, fruit quality, and return bloom in 'Honeycrisp' apple. *HortScience* 42:1622–25
 87. Wünsche JN, Ferguson IB. 2005. Crop load interactions in apple. *Horticultural Reviews* 31:231–90
 88. Robinson T, Lopez S, Iungerman K, Reginato G. 2009. Crop load management for consistent production of 'Honeycrisp' apples. *New York Fruit Quarterly* 17:24–28
 89. Verma P, Sharma S, Sharma NC, Chauhan N. 2023. Review on crop load management in apple (*Malus × domestica* Borkh.). *The Journal of Horticultural Science and Biotechnology* 98:299–321
 90. Robinson TL, Gonzalez L, Cheng L, Ziang Y, Peck G, et al. 2023. Studies in precision crop load management of apple. *Acta Horticulturae* 1366:219–26
 91. Robinson TL, Dominguez LI, Acosta F. 2016. Pruning strategy affects fruit size, yield and biennial bearing of 'Gala' and 'Honeycrisp' apples. *Acta Horticulturae* 1130:257–64
 92. Robinson T, Lakso A, Greene D, Hoying S. 2013. Precision crop load management. *New York Fruit Quarterly* 21:6–9
 93. Tromp J. 1982. Flower-bud formation in apple as affected by various gibberellins. *Journal of Horticultural Science* 57:277–82
 94. Elsyss MA, Hirst PM. 2023. Flowering in 'Honeycrisp' apple shows that spurs are semiautonomous organs. *Journal of the American Society for Horticultural Science* 148:108–16
 95. United States Department of Agriculture (USDA). 2023. *Rules and regulations for the production, handling, labeling, and enforcement of all organic products*. The National Organic Program (NOP). www.ams.usda.gov/rules-regulations/organic
 96. McCartney S, Palmer J, Davies S, Seymour S. 2006. Effects of lime sulfur and fish oil on pollen tube growth, leaf photosynthesis and fruit set in apple. *HortScience* 41:357–60
 97. Peck GM, DeLong CN, Combs LD, Yoder KS. 2017. Managing apple crop load and diseases with bloom thinning applications in an organically managed 'Honeycrisp'/'MM. 111' orchard. *HortScience* 52:377–81
 98. Peck GM, Combs LD, DeLong C, Yoder KS. 2016. Precision apple flower thinning using organically approved chemicals. *Acta Horticulturae* 1137:47–52

99. Kon TM, Schupp JR, Yoder KS, Combs LD, Schupp MA. 2018. Comparison of chemical blossom thinners using 'Golden Delicious' and 'Gala' pollen tube growth models as timing aids. *HortScience* 53:1143–51
100. Zhang S, Gottschalk C, van Nocker S. 2019. Genetic mechanisms in the repression of flowering by gibberellins in apple (*Malus × domestica* Borkh.). *BMC Genomics* 20:747
101. Schmidt T, Elfving DC, McFerson JR, Whiting MD. 2009. Crop load overwhelms effects of gibberellic acid and ethephon on floral initiation in apple. *HortScience* 44:1900–6
102. Robinson T, Lopez S, Iungerman K. 2010. Chemical thinning and summer PGRS for consistent return cropping of 'Honeycrisp' apples. *Acta Horticulturae* 884:635–42
103. Milyaev A, Kofler J, Moya YAT, Lempe J, Stefanelli D, et al. 2022. Profiling of phytohormones in apple fruit and buds regarding their role as potential regulators of flower bud formation. *Tree Physiology* 42:2319–35
104. Cuevas SL. 2009. *Effect of ground and foliar fertilization, irrigation, and cropland on yield and fruit quality at harvest and after cold storage of 'Honeycrisp' apple*. Thesis. Cornell University, Ithaca, NY, USA. <https://ecommons.cornell.edu/items/d656ef79-640f-4c45-a347-b59929d16478>
105. Raven P, Johnson G, Mason K, Losos J, Singer S. 2013. *Biology*. USA: McGraw Hill
106. Griffith C. 2025. *Fruit growth and hormonal regulation of bitter pit in 'Honeycrisp' apple*. Doctoral dissertation. Michigan State University, USA <https://www.proquest.com/docview/3238826493?fromopenview=true&pq-origsite=gscholar&sourcetype=Dissertations%20&%20Theses>
107. Lake Ontario Fruit Program CU. 2024. *The Role of Water in Tree Fruit Physiology & Quality*
108. Reid M, Kalcsits L. 2020. Water deficit timing affects physiological drought response, fruit size, and bitter pit development for 'Honeycrisp' apple. *Plants* 9:874
109. Behboudian M, Mills T, Janick J. 2010. Deficit irrigation in deciduous orchards. *Horticultural Reviews* 21:105–31
110. Kalcsits L, van der Heijden G, Reid M, Mullin K. 2017. Calcium absorption during fruit development in 'Honeycrisp' apple measured using ⁴⁴Ca as a stable isotope tracer. *HortScience* 52:1804–9
111. Telias A, Hoover E, Rosen C, Bedford D, Cook D. 2006. The effect of calcium sprays and fruit thinning on bitter pit incidence and calcium content in 'Honeycrisp' apple. *Journal of Plant Nutrition* 29:1941–57
112. Guerra M, Marcelo V, Valenciano JB, Casquero PA. 2011. Effect of organic treatments with calcium carbonate and bio-activator on quality of 'Reinette' apple cultivars. *Scientia Horticulturae* 129:171–75
113. Rosenberger DA, Schupp JR, Hoying SA, Cheng L, Watkins CB. 2004. Controlling bitter pit in 'Honeycrisp' apples. *HortTechnology* 14:342–49
114. Neilsen G, Neilsen D. 2002. Effect of foliar Zn, form and timing of Ca sprays on fruit Ca concentration in new apple cultivars. *Acta Horticulturae* 594:435–43
