## **Open Access**

## https://soilfun.org/, a web-based platform for soil-inhabiting Ascomycota species

Weeragalle Arachchillage Erandi Yasanthika<sup>1,2,3</sup>, Antonio Roberto Gomes de Farias<sup>1\*</sup>, Dhanushka N. Wanasinghe<sup>4</sup>, Kandawatte Wedaralalage Thilini Chethana<sup>1,2</sup>, Rasoul Zare<sup>5</sup>, Lei Cai<sup>6</sup>, Sajeewa S. N. Maharachchikumbura<sup>7</sup>, Danushka S. Tennakoon<sup>8,9</sup>, Rekhani Hansika Perera<sup>10</sup>, Thatsanee Luangharn<sup>1</sup> and Putarak Chomnunti<sup>2</sup>

<sup>2</sup> School of Science, Mae Fah Luang University, Chiang Rai 57100, Thailand

<sup>4</sup> Center for Mountain Futures, Kunming Institute of Botany, Honghe County 654400, Yunnan, PR China

- <sup>5</sup> Iranian Research Institute of Plant Protection (IRIPP), Agricultural Research, Education and Extension Organization, PO Box 1454, Tehran 19395, Iran
- <sup>6</sup> State Key Lab of Mycology, Institute of Microbiology, Chinese Academy of Sciences, Beijing 100101, PR China

<sup>7</sup> University of Electronic Science and Technology of China, Chengdu 611731, PR China

- <sup>8</sup> Department of Biology, Faculty of Science, Chiang Mai University, Chiang Mai 50200, Thailand
- <sup>9</sup> Research Center of Microbial Diversity and Sustainable Utilization, Chiang Mai University, Chiang Mai 50200, Thailand
- <sup>10</sup> School of Biological Sciences and Institute of Microbiology, Seoul National University, Seoul 08826, Republic of Korea

\* Corresponding author, E-mail: antonio.gom@mfu.ac.th

## Abstract

Soil covers a large area of the earth's crust and supports a high level of bio-diversity. This diversity is influenced by numerous geobiological variables, which impact soil fungal distribution worldwide. The taxonomy of soil-associated fungi has been investigated over the years using various approaches. To recover below-ground fungal species, researchers currently use culture-dependent and culture-independent approaches such as high-throughput techniques. As a result, a significant amount of taxonomic information is being generated and scattered throughout many publications. Most recent publications and databases of soil *Ascomycota* are derived from high-throughput techniques mainly focused on higher taxonomic ranks; however, information of the genera and species identified from soils are not conglomerated into a single database. Therefore, the Soil Fungi website aims to create an online platform to compile the soil-associated fungal species with comprehensive taxonomic descriptions and conglomerated on the website where users have quick and user-friendly access. This paper introduces the https://soilfun.org/ website with its objectives and functions and provides a guideline for accessing and retrieving information. Users will be able to access and retrieve information from the website easily. The Soil Fungi website will be an essential tool for researchers, students, and other interested parties in soil mycology.

Citation: Yasanthika WAE, Gomes de Farias AR, Wanasinghe DN, Chethana KWT, Zare R, et al. 2023. https://soilfun.org/, a web-based platform for soil-inhabiting Ascomycota species. Studies in Fungi 8:16 https://doi.org/10.48130/SIF-2023-0016

## Introduction

Soil is a remarkably complex substrate that provides various niches for microbial communities. In this environment, fungi are the main biotic component and mediate many below-ground functions. However, their distribution is variable due to inconsistencies in resource levels (nutrition, soil, water, and air)<sup>[1,2]</sup>. Besides, most fungi spend at least one stage of their life in the soil, showing different trophic modes such as pathogenic, saprobic, or mutualistic<sup>[3,4]</sup>. Studies have shown that *Ascomycota*, *Basidiomycota*, *Chytridiomycota*, *Glomeromycota*, *Mortierellomycota* and *Rozellomycota* are widely distributed in edaphic habitats<sup>[5–7]</sup>.

Culturing and high-throughput sequencing (HTS) methods are currently applied for detecting belowground *Ascomycota* groups<sup>[5,7–9]</sup>. *Ascomycota* is represented by dominant classes in soil, including *Arthoniomycetes*, *Dothideomycetes*, *Euro*-

© The Author(s)

tiomycetes, Leotiomycetes, and Sordariomycetes<sup>[5–7,10,11]</sup>. Of them, some genera, such as Fusarium, Penicillium, and Phoma, are prevalent<sup>[7,12]</sup>, while others, such as Ambrosiella, Arcopilus, Didymocyrtis, Kionochaeta, and Parathyridaria have rarely been isolated<sup>[13–15]</sup>. This is because exclusive adaptations to belowground environments and a lack of isolation techniques make it difficult to recover Ascomycota species. Most soil ascomycetes are identified at higher taxonomic ranks through HTS, which led to many recent publications and databases showing the taxonomy and distribution patterns of belowground fungi only up to the genus level<sup>[16]</sup>.

