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Exploring the potential of Arbuscular Mycorrhizal Fungi as a biofertilizer

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Abstract

Arbuscular Mycorrhizal Fungi (AMF) are a group of soil-borne fungi that form symbiotic relationships with the roots of most plants, including crops. In this relationship, the fungus provides the plant with nutrients, such as phosphorus, in exchange for carbohydrates produced by the plant through photosynthesis. The use of AMF as a biofertilizer involves the application of these fungi to soil to enhance plant growth and improve nutrient uptake. Studies have shown that AMF can increase plant growth, drought tolerance, and nutrient uptake, leading to improved crop yields. The fungi form a network of hyphae in the soil, which helps to increase the soil's water-holding capacity, as well as its ability to retain nutrients. This can lead to improved plant growth and health, even in nutrient-poor soils. In addition, the use of AMF as a biofertilizer can help to reduce the dependence on synthetic fertilizers, which can have negative environmental impacts. AMF can help to improve soil fertility, increase plant nutrient uptake, and reduce soil erosion, leading to more sustainable agriculture practices. However, it is important to note that the effectiveness of AMF as a biofertilizer can vary depending on several factors, including the species of AMF used, the type of crop being grown, and the conditions of the soil. Additionally, the proper application and management of AMF is important to ensure its effectiveness. In conclusion, the use of AMF as a biofertilizer has the potential to enhance plant growth, improve nutrient uptake, and promote sustainable agriculture practices.

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Introduction

Mycorrhizae are a type of symbiotic relationship that occurs between fungi and plant roots. There are two broad types of mycorrhiza: ectomycorrhizae and endomycorrhizae. Endomycorrhizae are further classified into five different kinds of mycorrhizal associations. Ectendomycorrhizas are another broad category of mycorrhizal association between plant roots and fungi[1-3]. Ectomycorrhizae form a sheath around the roots of certain plants, such as pines, oaks, and eucalyptus. The fungi involved in ectomycorrhizae forms a thick layer around the roots known as fungal sheath or mantle. They also penetrate between the epidermis and cortex through a structure known as the Hartig net. This allows for the exchange of nutrients and water between the plant and the fungus^[4]. Ectomycorrhizae are important for the health and growth of trees in forests, as they help to improve the uptake of nutrients and water from the soil. Endomycorrhizae, on the other hand, form a symbiotic relationship with the plant by actually invading the cells of the root^[4,5]. Endomycorrhizae can be categorized into different types, which are summarized in the following paragraph^[6].

Monotropoid mycorrhiza are characterized by the fact that the fungus is unable to form a visible sheath around the roots of the host plant, and instead forms a diffuse, intraradical mycelium that is intermingled with the plant's own root tissue. The second type of endomycorrhizae are Ericoid mycorrhiza. There is a symbiotic relationship between certain species of ericaceous plants (such as blueberries, rhododendrons, and

heaths) and a group of fungi known as ericoid mycorrhizal fungi (EMF). In this relationship, the EMF colonize the roots of the host plant and form a dense network of fungal structures called hyphae, which increase the plant's ability to absorb water and nutrients, particularly phosphorus. Ericoid mycorrhiza is considered a 'mycoheterotrophic' symbiosis, meaning that the host plant is dependent on the EMF for carbon, which is obtained by the fungus through its association with other plants. The third type are Orchidaceous mycorrhizae which are mutualistic associations between orchid plants and fungi. These relationships are formed in the roots of orchids, where the fungus colonizes the root cells and provide the plant with essential nutrients, such as phosphorus. In return, the orchid provides the fungus with organic carbon through photosynthesis. Orchid mycorrhizae are considered to be obligate, meaning that the orchid cannot survive without the fungus and vice versa. Instead, they rely on the fungus to provide them with this essential nutrient. The most common type of endomycorrhizae, Arbuscular mycorrhiza, are a group of soil-borne, obligate symbionts that form mutualistic associations with the majority of land plants. These fungi colonize plant roots, forming unique structures called arbuscules within the root cells. In exchange for sugars produced by the plant through photosynthesis, the fungi provide the plant with phosphorus and other essential nutrients that are otherwise difficult for the plant to acquire from the soil. AM fungi are ancient symbionts that have evolved alongside plants for over 400 million years, and they play important roles in ecosystem functioning and plant productivity^[7–10]. Various kinds of mycorrhizal associations are described below in Fig. 1.

Soil microorganisms such as AMF reflect a crucial connection between plants and mineral soil nutrients. They are also gaining rising attention as natural fertilisers. AMF are mandatory symbiotics of the phylum Glomeromycota^[12], establishing mutualistic symbiosis, with about 80% of the land plants, including many food crops. In return for photosynthetic materials, they provide the host plant with mineral nutrients and water^[13]. Some benefits provided by AMF in plant growth and yield are depicted in Fig. 2.

AMF mycelium from the root system is able to extract nutrients from soil volumes that are inaccessible to the roots^[14]. In comparison to plant roots, fungal hyphae are far thinner and can reach narrower pores^[15]. AM fungal hyphae colonise root cortex predominantly by forming profusely branchy structures inside the cells, i.e. arbuscules, which are known as the functional nutrient exchange sites^[16]. AMF therefore eliminates plant growth restrictions imposed by an insufficient supply of nutrients^[17,18]. In recent times a non-mycorrhizal state can be regarded as rare in natural habitats for most organisms^[19], even though the AM fungal populations below the ground are

significantly different depending on the species composition, soil and seasonal form or variation of these factors^[19]. AM association provide plants with additional advantages, in addition to an increased food source, such as enhanced drought and salinity resistance^[20,21]. While several studies have been performed on the impact of AM symbiosis on plant reaction to abiotic stress such as drought, salinity and flooding in the last few years, the processes that have contributed to an improved plant stress resistance still remains somewhat elusive^[20,22–25]. Metals such as Iron (Fe), Copper (Cu) and Zinc (Zn) perform important functions in a variety of sub-cellular compartments, but they constitute a highly reactive community of elements that are toxic at large concentrations^[26].