115. Yuri J, Retamales J, Moggia C, Vásquez J. 2002. Bitter pit control in apples cv. Braeburn through foliar sprays of different calcium sources. *Acta Horticulturae* 594:453–60
116. Peryea FJ, Neilsen GH, Faubion D. 2007. Start-timing for calcium chloride spray programs influences fruit calcium and bitter pit in 'Braeburn' and 'Honeycrisp' apples. *Journal of Plant Nutrition* 30:1213–27
117. Jemrić T, Fruk I, Fruk M, Radman S, Sinković L, et al. 2016. Bitter pit in apples: pre- and postharvest factors: a review. *Spanish Journal of Agricultural Research* 14:e08R01
118. de Freitas ST, Mitcham EJ. 2012. Factors involved in fruit calcium deficiency disorders. *Horticultural Reviews* 40:107–46
119. Kalcsits LA. 2016. Non-destructive measurement of calcium and potassium in apple and pear using handheld X-ray fluorescence. *Frontiers in Plant Science* 7:442
120. do Amarante CVT, Miqueloto A, de Freitas ST, Steffens CA, Silveira JPG, et al. 2013. Fruit sampling methods to quantify calcium and magnesium contents to predict bitter pit development in 'Fuji' apple: a multivariate approach. *Scientia Horticulturae* 157:19–23
121. Van Der Boon J. 1980. Prediction and control of bitter pit in apples. I. Prediction based on mineral leaf composition, cropping levels and summer temperatures. *Journal of Horticultural Science* 55:307–12
122. do Amarante CVT, Miqueloto A, Steffens CA, Maciel TM, Denardi V, et al. 2018. Optimization of fruit tissue sampling method to quantify calcium, magnesium and potassium contents to predict bitter pit in apples. *Acta Horticulturae* 1194:487–92
123. Autio WR, Bramlage WJ, Weis SA. 1986. Predicting poststorage disorders of 'Cox's Orange Pippin' and 'Bramley's Seedling' apples by regression equations. *Journal of the American Society for Horticultural Science* 111:738
124. Torres E, Recasens I, Àvila G, Lordan J, Alegre S. 2017. Early stage fruit analysis to detect a high risk of bitter pit in 'Golden Smoothee'. *Scientia Horticulturae* 219:98–106
125. Retamales JB, León L, Tomala K. 2001. Methodological factors affecting the prediction of bitter pit through fruit infiltration with magnesium salts in the apple cv. 'Braeburn'. *Acta Horticulturae* 564:97–104
126. Lötze E, Theron KI. 2006. Existing pre-harvest predictions and models for bitter pit incidence. *South African Fruit Journal* 5:20–25
127. Lötze E, Theron KI. 2006. Dynamics of calcium uptake with pre-harvest sprays to reduce bitter pit in 'Golden Delicious'. *Horticulturae* 721:313–20
128. Harb J, Gapper NE, Giovannoni JJ, Watkins CB. 2012. Molecular analysis of softening and ethylene synthesis and signaling pathways in a non-softening apple cultivar, 'Honeycrisp' and a rapidly softening cultivar, 'McIntosh'. *Postharvest Biology and Technology* 64:94–103
129. Telias A, Hoover E, Rother D. 2008. Plant and environmental factors influencing the pattern of pigment accumulation in 'Honeycrisp' apple peels using a novel color analyzer software tool. *HortScience* 43:1441–45
130. Watkins CB, Erkan M, Nock JF, Iungerman KA, Beaudry RM, et al. 2005. Harvest date effects on maturity, quality, and storage disorders of 'Honeycrisp' apples. *HortScience* 40:164–69
131. DeEll JR, Lum GB, Ehsani-Moghaddam B. 2016. Effects of delayed controlled atmosphere storage on disorder development in 'Honeycrisp' apples. *Canadian Journal of Plant Science* 96:621–29
132. Al Shoffe Y, Rudell D, Park D, Algul BE, Qin M, et al. 2024. Aroma volatiles and sensory quality of organically-grown apple cultivars after dynamic controlled atmosphere (DCA) storage and comparison with CA-stored fruit with and without 1-methylcyclopropene (1-MCP). *Postharvest Biology and Technology* 218:113162
133. Watkins CB, Nock JF. 2012. Controlled-atmosphere storage of 'Honeycrisp' apples. *HortScience* 47:886–92
134. Leisso RS, Hanrahan I, Mattheis JP, Rudell DR. 2017. Controlled atmosphere storage, temperature conditioning, and antioxidant treatment alter postharvest 'Honeycrisp' metabolism. *HortScience* 52:423–31
135. Mattheis J, Rudell DR. 2021. 'Honeycrisp' apple (*Malus domestica* Borkh.) fruit response to controlled atmosphere storage with the low oxygen limit established by monitoring chlorophyll fluorescence. *HortScience* 56:173–76
136. Contreras C, Alsmairat N, Beaudry R. 2014. Prestorage conditioning and diphenylamine improve resistance to controlled-atmosphere-related injury in 'Honeycrisp' apples. *HortScience* 49:76–81
137. Prange RK, Wright AH, DeLong JM, Zanella A. 2015. A review on the successful adoption of dynamic controlled-atmosphere (DCA) storage as a replacement for diphenylamine (DPA), the chemical used for control of superficial scald in apples and pears. *Acta Horticulturae* 1071:389–96



Copyright: © 2025 by the author(s). Published by Maximum Academic Press, Fayetteville, GA. This article is an open access article distributed under Creative Commons Attribution License (CC BY 4.0), visit <https://creativecommons.org/licenses/by/4.0/>.