High-throughput sequencing has impacted the estimation of the number of fungi and the endemicity of soil taxa in different parts of the world<sup>[8,17,18]</sup>. Some databases have attempted to gather information on soil fungal diversity based on HTS. For example, the GlobalFungi (https://globalfungi.com/) provides information on taxonomy, ecology, and reference sequences of

<sup>&</sup>lt;sup>1</sup> Center of Excellence in Fungal Research, Mae Fah Luang University, Chiang Rai 57100, Thailand

<sup>&</sup>lt;sup>3</sup> Innovative Institute for Plant Health, Zhongkai University of Agriculture and Engineering, Guang Dong Province 510225, PR China

taxa from different habitats, including soil<sup>[17]</sup>. The Global Soil Mycobiome consortium dataset, a global soil study recovered 722,682 fungal Operational Taxonomic Units (OTUs), representing 62.4% of the total OTUs<sup>[7]</sup>. Among them, 330,054 OTUs were assigned to *Ascomycota*. The total number of fungi is estimated as 6.28 million species<sup>[9]</sup> and found the highest diversity from soil and litter samples, revealing that soil fungal sequences have 75% overlap with previously known species. In addition, some studies have recovered unculturable fractions representing several new lineages at the phylum level<sup>[4–6,10,19]</sup>.

Few studies compare the taxa recovered by HTS and culturing methods from the same sample or location. For example, Stefani et al.<sup>[20]</sup> compared the fungal diversity from petrochemical-contaminated soils in Canada. They found that the number of OTUs ranged from 153 to 235 depending on the level of contamination (slightly to highly contaminated), and culturing methods vielded 8 to 43, respectively. Based on these results, we can roughly estimate that only around 5%-65% of the soil fungal community can be cultured in any environment and we need to develop methods to approach the rest of the 35%–95%. A study on Antarctic crypto-endolithic fungal communities identified several black fungal species that were only detectable through cultivation and found a 12% overlap of species from both culture-dependent and -independent approaches, which resulted in 4,618 colonies and 1,439,748 sequences<sup>[21]</sup>. The successful isolation of these species highlights the importance of using both complementary techniques<sup>[21]</sup>. Compared to overall HTS results, the cultured fraction is smaller as they have vast differences between the two approaches. However, HTS results can excel in culturing the soil taxa, for example, to plan on isolating targeted taxa or selecting suitable fungal isolating methods in a particular area<sup>[8,16]</sup>. Culturing is essential to obtain molecular and morphological data of species and recover the undetectable ones from HTS.

However, around 80% of all soil-inhabiting taxa are not assigned to species level, and 20% to known orders<sup>[5,19,22]</sup>. Thus, the soil can potentially reveal many undiscovered species in the future<sup>[8,9]</sup>. Contemporary studies have described and resolved soil-associated *Ascomycota* species from *Dothideomycetes*, *Eurotiomycetes*, *Leotiomycetes*, and *Sordariomycetes* with morphomolecular support<sup>[23–28]</sup>. However, no recently updated study or database shows the total number of species isolated from the soil<sup>[16]</sup>.

Over the last few decades, several books have been published based on morphology, such as Compendium of soil fungi<sup>[29–31]</sup>, Manual of soil fungi<sup>[32]</sup>, The Genera of Hyphomycetes from soil<sup>[33]</sup>, and Soil fungi: diversity and detection<sup>[34]</sup>, which provided the diversities of worldwide soil species with comprehensive morphological illustrations. The Atlas of soil ascomycetes<sup>[35]</sup> illustrated anamorphic and teleomorphic (fruit body and hyphomycetes form on cultures) structures, culture characteristics, and keys to the species. Currently, many taxonomists have adopted DNA sequences to determine species boundaries. However, species delineation criteria can differ among groups, such as cryptic species, pathogenic genera, and species complexes that need solid phylogenetic resolution<sup>[36-39]</sup>. In this regard, soil-associated species are a highly complex group, and taxonomists have been using the polyphasic approach, combining morphological, physiological, ecological, molecular methods and to determine species<sup>[36,38,40,41]</sup>.

Precise identification of fungal species/genera is critical for further use in taxonomy and biotechnology research<sup>[8,16,42-44]</sup>. For example, pathogens are important in disease management, while soil-borne extremophilic genera (e.g., Verruconis) are sources of bioactive compounds<sup>[42,45,46]</sup>. Thus a database integrating taxonomy, ecological, and economic information of soil fungi helps to identify these organisms by end-users. The Soil Fungi website aims to collect and provide accurately identified soil fungi and continuously update checklists and notes on genera. This will increase the availability of information in the field and provide a valuable resource for researchers and students studying soil fungi. The website will be a central database of soil-associated fungal species, including their taxonomic descriptions and other relevant information. Users will be able to easily access and retrieve information from the website, such as the distribution of certain fungal species and their ecological roles. The website will also provide a platform for collaboration between researchers and interested parties, allowing information and idea sharing. Furthermore, the Soil Fungi website will help estimate the number of soil genera discovered over time. This information will be useful for understanding the diversity and distribution of soil fungi and for developing conservation strategies for these important organisms.

# Why is it important to have a database for soil fungi?

The current information on soil fungal taxonomy is available as HTS and culture-dependent data<sup>[4,6,8,47-50]</sup>. These two forms of data can cause contradictions in species identifications and seem unstable and affect studies. Moreover, the number of fungal species in the soil has been estimated from HTS methods, but there is no updated account for recently identified or recorded species from the soil. In addition, morphomolecular identifications also face shortfalls as some genera are well-studied, while some have phylogenetic ambiguities<sup>[51-53]</sup>. For example, specious genera, such as Aspergillus, Cladosporium, Fusarium, Mucor, Penicillium, and Trichoderma are common in soil. Due to the similar morphologies resulting from hybridization, cryptic speciation, and convergent evolution, their species delineation is difficult<sup>[54]</sup>. Therefore, compiling updated phylogenetic and morphological data of the genera/ species present in the soil are essential. It is also important to gather data on the type of soil ecosystem, locality, identification method, and economic impact of the species or genera. Since most of the information is scattered among various publications and databases, compiling them into a single database is beneficial for mycologists, ecologists, clinical studies, and the general public.