AMF have been documented to minimise the toxicity of heavy metals in host plant and to withstand high metal concentrations in soil^[26–30]. Metal transporters play a vital function in homeostasis of heavy metals. A Zn transporter was identified in *Glomus intraradices* (GintZnT1)^[31] and multiple putative genes coding Cu, Fe and Zn transporters were identified in a genome-wide study of the recently published *Rhizophagus irregularis* (formally, *Glomus intraradices*) genome^[32]. The next steps would be characterisation of these carriers and discerning

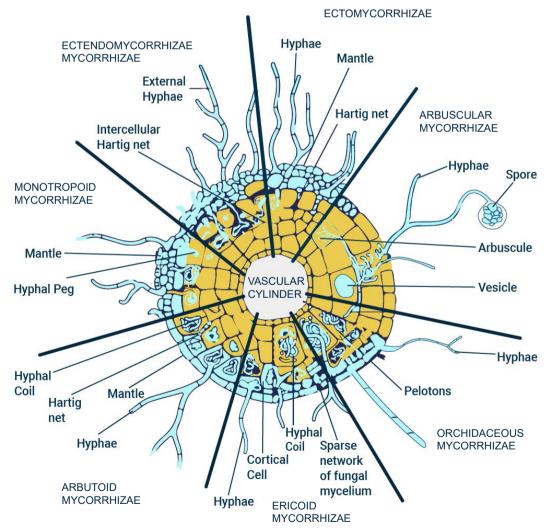


Fig. 1 Different types of mycorrhizal symbiosis in nature: Ectomycorrhizae, Monotropoid mycorrhizae, Arbutoid mycorrhizae, Ericoid mycorrhizae, Orchidaceous mycorrhizae, and Arbuscular mycorrhizae. (Modified image from Selosse & Le Tacon^[111])

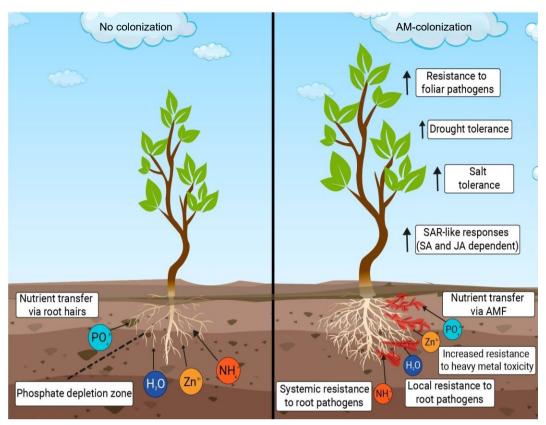


Fig. 2 A pictorial representation of a few of the benefits provided by AMF in plant growth and yield. Increased resistance to foliar pathogens, increased drought tolerance, increased salt tolerance, enhanced defense mechanism, help in uptake of essential nutrients and bioremediation are a few of the many benefits that AMF provide in the growth and development of a plant.

their role in the symbiosis. AMF may also have a significant influence on the environment, as they promote soil structure and aggregation^[33–36] and control the development and production of plant populations^[37].

The impact of AMF symbiosis has also recently been studied on greenhouse gas (GHG) emission^[38,39]. Evidence presented by Bender et al. suggests that AMF may have a role to play in climate change mitigation due to their ability to significantly reduce emissions of N₂O, a key greenhouse gas. By enhancing plant nitrogen (N) absorption and assimilation, AMF may be able to reduce N₂O emissions by decreasing soluble N in soil and, in turn, denitrification[38]. Correlations between AMF abundance and genes involved in primary N₂O production (nirK) and consumption (nosZ) suggest that AMF promote shifts in soil microbial biomass and community composition that lead to reduced N₂O emissions. According to Lazcano et al., AM symbiosis aids in N₂O emission management at high soil moisture levels, and it was suggested that AM plant N₂O emission control may be mediated by higher soil water use rather than increased N absorption^[39].

Therefore, AMF are primary biotic soil elements, which, when absent or degraded, for instance, by anthropic input, will contribute to a less effective functioning of the ecosystem. The process of re-establishing AMF may be a promising solution to industrial fertilisation methods in order to achieve organic cultivation, a significant goal for farmers in the midst of a global recession and an environmentally friendly consumer^[40,41]. The key technique for achieving this aim is to directly reintroduce AMF (inoculum) propagules in the target soil. However, in the

application of these fungi, awareness of how AMF adapts and reacts to the objective of soil management and ecosystem management and the events which result in a functional symbiosis, including the mechanisms involved in the transfer of nutrients is important^[42]. This paper gives a brief discussion of the latest studies on the nutritional aspects of AM symbiosis and a short description of the challenges of development of AMF inoculum, descriptions of the application of AM fungi are mentioned and addressed, both under regulated and open field conditions, with specific emphasis on identifying factors contributing to success of the biofertilizer.

Essential features of AMF symbiosis

The symbiosis of AMF with plants was originally discovered 400 million years ago^[43]. Such links are a series of biological processes that have beneficial impacts on the ecosystems of both wild and cultivated biota^[44]. An example of a reciprocal interaction that may regulate the growth and advancement of a plant is the symbiotic relationship between AMF and its partners. The plant's roots may absorb nutrients that would not otherwise be available thanks to the mycelial fungal network that has grown throughout them^[45]. The fungal mycelium colonises the roots of several plants, even if they are from different species, and creates a shared mycorrhizal network. According to Pringle et al.^[46], this mycorrhizal network is regarded as the main element of the terrestrial ecosystem for many plant populations, including invasive species, and it facilitates the transfer of phosphorus (P) and nitrogen (N) to plants

via fungi[13,47]. They may improve soil qualities and encourage plant development in both natural and challenging environments^[48–50]. Plant resilience to harsh environments is increased by AMF colonisation, which results in several improvements in morpho-physiological traits^[49–52]. Researchers encourage the use of AMF as influential bio-fertilizers in sustainable agricultural production, having been employed as bioinoculants^[53]. In comparison to untreated soils, AMF-inoculated soil often forms more consistent masses and much more extraradical hyphal mycelium^[54]. Glomalin-related soil protein is thought to keep soils with complex abiotic stressors moist^[55], which then controls water levels between soil and plants and naturally promotes plant development. Glomalin contains 30%-40% carbon (C) and related chemicals to protect soil from desiccation by enhancing its ability to retain soil-water^[56]. Growthrelated processes that affect AMF inoculation include stomatal conductance, leaf water capacity, relative water content (RWC), PSII quality, and CO₂ assimilation^[57,58]. By altering the biochemistry of the organ and tissues above ground, AMF also contributes to improving water stress tolerance^[24]. Additionally, AMF inoculation promotes the accumulation of dry matter and improves moisture uptake, boosting plant tolerance to stressors like salt and drought. AMF extraction for plant growth in various biological conditions will significantly boost organic farming's ability to promote growth and raise production[59].