Even though information about soil fungi is available in online databases e.g., Fungal Genera<sup>[55]</sup>, GlobalFungi<sup>[17]</sup>, UNITE<sup>[56]</sup>, and Global Soil Mycobiome consortium (GSMc)<sup>[7]</sup>, they lack specific information on fungal species and relevant morpho-molecular data. Further, in other databases like 'Faces of Fungi' (www.facesoffungi.org)<sup>[57]</sup>, 'Dothideomycetes' (www. dothideomycetes.org/)<sup>[58]</sup>, 'Sordariomycetes' (https://sordariomycetes.org/)<sup>[59]</sup>, GMS microfungi (www.gmsmicrofungi.org/)<sup>[60]</sup>, and 'Coelomycetes' (https://coelomycetes.org/)<sup>[61]</sup> information about *Ascomycota* studies are commonly found. However, a website compiling morpho-molecular data of soil fungi has not been developed. Therefore, we developed and

#### A web-based platform for Ascomycota in soil

implemented a web-based platform as the information hub for soil fungi research. The website provides a checklist of all the soil-associated *Ascomycota* genera. These generic notes include taxonomic descriptions, phylogenetic data, and other hosts/substrates for each species, accompanied by photographic plates or drawings and detection methods. It also provides a list of novel or existing species of soil fungi for each genus, which will be updated over time. The data from previous studies will be periodically included to disseminate information in a holistic approach, enhancing the in-depth understanding of the soilinhabiting fungi and resource for retrieving multiple data.

## Soil Fungi website

The website provides a comprehensive checklist of all the soil-associated genera in Ascomycota essential for identifying and classifying soil fungi. The generic notes contain morphological descriptions, information on phylogenetic analyses such as gene regions, a summary of the latest phylogenetic revisions, ecological and economic impacts and the distribution. For each genus, a list of novel or existing soil-inhabiting species is provided, which will be uploaded and updated over time. Each species is accompanied by taxonomic descriptions, photographic plates or drawings, and detection methods. Each species/genera description is linked to the original publication(s), thus, users have-real time access to further details (Fig. 4). The website ensures that researchers can access the latest information about soil fungal taxonomy and keep up with the latest research developments. The data from previous studies are rearranged, and future studies will be periodically included to disseminate information in a holistic approach. Therefore, the website remains an up-to-date resource for researchers, providing them access to the latest research on soil fungi. These data will enhance an in-depth understanding of the soilinhabiting fungi and will be important for future research. We encourage all the taxonomists and interested parties who work on soil fungi to contribute their published data to this website which will enhance the information availability and ensures that the website becomes an invaluable resource for retrieving multiple data, providing researchers a platform to share their research and collaborate with others in the field.

#### Construction

On the website, taxa are arranged according to the latest classification<sup>[53]</sup>. The history of soil taxonomy is briefly described, and links to related literature are provided. All taxonomic entries are checked by the curators prior to uploading and published in a consistent format. The list of curators and their contact details are displayed in Table 1.

## Interface and visualization of the database

The website is available at https://soilfun.org/ and provides a user-friendly interface to retrieve updated information on the classification of soil-inhabiting *Ascomycota*. It is easy to access and navigate. The homepage comprises a navigation bar and a search tool to retrieve information.

## Home page

The Home page displays the summary of the project (Fig. 1). A navigation bar is displayed at the top of the page. Below the navigation bar, the main objectives of the webpage are Table 1. List of curators for the https://soilfun.org/ website.

Position	Name	Contact details
Head curator	Dr. A. Roberto G. Farias	rfariasagro@gmail.com
Expert curator	Prof. Dr. Rasoul Zare	simplicillium@yahoo.com
Managing curator	Ms. W. A. Erandi Yasanthika	eyasanthika@gmail.com
Curators	Prof. Dr. Kevin D. Hyde	kdhyde3@gmail.com
	Prof. Dr. Lei Cai	cail@im.ac.cn
	Prof. Dr. Sajeewa Maharachchikumbura	sajeewa83@yahoo.com
	Dr. Dhanushka	dnadeeshan@gmail.com,
	Wanasinghe	wanasinghe@mail.kib.ac.cn
	Dr. K. W. Thilini Chethana	kandawatte.thi@mfu.ac.th
	Dr. Danushka Sandaruwan Tennakoon	danushkasandaruwanatm@gmail.com
	Dr. Rekhani Hansika Perera	hansi.rekha2@gmail.com

displayed in the left column, while the right column displays the search tool and the recently updated entries. The bottom of the page shows the publisher details, copyright information, and contact details. The navigation bar consists of eight menu items (Home, Outline, Archives, Reference, Curators, History, Reference, Notes, and Contact). These menu items facilitate easy navigation throughout the website, allowing users to access the information they need, quickly and efficiently

#### Outline

Under this navigation menu item, the website presents the latest taxonomic representation of genera that are recorded from soil obtained from peer-reviewed publications (Fig. 2). The user can scroll down to find any taxa or the search bar is optional, where the user can type preferred taxa, and be directed to the exact location. Each taxon is linked to its corresponding taxonomic description, which can be accessed by clicking on the relevant name.