Here are the key characteristics of AM fungus symbiosis:

- a) Establishment: The AM fungus forms a symbiotic relationship with plant roots through the process of colonization, in which the fungus penetrates the root cells and forms structures known as arbuscules^[60].
- b) Nutrient exchange: The AM fungus enhances the plant's nutrient uptake by extending the absorptive surface area of the root system and solubilizing soil nutrients. In return, the plant provides the AM fungus with carbon^[61,62].
- c) Soil structure improvement: AM fungi can contribute to soil structure improvement through the production of extraradical mycelium, which helps bind soil particles and improve soil stability^[40].
- d) Enhanced plant growth: The symbiotic relationship between AM fungi and plants results in improved plant growth

and health, with higher root and shoot biomass, enhanced root system architecture, and increased stress tolerance^[45].

- e) Wider host range: AM fungi have a wide host range, forming symbiotic relationships with many different plant species, including some of the most important crops such as maize, wheat, and soybean $^{[4,63-66]}$.
- f) Environmental importance: AM fungi play a crucial role in the ecosystem, by improving soil fertility and plant productivity. They also help to maintain soil health and reduce soil degradation by contributing to soil aggregation and water retention. Overall, the AM fungus symbiosis is a vital component of the terrestrial ecosystem, providing many benefits to plants and the environment^[67,68].

General life-cycle of an AMF

The life-cycle of AMF is basically divided in two stages; presymbiosis and symbiosis (Fig. 3). Pre-symbiosis is characterized by the following steps: Plant root exudating certain chemicals, spore germination and hyphal growth, and attachment of appressorium to the root. This step is followed by the symbiosis stage: the hyphae of the AMF colonize the root system of a host plant by penetrating the root cells and form arbuscules, and vesicles^[69,70]. The hyphae of the AMF extend into the soil and increase the absorptive surface area of the root system (Fig. 4). The relationship between the AMF and the host plant becomes established, and the fungus continues to grow and spread throughout the root system. Extraradical mycelium also gives rise to formation of chlamydospores on soils at maturity. When the host plant dies, the AMF begins to decompose the plant's organic matter and return nutrients to the soil. The AMF spores, in time will make association with other plants and continue its life cycle.

Methods and source of AMF inoculum propagation

Reviewing the work from the past few decades, the most successful form of AMF propagation was the use of trap cultures using certain plants as a host species, and it accounted

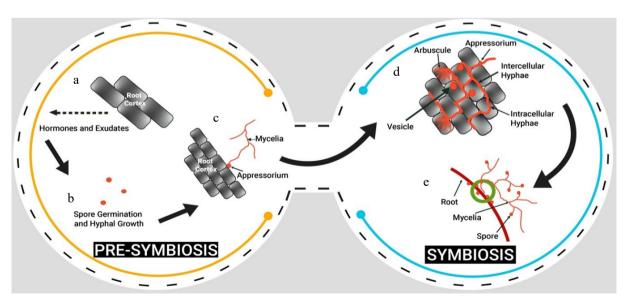


Fig. 3 Life cycle of AMF showing pre-symbiosis and symbiosis stages.

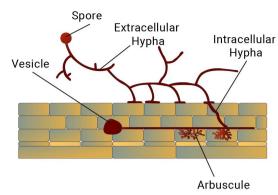


Fig. 4 Fig. 4- Graphical representation of AMF infection in the root. AMF colonisation is characterised by the formation of arbuscules, vesicles, hyphae and spores.

for almost 75% of AMF propagation and interestingly, only marginalised usage of other approaches was observed. Approaches such as aeroponics and hydroponics, are also contributing to the development of pure clean spores and the maximisation of the host plant growth conditions^[71]. The monoxenic root organ culture is another approach that permits the effective large-scale propagation of AMF that can be directly used as inoculum. The system consists of culture of inoculated excised roots (the so-called hairy roots), which have acquired the capacity to proliferate without developing any epigeous portion, after processing with the soil-borne plasmid Ri (root-inducing) Agrobacterium rhizogenes^[72]. Within a few months, a significant number of spores, mycelium and colonised roots were produced^[73]. Since, AMF may use a variety of propagules in order to expand and colonise new roots with different levels of efficiency^[74], the selection of the source of the inoculum is a factor of primary importance for a successful colonisation. Spores, mycelium fragments fragmented from the lower hyphal network and other complexes inside both living and dead root fragments are all components of the extraradical and intraradical systems of AMF. The main cause of regrowth for some AM fungal organisms was in specific intraradical vesicles^[75]. Different AM fungal taxonomic ranks vary in their capacity to disperse from a given propagule. The propagation of mycelial fragmentation appears to be of greater importance for organisms of the Glomeraceae family, whereas for representatives of other groups such as Gigasporaceae, Acaulosporaceae and Scutellosporaceae spore germination would be preferential methods of propagation^[76]. The most effective and user-friendly method to apply a multi species inoculum, as propagation through trap culture is the most widely utilised strategy, is to sieve the substrate and finely cut the root of the trap culture plant so that all the various kinds of fungal propagules (crude inoculum) can be retrieved.

Inoculum structure

In most of the reviews studied, the latest general tendency is to pursue one or more types of AM fungi for individual inoculation (monospecies inoculum). Gosling et al.^[77] have examined the impact on plant growth after inoculating diverse AM fungal populations with functionally distinct characteristics, fewer fungal species that are able to mitigate stress are likely to be of utmost benefit to the hosts when a host plant is subjected to one cause, such as greenhouse experiments. In the case of

shoot biomass, single species inoculation experiments are more effective than inoculation experiments with more than one species concurrently used. Another greenhouse research has shown that species composition instead of variety may be more critical in deciding how the species works^[78].

According to Opik et al.[79], for single species inoculation, three AMF species viz. Rhizophagus intraradices, Rhizophagus irregularis and Funneliformis mossae are subjected to most scientific studies because these are extremely versatile symbionts that can colonise a wide range of host plants, maintain long-term storage, disperse widely across the world and quickly and massively reproduce. These species have been ideal for premium inoculum components due to the above described characteristics. Several studies have shown that different isolates from the same species can have a greater impact on plant response compared to the variations among different species^[80-82]. This suggests that the exclusive use of individual AMF organisms such as R. intraradices, R. irregularis and F. mossae should not be considered a drawback in inoculation experiments as these species may have significant functional diversity. In this context, in the presence of R. irregularis reference genome^[32,83] the partial genome re-sequencing of several isolates from various geographical backgrounds can open the door to exploring the roles of genetic variation in AMF communities so that it is possible to develop and choose more successful and productive AMF for crop plants. Another factor that needs to be addressed is that the receptivity of plant organisms, including seeds, to AMF inoculation differs greatly[84-86]. The plants' reaction to AM fungi can be used as a selection function in modern farming, resulting in varieties or cultivars with different genetic differences.

AMF as a bio-fertilizer

Bio-fertilizers are a combination of naturally occuring microbes used to increase soil fertility. These fertilisers are very useful for soil health and plant growth[87]. Various scientific experiments on AMF in the last two decades have demonstrated their innumerable benefits in terms of soil quality and crop productivity. Therefore, it is commonly assumed that AMF may, in the foreseeable future, be seen as a substitute for inorganic fertilisers, since mycorrhizal applications will effectively reduce the quantitative use of the chemical fertiliser input, especially phosphorus^[88]. Owing to the adverse influence on food safety, crop health, and air and water systems by inorganic fertilisers, herbicides and fungicides, the continued usage of these have triggered numerous land, plant and human health concerns^[89]. AMF may be able to minimise chemical fertiliser usage up to 50% for optimal agricultural output, although this calculation depends on the variety of plant species and the prevalence of stressful environments. In order for sustainable agriculture to be accomplished, AMF as a biofertilizer becomes more significant because the proper treatment of these symbiotic fungi might significantly minimise the usage of agrochemicals. Inoculation of AMF propagules (inoculum) into a target soil is the key technique embraced for this aim. Sadly, AMF are obligate symbionts and cannot be produced without the host plants in pure cultures. The large-scale development of AMF inocula is very difficult and complex because of this constraining feature.

There are three major forms of AMF inocula. First, AMF soil may be used as an inoculum from the root zone of a plant since

it usually includes colonised root parts, AMF spores, and hyphae. However, without adequate knowledge regarding propagule quantity, variety and infectivity, soil inocula cannot be effective and may be at risk of transmitting weed seeds and pathogenic agents. Spores removed from soil can be used instead as starters for the development of crude inoculum. Crude inoculum can be collected from the inert medium adapted for AMF propagation after the known AMF isolate and the host trap plant (i.e. plant which can be colonised with a lot of AMF species) have been cultivated together^[90–92]. This is the most common form of inoculum used for large-scale inoculation, since it normally includes a more condensed collection of propagules of the same sort present in the soil inoculum. Finally, colonised root fragments alone from an established AMF host that are isolated from a trap crop may even be used as an inoculum source. AMF's large-scale development of crude inoculum remains very demanding even while new mass processing methods^[71], and the technology of seed coating have been developed in recent years^[44]. The biggest challenge to an AMF inoculum is the unavoidable symbiotic conduct of the AMF, i.e., its requirement for a host facility to expand and complete its life cycles. Which implies that the propagation stage must involve a time and space-demanding process of cultivation with the host plant. As a result, the establishment of AMF reference collections involves methodologies which are very different and more binding than those for other microbial collections^[93–96]. In addition, the lack of a prompt way to determine when and how often the host plant is colonised by AMF often contributes to threatening AMF 's agricultural usability. The management of the high inoculum needed for large-scale usage is also a challenging operation. However, AMF inoculation for plant manufacturing systems with a transplant stage is simpler since smaller quantities of inoculum are required.

Once AMF biodiversity has been preserved and well developed, and an AMF-friendly management such as fall cover^[97] and conservation tillage^[98] is introduced, the AMF population will prosper and if no damage is done before and after cultivation, it is understood that in the future, the network of mycorrhizal hyperbiotics of biodiversity will remain unaltered and contagious. Few of the experiments carried out to study the effectiveness of AMF as a biofertilizer either alone or in combination of other soil microorganisms are as follows: The maximum grain production of Triticum aestivum at phosphorus levels was recorded on being inoculated with Pseudomonas striata, followed by Glomus fasciculatum[99]. Rajendran & Jayasree[100] have examined the impact of biofertilizers such as Rhizobium, AMF etc. for Acacia nilotica and also demonstrated a substantial improvement in the length of seedlings and biomass compared with control. Microbial bioagents have been explored in chickpea for the prevention of collar rot disease^[101]. When they co-inoculated Rhizobium with AMF, optimum decrease in mortality was observed as compared to control (100% seedling mortality). Different yield parameters have been effectively improved in each of the adjustments by disease control and seedling mortality reduction. The impact of single and dual inoculum of AMF species such as that of Gigaspora rosea, Glomus intraradices + Gigaspora rosea, and Glomus etunicatum + Glomus intraradices on dry shoot and root weight of Medicago sativa plants was studied by Khan et al.[102] and they reported a significant growth in both the parameters compared to control. Increase in nutrient absorption in inoculated plants was also reported. The inoculation of AMF is promising because it is inexpensive, simple to manage and promotes the growth of plants and the seed quality. Growth and return reaction to inoculation of *Cicer arietinum* with *Rhizobium* sp. and AMF has been studied by Giri^[103]. These findings showed an increase of 10.83% of the total weight and 9.0% of the germination over control in pot experiments.