#### Archives

The orders to which different fungi isolated from soil belong are listed alphabetically under this menu item (Fig. 3). When the user clicks on a preferred order, a dropdown list containing its families and a 'Read more about...' link to the description of each hierarchical taxonomic level is displayed. Similarly, when the user clicks on a family, it shows the related genera and a 'Read more about...' link to the family description. When the user selects a preferred genus, a 'Read more about...' link to the description of the genus and a list of soil-associated species linked to their taxonomic descriptions and illustrations are shown (Fig. 4). Each taxonomic description includes a web source link, enabling the reader to access the original publication directly. This information is continuously updated according to the latest publications, ensuring user access to the most up-to-date and accurate information on soil-inhabiting Ascomycota.

#### Curators

Curators and contact details are provided under this menu item (Fig. 5). These curators are expert fungal taxonomists who continuously monitor the webpage and suggest improvements.

#### History

Under this menu item, the evolution of soil fungal taxonomy is briefly discussed, together with the available culture-dependent and independent approaches (Fig. 6).

#### References

References used in the entries are provided under this menu.

#### Notes

Important events, information, and the latest trends in soil fungal taxonomy are provided here.

#### Contact

Under this menu item, a dialogue box is provided for the users to exchange information directly or share their suggestions and/or comments. Furthermore, the email and the address of the publisher are also given.

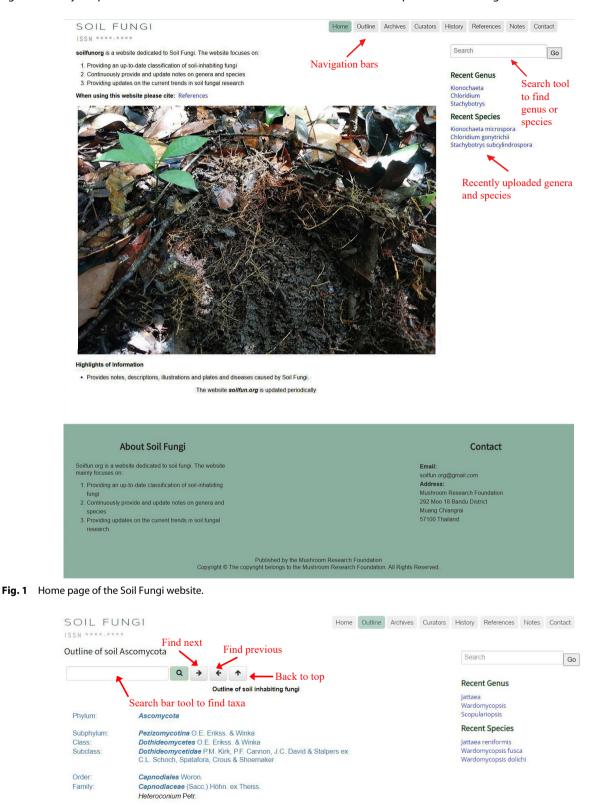


Fig. 2 'Outline' contains soil inhabiting Ascomycota taxa according to the latest classification.

## A web-based platform for Ascomycota in soil

SOIL FUNGI	Home Outline Archives Curators History References Notes Contact
Soil Fungi Heirarchy	Search
Amphisphaeriales	Search Go
Calosphaeriales	Recent Genus
Chaetosphaeriales	Jattaea
Hypocreales - Order	Wardomycopsis Scopulariopsis
Read more about Hypocreales orders »	Recent Species
Cordycipitaceae - Family	Jattaea reniformis
Read more about Cordycipitaceae family »	Wardomycopsis fusca Wardomycopsis dolichi
Flavocillium ← Genus	wardoniyeopsis doichi
Read more about Flavocillium genus »	
Flavocillium primulinum Species	
Flavocillium subprimulinum	
Gamszarea	
Lecanicillium	
Simplicillium	
Stachybotryaceae	
Microascales	

Fig. 3 'Archives' configured with a dropdown list of taxonomic hierarchy with links to order, family, genera, and species descriptions.