Conclusions

According to some estimates, the global population will reach 9 billion by the year 2050^[104]. In order to preserve both human and environmental health, the world's agricultural sector is now faced with the problem of almost tripling the amount of food that is produced while simultaneously lowering farmers' dependency on agricultural chemicals. The increase in yield that is anticipated is more than the current capacity for the production of food throughout the world[104], which highlights the need for the development or revitalization of environmentally friendly technologies such as AMFbased biofertilization. Despite the enormous potential it has, agriculturalists have not yet entirely accepted the use of AMF. According to the findings of this study, overall AMF inoculation has beneficial effects on plant growth in both controlled and open-field conditions. This is mostly due to the many nutritional advantages that this class of soil fungus symbionts provides to the host plant. In point of fact, it has been shown that AMF inoculation in the field is just as effective as inoculation in the greenhouse, where, in contrast to open field conditions, non-inoculated controls are often free of AMF. Because of this, the next significant step for the consistent use of AMF in agriculture is to conduct large-scale field experiments and a cost-benefit study, such as the one suggested by Ceballos et al.[105], to enable future end-users to be better aware of the benefits of AMF inoculant. Farmers are urged to create their own AMF inoculant from their local soils since research has shown that the indigenous AMF is as potent as or stronger than commercial or cultural isolates. Even farmers in developing nations like India, who are in desperate need of a technique of crop production that is sustainable, would have an easier time gaining access to biofertilization technology as a result of this.

Author contributions

The authors confirm their contribution to the paper as follows: study conception and design: Jha SS, Songachan LS. Data collection: Jha SS; analysis and interpretation of results: Jha SS, Songachan LS. All authors reviewed the results and approved the final version of the manuscript

Data availability

The data supporting the findings of the study are available within this published article.

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Conflict of interest

The authors declare that they have no conflict of interest.

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References

- Harley JL, Smith SE. 1983. Mycorrhizal Symbiosis. London: Academic Press.
- Smith SE, Read DJ. 1997. Mycorrhizal symbiosis. 2nd Edition. San Diego: Academic Press. https://doi.org/10.1016/B978-0-12-652840-4.X5000-1
- 3. Read DJ. 1999. Mycorrhiza the state of the art. In *Mycorrhiza:* structure, function, molecular biology and biotechnology, eds.Varma A, Hock B. 2nd Edition. Berlin, Heidelberg: Springer. pp. 3–34. https://doi.org/10.1007/978-3-662-03779-9_1
- Trappe JM. 1987. Phylogenetic and ecological aspects of mycotrophy in the angiosperms from an evolutionary standpoint. In *Ecophysiology of VA mycorrhizal plants*, ed. Safir GR. Boca Raton: CRC Press. pp. 5–26.
- Harley JL, Harley EL. 1987. A check-list of mycorrhiza in the British flora—addenda, errata and index. New Phytologist 107:741–49
- Morton JB. 1993. Problems and solutions for the integration of glomalean taxonomy, systematic biology, and the study of endomycorrhizal phenomena. Mycorrhiza 2:97–109
- Janos DP. 1980. Vesicular-arbuscular mycorrhizae affect lowland tropical rain forest plant growth. *Ecology* 61:151–62
- Zhao ZW, Xia YM, Qin XZ, Li XW, Cheng LZ, et al. 2001. Arbuscular mycorrhizal status of plants and the spore density of arbuscular mycorrhizal fungi in the tropical rain forest of Xishuangbanna, southwest China. Mycorrhiza 11:159–62
- Haselwandter K. 1987. Mycorrhizal infection and its possible ecological significance in climatically and nutritionally stressed alpine plant communities. Angewandte Botanik 61:107–14
- Muthukumar T, Udaiyan K. 2006. Growth of nursery-grown bamboo inoculated with arbuscular mycorrhizal fungi and plant growth promoting rhizobacteria in two tropical soil types with and without fertilizer application. New Forests 31:469–85
- Selosse MA, Le Tacon F. 1998. The land flora: a phototrophfungus partnership? *Trends in Ecology & Evolution* 13:15–20
- Schüβler A, Schwarzott D, Walker C. 2001. A new fungal Phylum, the *Glomeromycota*: phylogeny and evolution. *Mycological Research* 105:1413–21
- Smith SE, Read DJ. 2008. Mycorrhizal Symbiosis. 3rd Edition. London: Academic Press. https://doi.org/10.1016/B978-0-12-370526-6.X5001-6
- Smith FA, Jakobsen I, Smith SE. 2000. Spatial differences in acquisition of soil phosphate between two arbuscular mycorrhizal fungi in symbiosis with *Medicago truncatula*. New Phytologist 147:357–66
- Allen MF. 2011. Linking water and nutrients through the vadose zone: a fungal interface between the soil and plant systems. *Journal of Arid Land* 3:155–63
- Balestrini R, Lumini E, Borriello R, Bianciotto V. 2015. Plant-soil biota interactions. In Soil Microbiology, Ecology and Biochemistry, ed. Paul EA. Amsterdam: Academic Press, Elsevier. pp. 311–38. https://doi.org/10.1016/b978-0-12-415955-6.00011-6
- Nouri E, Breuillin-Sessoms F, Feller U, Reinhardt D. 2014. Phosphorus and nitrogen regulate arbuscular mycorrhizal symbiosis in *Petunia* hybrida. *PLoS One* 9:e90841
- Lewe N. 2023. The interplay between host plants and priority effects in community assembly processes of arbuscular mycorrhizal fungi.

- Doctoral dissertation. Victoria University of Wellington Library. https://doi.org/10.26686/wqtn.21974357
- Smith SE, Smith FA. 2012. Fresh perspectives on the roles of arbuscular mycorrhizal fungi in plant nutrition and growth. Mycologia 104:1–13
- Augé RM. 2001. Water relations, drought and vesicular-arbuscular mycorrhizal symbiosis. Mycorrhiza 11:3–42
- Porcel R, Aroca R, Ruiz-Lozano JM. 2012. Salinity stress alleviation using arbuscular mycorrhizal fungi. A review. Agronomy for Sustainable Development 32:181–200
- Ruiz-Lozano JM. 2003. Arbuscular mycorrhizal symbiosis and alleviation of osmotic stress. New perspectives for molecular studies. Mycorrhiza 13:309–17
- Ruiz-Lozano JM, Aroca R. 2010. Modulation of aquaporin genes by the arbuscular mycorrhizal symbiosis in relation to osmotic stress tolerance. In *Symbioses and stress*, eds. Seckbach J, Grube M. Vol. 17. Dordrecht: Springer. pp. 357–74. https://doi.org/ 10.1007/978-90-481-9449-0_17
- Bárzana G, Aroca R, Paz JA, Chaumont F, Martinez-Ballesta MC, et al. 2012. Arbuscular mycorrhizal symbiosis increases relative apoplastic water flow in roots of the host plant under both wellwatered and drought stress conditions. Annals of Botany 109:1009–17
- Bárzana G, Aroca R, Ruiz-lozano JM. 2015. Localized and nonlocalized effects of arbuscular mycorrhizal symbiosis on accumulation of osmolytes and aquaporins and on antioxidant systems in maize plants subjected to total or partial root drying. *Plant, Cell* & Environment 38:1613–27
- Tamayo E, Gómez-Gallego T, Azcón-Aguilar C, Ferrol N. 2014. Genome-wide analysis of copper, iron and zinc transporters in the arbuscular mycorrhizal fungus *Rhizophagus irregularis*. Frontiers in Plant Science 5:547
- Göhre V, Paszkowski U. 2006. Contribution of the arbuscular mycorrhizal symbiosis to heavy metal phytoremediation. *Planta* 223:1115–22
- Lingua G, Franchin C, Todeschini V, Castiglione S, Biondi S, et al. 2008. Arbuscular mycorrhizal fungi differentially affect the response to high zinc concentrations of two registered poplar clones. Environmental Pollution 153:137–47
- Cornejo P, Pérez-Tienda J, Meier S, Valderas A, Borie F, et al. 2013.
 Copper compartmentalization in spores as a survival strategy of arbuscular mycorrhizal fungi in Cu-polluted environments. Soil Biology and Biochemistry 57:925–28
- Meier S, Cornejo P, Cartes P, Borie F, Medina J, et al. 2015. Interactive effect between Cu-adapted arbuscular mycorrhizal fungi and biotreated agrowaste residue to improve the nutritional status of Oenothera picensis growing in Cu-polluted soils. Journal of Plant Nutrition and Soil Science 178:126–35
- González-Guerrero M, Azcón-Aguilar C, Mooney M, Valderas A, MacDiarmid CW, et al. 2005. Characterization of a Glomus intraradices gene encoding a putative Zn transporter of the cation diffusion facilitator family. Fungal Genetics and Biology 42:130–40
- 32. Tisserant E, Malbreil M, Kuo A, Kohler A, Symeonidi A, et al. 2013. Genome of an arbuscular mycorrhizal fungus provides insight into the oldest plant symbiosis. Proceedings of the National Academy of Sciences of the United States of America 110:20117–22
- Rillig MC, Mummey DL. 2006. Mycorrhizas and soil structure. New Phytologist 171:41–53
- Leifheit EF, Veresoglou SD, Lehmann A, Morris EK, Rillig MC. 2014.
 Multiple factors influence the role of arbuscular mycorrhizal fungi in soil aggregation—a meta-analysis. *Plant and Soil* 374:523–37
- Leifheit EF, Verbruggen E, Rillig MC. 2015. Arbuscular mycorrhizal fungi reduce decomposition of woody plant litter while increasing soil aggregation. Soil Biology and Biochemistry 81:323–28
- Rillig MC, Aguilar-Trigueros CA, Bergmann J, Verbruggen E, Veresoglou SD, et al. 2015. Plant root and mycorrhizal fungal

- traits for understanding soil aggregation. *New Phytologist* 205:1385–88
- van der Heijden MGA, Klironomos JN, Ursic M, Moutoglis P, Streitwolf-Engel R, et al. 1998. Mycorrhizal fungal diversity determines plant biodiversity, ecosystem variability and productivity. Nature 396:69–72
- 38. Bender SF, Plantenga F, Neftel A, Jocher M, Oberholzer HR, et al. 2014. Symbiotic relationships between soil fungi and plants reduce N₂O emissions from soil. *The ISME Journal* 8:1336–45
- Lazcano C, Barrios-Masias FH, Jackson LE. 2014. Arbuscular mycorrhizal effects on plant water relations and soil greenhouse gas emissions under changing moisture regimes. Soil Biology and Biochemistry 74:184–92
- Miller RM, Jastrow JD. 2000. Mycorrhizal fungi influence soil structure. In *Arbuscular Mycorrhizas: Physiology and Function*, eds. Kapulnik Y, Douds DD. Dordrecht: Springer. pp. 3–18. https://doi.org/10.1007/978-94-017-0776-3_1
- Wright SF, Franke-Snyder M, Morton JB, Upadhyaya A. 1996. Time-course study and partial characterization of a protein on hyphae of arbuscular mycorrhizal fungi during active colonization of roots. *Plant and Soil* 181:193–203
- Kuila D, Ghosh S. 2022. Aspects, problems and utilization of Arbuscular Mycorrhizal (AM) application as bio-fertilizer in sustainable agriculture. Current Research in Microbial Sciences 3:100107
- Selosse MA, Strullu-Derrien C, Martin FM, Kamoun S, Kenrick P. 2015. Plants, fungi and oomycetes: a 400-million year affair that shapes the biosphere. New Phytologist 206:501–6
- van der Heijden MGA, Martin FM, Selosse MA, Sanders IR. 2015. Mycorrhizal ecology and evolution: the past, the present, and the future. New Phytologist 205:1406–23
- Jha SS, Songachan LS. 2020. Research on diversity and community composition of arbuscular mycorrhizal fungi species in India: a review. *Plant Archives* 20(2):4201–26
- Pringle A, Bever JD, Gardes M, Parrent JL, Rillig MC, et al. 2009. Mycorrhizal symbioses and plant invasions. *Annual Review of Ecology, Evolution, and Systematics* 40:699–715
- Jha SS, Songachan LS. 2022. Mycorrhizoremediation: understanding the science behind it and it's future prospects. *Materials Today: Proceedings* 51:2431–36
- Navarro JM, Pérez-Tornero O, Morte A. 2014. Alleviation of salt stress in citrus seedlings inoculated with arbuscular mycorrhizal fungi depends on the rootstock salt tolerance. *Journal of Plant Physiology* 171:76–85
- Alqarawi AA, Abd Allah EF, Hashem A. 2014. Alleviation of saltinduced adverse impact via mycorrhizal fungi in Ephedra aphylla Forssk. Journal of Plant Interactions 9:802–10
- Alqarawi AA, Hashem A, Abd_Allah EF, Alshahrani TS, Huqail AA.
 2014. Effect of salinity on moisture content, pigment system, and lipid composition in *Ephedra alata* decne. *Acta Biologica Hungarica* 65:61–71
- Hashem A, Abd_Allah EF, Alqarawi AA, Aldubise A, Egamberdieva D. 2015. Arbuscular mycorrhizal fungi enhances salinity tolerance of *Panicum turgidum* Forssk by altering photosynthetic and antioxidant pathways. *Journal of Plant Interactions* 10:230–42
- Basit A, Shah ST, Ullah I, Muntha ST, Mohamed HI. 2021. Microbeassisted phytoremediation of environmental pollutants and energy recycling in sustainable agriculture. Archives of Microbiology 203:5859–85
- Barrow CJ. 2012. Biochar: potential for countering land degradation and for improving agriculture. Applied Geography 34:21–28
- Syamsiyah J, Herawati A, Mujiyo. 2018. The potential of arbuscular mycorrhizal fungi application on aggregrate stability in alfisol soil. IOP Conference Series: Earth and Environmental Science 142:012045