S O LI	FUNGI					Home	Outline	Archives	Curators	History	References	Notes	Contact
ISSN ****						TIOTTIC	Outime	Archives	Culators	Thatory	References	140(03	Contact
Amphisphaer	riales » Beltraniac	ese » Beltrar	niella										
Amphisphaeriales » Beltraniaceae » Beltraniella Beltraniella fertilis Species name								Recent Genus					
Beltraniella fertilis Heredia, R.M. Arias, M. Reves & R.F. Castañeda, Fungal Diversity 11: 100 (2002) Fig. X							Pseudopithomyces Yunnania Eosphaeria						
Index Fungorum number: IF489903; Faces of fungi number: FoF 03632													
Colonie	-ungorum number es on PDA pale w	hite, reaching	aces of fung q a diam. of :	2-3 cm in 4 days	s at 25°C, flat and cir	cular-shap	ed, pale br	own, smoot	h at surface				
Colonies on PDA pale white, reaching a diam. of 2–3 cm in 4 days at 25°C, flat and circular-shaped, pale brown, smooth at surface and produce highly branched melanized hyphae with brownish exudates in old cultures, after 16 weeks, with conidophores forming on the mycelium; reverse light vellow to dark brown. Sexual morph: Hyphomycetous. Mycelium mostly immersed in							Recent Species						
the substrature small groups, at the base, thick-walled to prown to sub to polyblastic,	m, composed of s , thick-walled, verr tapering to a poir o smooth-walled, t hyaline at the sw , integrated, termi	eptate, bran rucose, dark l nted apex. <i>C</i> 6.4–91.5 μm ollen base, p nal. <i>Conidia</i>	ched subhya brown at the conidiophores long, 2.3–6.5 aler and slig solitary to ag	line hyphae. Set base, paler at ap macronematous $5 \mu m$ wide ( $\overline{x} = 37$ htly tapering towa gregated, acroge	so. Asexual morph: H; ae numerous, erect, s bex, 61.7-149.8 µm liss, sometimes setiform 7 µm × 3.4, n = 30), s ards a pointed apex. anous, simple, dry, st yaline to sub hyaline.	straight or ong, 2.5–7 n; single, s ometimes l <i>Conidiogen</i> raight, smo	flexuous, u µm wide (3 straight, se branched a nous cells h both, thin-w	nbranched, $\overline{x} = 113 \times 4$ ptate, partly t the apical holoblastic, i alled, bicon	single or in µm, n = 30) verrucose, region, dark monoblastic ic, turbinate	Stella Pseud	naeria uliginosa tospora terrico doneurospora a	la	oorcata
07.000 0 -	20) in the broodes	tnort								lorpho	logical de	scripti	on
Distribu	ution – Atlantic fore hosts – Dead leav	ests, Brazil, N ves of Manai	Aexico, Thaila fera indica ar	and <b>erinari alvimi</b>	Host-substrate	; and d	Istribut	ION		1	0	1	
Materia	al examined – Tha	iland, Chiang	Mai Provinc	e, Mae Tang dist	rict, Ban Pa Deng, Mi								
	", 905 m, in forest UCC 20-0119.	soil (dominat	ted by Dipter	ocarpaceae), 20 <sup>4</sup>	<sup>h</sup> March 2019, Erandi	Yasanthik	a, Erscm11	(MFLU 20-0	0506), living				
lotes:	artilla was introduc	od by Horodi	in at al (200	2) based on mor	phological characteris	tion The s	oguopoo d	ata for this					
					g leaves collected in		sequence as	ata for this s	species was				
2			State State	and the second									
			1 mil	a sta									
1. its	1	-	1.1.1.1	38 29									
A State	a contra la	1	1.723	1									
Re Way			Nº F	6 2 3									
			- Martine	- Colorest									
			1	h									
	(the state	1	E	0									
	A		B	Y									
23	all as		9										
24	SHE	F	1										
Te		13											
F	1 and	11											
i j	~ <sup>k</sup> Q	- Ø											
82	9		b										
7	LA I	g	Reg .										
U.	4	R	6	1									
m	0 n	0	р	q									
	e a	8	0	(23)									
	9	5/	1		Figure le	aand							
Office		U.	2		rigure le	genu							
Eigure Y P.	eltraniella fertilia (	MELLICC 20	0119) a Mat		A after 16 weeks with	the encru	lation b Dr	warea of the	a colonies o				
PDA after 16	6 weeks. c Sporu	lation of the	colony with	conidial attach	ments on the myceli	um. d Imr	nature ase	ptate hypha	e. e Matur	e			
					cose pigmented setar le bars: g = 25 µm d,								
ım.				,									
References:													

Heredia G, Arlas RM, Reyes M, Castañeda–Ruiz R. 2002 – New anamorph fungi with rhombic conidia from Mexican tropical forest litter. Fungal Diversity 11, 99–107.

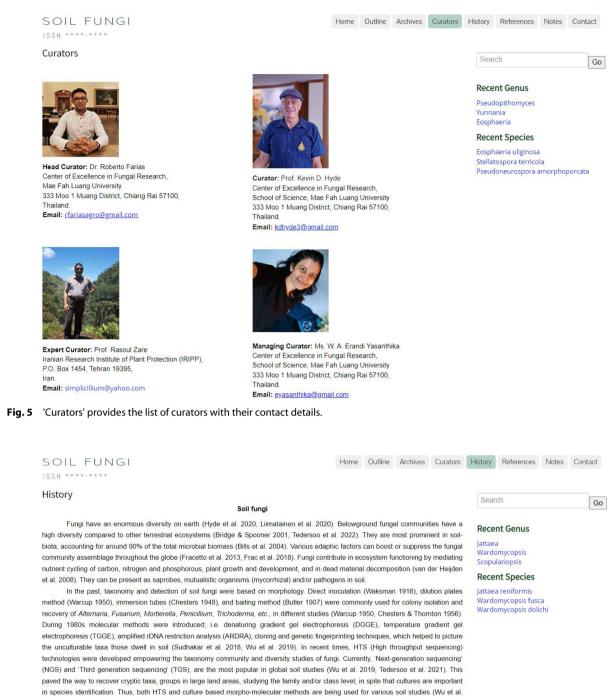
Lin CG, Hyde KD, Lumyong S, Mckenzie EHC. 2017b – Beltrania–like taxa from Thailand. Cryptogamie, Mycologie 38, 301–319

#### Source:

Data and figures were reprinted with permission from: Yasanthika WAE, Wanasinghe DN, Karunarathna SC, Bhat DJ, Samarakoon SMBC, Ren GC, Monkal J, Mortimer PE, Hyde KD 2020 – Two new Sordariomycetes records from forest soils in Thailand. Asian Journal of Mycology 3(1), 456–472, Doi 10.5943/ajom/3/1/16-in AJOM

Fig. 4 Information provided for the soil-inhabiting species with taxonomic description.

#### A web-based platform for Ascomycota in soil



Aspergillus, Fusarium, Penicillium and Trichoderma are common genera in soil, recovered from many parts of the world by the use of both culturing and HTS methods (Tedersoo et al. 2017, Wang & Zhuang 2022). Recent studies have isolated strains of genera which used to be rare in the soil; i.e. Calvatia, Chloridium, Cortinarius, Exophiala, Juxtiphoma, Kionochaeta, Kosmimatamyces, Ogataea, Phialoparvum, Pichia, Plectosphaerella, and Zygoascus (Groenewald et al. 2018, Giraldo, et al. 2019, Yasanthika et al. 2022).

Species identification in soil is critical and some of the cultures are sources of biotechnology as HTS methods are still lagged behind (Gutleben et al. 2018, Hyde et al. 2019). Thus, we use culturing and morpho-molecular approaches in their identification. In https://soilfun.org/ present genera and species introduced or recorded from soils all over the world.

Fig. 6 'History' provides the history and development of studies on soil fungal taxonomy.

### Acknowledgments

2019, Chethana et al. 2020).

The authors would like to thank all the contributors who will provide entries for this website. This project is supported by The National Research Council of Thailand (NRCT) grant 'Total fungal diversity in a given forest area with implications towards species numbers, chemical diversity and biotechnology' (grant number N42A650547) and the Mushroom Research Foundation (MRF). A.R. Gomes de Farias thanks Mae Fah Luang University new researcher grant 'Diversity and host specificity of soil fungi in forest ecosystems in Chiang Rai, Thailand' (grant number 662A16047). W.A.E. Yasanthika thanks Theisis support grant from Mae Fah Luang University; reference number 7702(6)/580.

## **Conflict of interest**

The authors declare that they have no conflict of interest.

## Dates

Received 7 July 2023; Accepted 22 August 2023; Published online 20 October 2023

#### References

- Coleine C, Selbmann L, Guirado E, Singh BK, Delgado-Baquerizo M. 2022. Humidity and Low PH Boost Occurrence of Onygenales Fungi in Soil at Global Scale. *Soil Biology and Biochemistry* 167:108617
- Li J, Cui L, Delgado-Baquerizo M, Wang J, Zhu Y, et al. 2022. Fungi drive soil multifunctionality in the coastal salt marsh ecosystem. *Science of The Total Environment* 818:151673
- Wilkes TI, Warner DJ, Edmonds-Brown V, Davies KG, Denholm I. 2021. Zero tillage systems conserve arbuscular mycorrhizal fungi, enhancing soil glomalin and water stable aggregates with implications for soil stability. *Soil Systems* 5:4
- Senanayake IC, Pem D, Rathnayaka AR, Wijesinghe SN, Tibpromma S, et al. 2022. Predicting global numbers of teleomorphic ascomycetes. *Fungal Diversity* 114:237–78
- Tedersoo L, Bahram M, Põlme S, Kõljalg U, Yorou NS, et al. 2014. Global diversity and geography of soil fungi. *Science* 346:e1256688
- Tedersoo L, Anslan, S, Bahram M, Kõljalg, U, Abarenkov K. 2020. Identifying the 'Unidentified' fungi: A global-scale long-read thirdgeneration sequencing approach. *Fungal Diversity* 103:273–93
- Tedersoo L, Mikryukov V, Anslan S, Bahram M, Khalid AN, et al. 2021. The global soil mycobiome consortium dataset for boosting fungal diversity research. *Fungal Diversity* 111:573–88
- Wu B, Hussain M, Zhang W, Stadler M, Liu X, et al. 2019. Current insights into fungal species diversity and perspective on naming the environmental DNA sequences of fungi. *Mycology* 10:127–40
- Baldrian P, Větrovský T, Lepinay C, Kohout P. 2022. High-throughput sequencing view on the magnitude of global fungal diversity. *Fungal Diversity* 114:539–47
- 10. Blackwell M. 2011. The fungi: 1, 2, 3...5.1 million species? American Journal of Botany 98:426–38
- Zhou J, Jiang X, Zhou B, Zhao B, Ma M, et al. 2016. Thirty four years of nitrogen fertilization decreases fungal diversity and alters fungal community composition in black soil in northeast China. *Soil Biology and Biochemistry* 95:135–43
- Crous PW, Hernández-Restrepo M, van Iperen AL, Starink-Willemse M, Sandoval-Denis M, et al. 2021. Citizen science project reveals novel fusarioid fungi (*Nectriaceae, Sordariomycetes*) from urban soils. *Fungal Systematics and Evolution* 8:101–27
- Park S, Ten L, Lee SY, Back CG, Lee JJ, et al. 2017. New recorded species in three genera of the sordariomycetes in Korea. *Mycobiology* 45:64–72
- Lee SH, Park HS, Nguyen TTT, Lee HB. 2019. Characterization of three species of *Sordariomycetes* isolated from freshwater and soil samples in Korea. *Mycobiology* 47:20–30
- Das K, Lee SY, Jung HY. 2021. Morphology and phylogeny of two novel species within the class *Dothideomycetes* collected from soil in Korea. *Mycobiology* 49:15–23
- Yasanthika WAE, Wanasinghe DN, Mortimer PE, Monkai J, Farias ARG. 2022. The importance of culture-based techniques in the genomic era for assessing the taxonomy and diversity of soil fungi. *Mycosphere* 13:724–51