- Wu Z, McGrouther K, Huang J, Wu P, Wu W, et al. 2014. Decomposition and the contribution of glomalin-related soil protein (GRSP) in heavy metal sequestration: field experiment. Soil Biology and Biochemistry 68:283–90
- Sharma S, Prasad R, Varma A, Sharma AK. 2017. Glycoprotein associated with Funneliformis coronatum, Gigaspora margarita and Acaulospora scrobiculata suppress the plant pathogens in vitro. Asian Journal of Plant Pathology 11:199–202
- He F, Sheng M, Tang M. 2017. Effects of Rhizophagus irregularis on photosynthesis and antioxidative enzymatic system in Robinia pseudoacacia L. under drought stress. Frontiers in Plant Science 8:183
- Chandrasekaran M, Chanratana M, Kim K, Seshadri S, Sa T. 2019. Impact of arbuscular mycorrhizal fungi on photosynthesis, water status, and gas exchange of plants under salt stress – a metaanalysis. Frontiers in Plant Science 10(10):457
- Lahlali R, Ezrari S, Radouane N, Kenfaoui J, Esmaeel Q, et al. 2022.
 Biological control of plant pathogens: a global perspective.
 Microorganisms 10:596
- Jeffries P, Gianinazzi S, Perotto S, Turnau K, Barea JM. 2003. The contribution of arbuscular mycorrhizal fungi in sustainable maintenance of plant health and soil fertility. *Biology and Fertility of* Soils 37:1–16
- Fracchia S, Sampedro I, Scervino J, Garcia-Romera I, Ocampo J, et al. 2004. Influence of saprobe fungi and their exudates on arbuscular mycorrhizal symbioses. Symbiosis 36(2):169–82
- Rambelli A. 1973. The rhizosphere of mycorrhizae. In Ectomyhyphencorrhizae, eds. Marks GC, Kozlowski TT. New York: Academic Press. pp. 299–349. https://doi.org/10.1016/b978-0-12-472850-9.50014-1
- 63. Rajkumar HG, Seema HS, Sunil Kumar CP. 2012. Diversity of arbuscular mycorrhizal fungi associated with some medicinal plants in Western Ghats of *Karnataka* region, India. *World Journal of Nuclear Science and Technology* 2:13–20
- 64. Gemma JN, Koske RE, Flynn T. 1992. Mycorrhizae in Hawaiian pteridophytes: occurrence and evolutionary significance. American Journal of Botany 79:843–52
- 65. Muthukumar T, Sathiyaraj G, Priyadharsini P, Uma E, Sathiyadash K. 2014. Arbuscular mycorrhizal and dark septate endophyte fungal associations in ferns and lycophytes of Palni Hills, Western Ghats, southern India. *Brazilian Journal of Botany* 37:561–81
- 66. Santhosh Kumar S, Nagarajan N. 2014. AM fungal association in the Rhizosphere soil of some Pteridophytic plant species in Valparai Hills, Western Ghats of Tamilnadu, India. *International Journal of Life Sciences* 2:201–6
- Vosátka M, Gryndler M. 1999. Treatment with culture fractions from *Pseudomonas putida* modifies the development of *Glomus* fistulosum mycorrhiza and the response of potato and maize plants to inoculation. *Applied Soil Ecology* 11:245–51
- Gryndler M. 2000. Interactions of arbuscular mycorrhizal fungi with other soil organisms. In *Arbuscular Mycorrhizas: Physiology* and Function, eds. Kapulnik Y, Douds DD. Dordrecht: Springer. pp. 239–62. https://doi.org/10.1007/978-94-017-0776-3_11
- Giovannetti M, Sbrana C, Logi C. 1994. Early processes involved in host recognition by arbuscular mycorrhizal fungi. New Phytologist 127:703–9
- Hart MM, Reader RJ. 2004. Do arbuscular mycorrhizal fungi recover from soil disturbance differently? *Tropical Ecology* 45:97–111
- IJdo M, Cranenbrouck S, Declerck S. 2011. Methods for large-scale production of AM fungi: past, present, and future. *Mycorrhiza* 21:1–16
- Bécard G, Fortin JA. 1988. Early events of vesicular-arbuscular mycorrhiza formation on Ri T-DNA transformed roots. New Phytologist 108:211–18
- Declerck S, Strullu DG, Plenchette C. 1998. Monoxenic culture of the intraradical forms of Glomus sp. isolated from a tropical

- ecosystem: a proposed methodology for germplasm collection. *Mycologia* 90:579–85
- Klironomos JN, Hart MM. 2002. Colonization of roots by arbuscular mycorrhizal fungi using different sources of inoculum. Mycorrhiza 12:181–84
- Biermann B, Linderman RG. 1983. Use of vesicular-arbuscular mycorrhizal roots, intraradical vesicles and extraradical vesicles as inoculum. New Phytologist 95:97–105
- Brundrett MC, Jasper DA, Ashwath N. 1999. Glomalean mycorrhizal fungi from tropical Australia. Mycorrhiza 8:315–21
- Gosling P, Jones J, Bending GD. 2016. Evidence for functional redundancy in arbuscular mycorrhizal fungi and implications for agroecosystem management. Mycorrhiza 26:77–83
- Wagg C, Barendregt C, Jansa J, van der Heijden MGA. 2015. Complementarity in both plant and mycorrhizal fungal communities are not necessarily increased by diversity in the other. Journal of Ecology 103:1233–44
- Öpik M, Vanatoa A, Vanatoa E, Moora M, Davison J, et al. 2010. The online database MaarjAM reveals global and ecosystemic distribution patterns in arbuscular mycorrhizal fungi (Glomeromycota). New Phytologist 188:223–41
- Munkvold L, Kjøller R, Vestberg M, Rosendahl S, Jakobsen I. 2004.
 High functional diversity within species of arbuscular mycorrhizal fungi. New Phytologist 164:357