Yasanthika et al. Studies in Fungi 2023, 8:16

- Větrovský T, Morais D, Kohout P, Lepinay C, Algora C, et al. 2020. GlobalFungi, a global database of fungal occurrences from highthroughput-sequencing metabarcoding studies. *Scientific Data* 7:228
- Tedersoo L, Mikryukov V, Zizka A, Bahram M Hagh-Doust, N, et al. 2022. Global Patterns in endemicity and vulnerability of soil fungi. *Global Change Biology* 28:6696–710
- Tedersoo L, Bahram M, Puusepp R, Nilsson RH, James TY. 2017. Novel soil-inhabiting clades fill gaps in the fungal tree of life. *Microbiome* 5:42
- Stefani FOP, Bell TH, Marchand C, De La Providencia IE, El Yassimi A, et al. 2015. Culture-dependent and -independent methods capture different microbial community fractions in hydrocarboncontaminated soils. *PLoS One* 10:e0128272
- Selbmann L, Stoppiello GA, Onofri S, Stajich JE, Coleine C. 2021. Culture-dependent and amplicon sequencing approaches reveal diversity and distribution of black fungi in Antarctic Cryptoendolithic Communities. *Journal of Fungi* 7:213
- Hawksworth DL. 2001. The magnitude of fungal diversity: The 1.5 million species estimate revisited. *Mycological Research* 105:1422–1432
- Groenewald M, Lombard L, de Vries M, Lopez AG, Smith M, et al. 2018. Diversity of yeast species from Dutch garden soil and the description of six novel Ascomycetes. *FEMS Yeast Research* 18:0y076
- Valenzuela-Lopez N, Cano-Lira JF, Guarro J, Sutton DA, Wiederhold N, et al. 2018. Coelomycetous *Dothideomycetes* with emphasis on the families *Cucurbitariaceae* and *Didymellaceae*. *Studies in Mycology* 90:1–69
- Hou L, Hernández-Restrepo M, Groenewald JZ, Cai L, Crous PW. 2020. Citizen science project reveals high diversity in *Didymellaceae* (*Pleosporales*, *Dothideomycetes*). *MycoKeys* 65:49–99
- 26. Sun BD, Chen AJ, Houbraken J, Frisvad JC, Wu WP, et al. 2020. New section and species in *Talaromyces*. *MycoKeys* 68:75–113
- Zhang ZF, Zhou SY, Eurwilaichitr L, Ingsriswang S, Raza M, et al. 2021. Culturable mycobiota from Karst caves in China II, with descriptions of 33 new species. *Fungal Diversity* 106:29–136
- Kandemir H Dukik K, de Melo Teixeira M, Stielow JB, Delma FZ, et al. 2022. Phylogenetic and Ecological Reevaluation of the Order Onygenales. Fungal Diversity 115:1–72
- 29. Domsch KH, Gams W, Anderson TH. 1980. *Compendium of Soil Fungi*. Vol 1. London: Academic Press.
- 30. Domsch KH, Gams W, Anderson TH. 1993. *Compendium of Soil Fungi*. Germany: IHW-Verlag Press.
- Domsch KH, Gams W, Anderson TH. 2007. Compendium of Soil Fungi. 2<sup>nd</sup> Edition. Taxono: IHW, Eching.
- 32. Gilman JC. 1968. A Manuel of Soil Fungi. USA: The Iowa State University Press.
- Barron GL, Peterson JL. 1968. The genera of hyphomycetes from soil. Soil Science 106:477
- Bridge P, Spooner B. 2001. Soil fungi: Diversity and detection. *Plant* Soil 232:147–54
- Guarro J, Gené J, Stchigel AM, Figueras MJ. 2012. Atlas of Soil Ascomycetes. (CBS Biodiversity Series No. 10). Netherlands: CBS-KNAW Fungal Biodiversity Centre, Utrecht.
- Chethana KWT, Manawasinghe IS, Hurdeal VG, Bhunjun CS, Appadoo MA, et al. 2021. What Are Fungal Species and How to Delineate Them? *Fungal Diversity* 109:1–25
- Manawasinghe IS, Phillips AJL, Xu J, Balasuriya A, Hyde KD, et al. 2021. Defining a species in fungal plant pathology: Beyond the species level. *Fungal Diversity* 109:267–82
- Jayawardena RS, Hyde KD, de Farias ARG, Bhunjun CS, Ferdinandez HS, et al. 2021. What Is a species in fungal plant pathogens? *Fungal Diversity* 109:239–66
- Bhunjun CS, Niskanen T, Suwannarach N, Wannathes N, Chen YJ, et al. 2022. The numbers of fungi: Are the most speciose genera truly diverse? *Fungal Diversity* 114:387–462

- Jeewon R, Hyde KD. 2016. Establishing species boundaries and new taxa among fungi: Recommendations to resolve taxonomic ambiguities. *Mycosphere* 7:1669–77
- 41. Maharachchikumbura SSN, Chen, Y, Ariyawansa HA, Hyde KD, Haelewaters D, et al. 2021. Integrative approaches for species delimitation in *Ascomycota*. *Fungal Diversity* 109:155–79
- 42. Hyde KD, Xu J, Rapior S, Jeewon R, Lumyong S, et al. 2019. The amazing potential of fungi: 50 ways we can exploit fungi industrially. *Fungal Diversity* 97:1–136
- 43. Anugraha AC, Thomas T. 2021. A review on pigment producing soil fungi and its applications. *Asian Journal of Mycology* 4:89–112
- 44. Wei DP, Gentekaki E, Hyde KD, Xiao YP, Luangharn T, et al. 2022. https://Invertebratefungi.org/: An expert-curated web-based platform for the identification and classification of invertebrate-associated fungi and fungus-like organisms. *Database* 2022:baac021
- 45. Chambergo FS, Valencia EY. 2016. Fungal Biodiversity to Biotechnology. *Applied Microbiology and Biotechnology* 100:2567–2577
- 46. Gutleben J, Chaib De Mares M, van Elsas JD, Smidt H, Overmann J, et al. 2018. The multi-omics promise in context: From sequence to microbial isolate. *Critical Reviews in Microbiology* 4:212–29
- 47. Alshuwaili FE, Al Anbagi RA, Nelsen DJ, Semenova-Nelsen TA, Stephenson SL. 2021. Mycobiome sequencing and analysis of the assemblages of fungi associated with leaf litter on the Fernow Experimental Forest in the Central Appalachian Mountains of West Virginia. Current Research in Environmental & Applied Mycology 11:315–32
- 48. Aritonang R, Mamangkey J, Suryanto D, Munir E, Hartanto A, et al. 2022. Potential of keratinolytic fungi isolated from chicken and goat farms soil, North Sumatra. *Current Research in Environmental & Applied Mycology* 12:15–27
- 49. Iliushin VA, IYu K, Sazanova NA. 2022. Diversity of microfungi of coal mine spoil tips in the Magadan Region, Russia. *Current Research in Environmental & Applied Mycology* 12:136–46
- 50. Gautam AK, Verma RK, Avasthi S, Sushma, Bohra Y, et al. 2022. Current insight into traditional and modern methods in fungal diversity estimates. *Journal of Fungi* 8:226
- Hongsanan S, Hyde KD, Phookamsak R, Wanasinghe DN, McKenzie EHC, et al. 2020. Refined Families of Dothideomycetes: Orders and families incertae sedis in *Dothideomycetes*. *Fungal Diversity* 105:17–318
- Hyde KD, Norphanphoun C, Maharachchikumbura SSN, Bhat DJ, Jones EBG, et al. 2020. Refined Families of Sordariomycetes. *Mycosphere* 11:305–1059

- Wijayawardene NN, Hyde KD, Dai DQ, Sánchez-García M, Goto BT, et al. 2022. Outline of fungi and fungus-like taxa – 2021. *Mycosphere* 13:53–453
- Montoya-Castrillón M, Serna-Vasco KJ, Pinilla L, Quiceno-Rico JM, Cardona-Bermúdez LM, et al. 2021. Isolation and characterization of filamentous fungi from wood and soil samples of "La Lorena", Sonsón, Antioquia (Colombia). DYNA 88:171–180
- Monkai J, Mckenzie E, Phillips A, Hongsanan S, Pem D. et al. 2019. https://fungalgenera.org/: A comprehensive database providing webbased information for all fungal genera. *Asian Journal of Mycology* 2:298–305
- Nilsson RH, Anslan S, Bahram M, Wurzbacher C, Baldrian P, et al. 2019. Mycobiome diversity: High-throughput sequencing and identification of fungi. *Nature Reviews Microbiology* 17:95–109
- 57. Jayasiri SC, Hyde KD, Ariyawansa HA, Bhat J, Buyck B, et al. 2015. The faces of fungi database: Fungal names linked with morphology, phylogeny and human impacts. *Fungal Diversity* 74:3–18
- Pem D, Hongsanan S, Doilom M, Tibpromma S, Wanasinghe DN, et al. 2019. Https://www.dothideomycetes.org: An online taxonomic resource for the classification, identification, and nomenclature of *Dothideomycetes*. *Asian Journal of Mycology* 2:287–97
- Bundhun D, Maharachchikumbura SSN, Jeewon R, Senanayake IC, Jayawardena RS, et al. 2020. https://sordariomycetes.org/, a Platform for the identification, ranking and classification of taxa within Sordariomycetes. Asian Journal of Mycology 3:13–21
- Chaiwan N, Gomdola D, Wang S, Monkai J, Tibpromma S, et al. 2021. https://gmsmicrofungi.org: An online database providing updated information of microfungi in the Greater Mekong Subregion. *Mycosphere* 1:1513–26
- Huanraluek N, Padaruth OD, Jayawardena RS, Li WJ, Hongsanan S, et al. 2021. https://www.coelomycetes.org: Databank that contributes for the classification, identification and nomenclature of Coelomycetes. Asian Journal of Mycology 4:114–22

Copyright: © 2023 by the author(s). Published by Maximum Academic Press, Fayetteville, GA. This article is an open access article distributed under Creative Commons Attribution License (CC BY 4.0), visit https://creativecommons.org/licenses/by/4.0/.