 –64
- 81. Gai JP, Feng G, Christie P, Li XL. 2006. Screening of arbuscular mycorrhizal fungi for symbiotic efficiency with sweet potato. *Journal of Plant Nutrition* 29:1085–94
- Angelard C, Colard A, Niculita-Hirzel H, Croll D, Sanders IR. 2010.
 Segregation in a mycorrhizal fungus alters rice growth and symbiosis-specific gene transcription. Current Biology 20:1216–21
- Lin K, Limpens E, Zhang Z, Ivanov S, Saunders DGO, et al. 2014.
 Single nucleus genome sequencing reveals high similarity among nuclei of an endomycorrhizal fungus. PLoS Genetics 10:e1004078
- 84. Johnson NC, Graham JH, Smith FA. 1997. Functioning of mycorrhizal associations along the mutualism-parasitism continuum. New Phytologist 135:575–85
- Smith SE, Smith FA. 2011. Roles of arbuscular mycorrhizas in plant nutrition and growth: new paradigms from cellular to ecosystem scales. *Annual Review of Plant Biology* 62:227–50
- Smith SE, Jakobsen I, Grønlund M, Smith FA. 2011. Roles of arbuscular mycorrhizas in plant phosphorus nutrition: interactions between pathways of phosphorus uptake in arbuscular mycorrhizal roots have important implications for understanding and manipulating plant phosphorus acquisition. *Plant Physiology* 156(3):1050–57
- 87. Sadhana B. 2014. Arbuscular mycorrhizal fungi (AMF) as a biofertilizers—a review. *International Journal of Current Microbiology* and Applied Sciences 3(4):384–400
- 88. Ortas I. 2012. The effect of mycorrhizal fungal inoculation on plant yield, nutrient uptake and inoculation effectiveness under long-term field conditions. *Field Crops Research* 125:35–48
- 89. Yang S, Li F, Malhi SS, Wang P, Suo D, et al. 2004. Long-term fertilization effects on crop yield and nitrate nitrogen accumulation in soil in northwestern China. *Agronomy Journal* 96:1039–49
- Mehrotra VS. 1996. Use of revegetated coal mine spoil as source of arbuscular mycorrhizal inoculum for nursery inoculations. Current Science 71:73–77
- Chaurasia B, Pandey A, Palni LMS. 2005. Distribution, colonization and diversity of arbuscular mycorrhizal fungi associated with

- central Himalayan rhododendrons. Forest Ecology and Management 207:315–24
- Stutz JC, Copeman R, Martin CA, Morton JB. 2000. Patterns of species composition and distribution of arbuscular mycorrhizal fungi in arid regions of southwestern North America and Namibia, Africa. Canadian Journal of Botany 78:237–45
- 93. An ZQ, Grove JH, Hendrix JW, Hershman DE, Henson GT. 1990. Vertical distribution of endogonaceous mycorrhizal fungi associated with soybean, as affected by soil fumigation. *Soil Biology and Biochemistry* 22:715–19
- Oehl F, Sieverding E, Ineichen K, Ris EA, Boller T, et al. 2005.
 Community structure of arbuscular mycorrhizal fungi at different soil depths in extensively and intensively managed agroecosystems. New Phytologist 165:273–83
- 95. Verma N, Tarafdar JC, Srivastava KK. 2010. Periodic changes in *Prosopis cineraria* associated AM population at different soil depth and its relationship with organic carbon and soil moisture. *African Journal of Microbiology Research* 4:11–12
- 96. Brady NC, Weil RR. 2008. The nature and properties of soils. Upper Saddle River, N.J.: Pearson Prentice Hall. pp. 662–710.
- Lehman RM, Taheri WI, Osborne SL, Buyer JS, Douds DD. 2012.
 Fall cover cropping can increase arbuscular mycorrhizae in soils supporting intensive agricultural production. Applied Soil Ecology 61:300–4
- 98. Säle V, Aguilera P, Laczko E, Mäder P, Berner A, et al. 2015. Impact of conservation tillage and organic farming on the diversity of arbuscular mycorrhizal fungi. *Soil Biology and Biochemistry* 84:38–52
- Mukherjee PK, Rai RK. 2000. Effect of vesicular arbuscular mycorrhizae and phosphate-solubilizing bacteria on growth, yield and phosphorus uptake by wheat (*Triticum aestivum*) and chickpea (*Cicer arietinum*). *Indian Journal of Agronomy* 45:602–7
- Rajendran K, Jayasree R. 2007. Effect of biofertilizers on quality seedling production of Acacia nilotica. Journal of Non-Timber Forest Products 14:5–11
- Zahid MA, Iqbal SM, Ali A, Hussain S. 2007. Efficacy of microbial bioagents for the control of collar rot disease in chickpea. Pakistan Journal of Botany 39:2667–72
- 102. Khan, Ayub N, Mirza SN, Nizami SM, Azam M. 2008. Synergistic effect of dual inoculation (vesicular arbuscular mycorrhizae) on the growth and nutrients uptake of *Medicago sativa*. *Pakistan Journal of Botany* 40:939–45
- Giri N. 2010. Growth and yield response of chick pea (Cicer arietinum) to seed inoculation with Rhizobium sp. Nature and Science 8(9):232–36
- Rodriguez A, Sanders IR. 2015. The role of community and population ecology in applying mycorrhizal fungi for improved food security. The ISME Journal 9:1053–61
- 105. Ceballos I, Ruiz M, Fernández C, Peña R, Rodríguez A, et al. 2013. The in vitro mass-produced model mycorrhizal fungus, Rhizo-phagus irregularis, significantly increases yields of the globally important food security crop cassava. PLoS One 8:e70633



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