

# Greenhouse gas emissions from managed freshwater wetlands under intensified aquaculture

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## Abstract

Wetlands are important niches for aquatic organisms and biodiversity as well as for organic carbon sequestration. However, wetlands are frequently managed for aquaculture production, which jeopardized their ability to act as a natural carbon sink. In this experiment, greenhouse gases (GHG) including CO<sub>2</sub>, CH<sub>4</sub> and N<sub>2</sub>O fluxes at the water-air interface, were monitored on a daily and monthly basis in four aquaculture wetlands: a natural wetland (NW), an enclosure wetland for intensive aquaculture (EW<sub>IA</sub>), a constructed wetland made from reclaimed paddy land for intensive aquaculture (CW<sub>IA</sub>), and a constructed wetland made from reclaimed paddy land for extensive aquaculture (CW<sub>EA</sub>). As a result, annual GHG fluxes for CW<sub>EA</sub>, CW<sub>IA</sub>, EW<sub>IA</sub> and NW wetlands averaged 0.81, 1.06, 2.43 and 1.12 kg N<sub>2</sub>O-N ha<sup>-1</sup>, 23.83, 457.08, 1,360.27 and 38.29 kg CH<sub>4</sub>-C ha<sup>-1</sup>, 1,321.32, 1,877.04, 2,246.79 and 1,305.48 kg CO<sub>2</sub>-C ha<sup>-1</sup>, respectively. Also, the highest yearly global warming potential (GWP) for EW<sub>IA</sub>, which was 60.02 t CO<sub>2</sub>-eq ha<sup>-1</sup>, was roughly ten times greater than that of NW, which was 6.04 t CO<sub>2</sub>-eq ha<sup>-1</sup>. Furthermore, the GHG emission intensity per unit of production ranged from 1.21 (CW<sub>EA</sub>)–5.30 (EW<sub>IA</sub>) kg CO<sub>2</sub>-eq kg<sup>-1</sup>, with EW<sub>IA</sub> having the highest N<sub>2</sub>O and CH<sub>4</sub> emission factors per unit of aquaculture output at 0.34 g N<sub>2</sub>O kg<sup>-1</sup> and 0.19 kg CH<sub>4</sub> kg<sup>-1</sup>. The study also revealed that feed quantity and water parameters positively linked with GHG emissions, suggesting that improving human management and water environmental quality could achieve large mitigation for both natural and artificial aquaculture wetlands. Given the high heterogeneity of GHGs emissions in aquaculture wetlands, future research should conduct long-term continuous automatic monitoring of aquaculture wetlands to understand the patterns of GHGs emissions and their responses and mechanisms to human management.

**Citation:** Yue Q, Cheng K, Sheng J, Wang L, Ji C, et al. 2023. Greenhouse gas emissions from managed freshwater wetlands under intensified aquaculture. *Soil Science and Environment* 2:3 <https://doi.org/10.48130/SSE-2023-0003>

## Introduction

Wetlands are critical terrestrial ecosystems that contain a huge global carbon (C) store (Gorham, 1991; Fourqurean et al., 2012; Mitsch et al., 2013) and may exert a significant impact on climate change (Zhang et al., 2002). Peat-accumulating wetlands currently hold approximately 390–455 Pg of terrestrial C, or nearly one-third of the world's soil C stock (Jenkinson et al., 1991). C capture and sequestration within wetlands is a complicated process as it involves both aerobic and anaerobic processes (Kayranli et al., 2010). Additionally, a variety of substrates could potentially be used to capture and sequester CO<sub>2</sub>, such as microorganism, algae (Sayre, 2010). In contrast, wetlands also have a major part to play in C cycling, with high GHG emission, estimated at 0.65 Pg C per year in the form of CH<sub>4</sub> (Bastviken et al., 2011), totalling up to 2.1 Pg C per year as CO<sub>2</sub> (Aufdenkampe et al., 2011). Therefore, maintaining wetlands as a large carbon sequestration site is an essential way to balance the whole terrestrial ecosystem.

Wetland reclamation under human factors would generate many additional anthropogenic GHG emissions and removals, of which aquaculture could be a typical artificially managed freshwater wetland system (Pendleton et al., 2012; Tan et al., 2020; Yang et al., 2022b). Yang et al. (2022a) reported a large

increase in CH<sub>4</sub> emission following conversion of coastal marsh to aquaculture ponds. Zhang et al. (2022) reported that aquaculture-emissions in China offset about 7% of terrestrial carbon burial, and revealed that the growth and intensification of the aquaculture sector raised climate concerns. Seitzinger (2000) showed that one third of the global anthropogenic N<sub>2</sub>O emissions were from aquatic ecosystems, and Hu et al. (2012) reported that the global N<sub>2</sub>O emission from aquaculture would be up to 383 Gg comprising 5.72% of anthropogenic N<sub>2</sub>O emission by 2030 from 93 Gg in 2009 if the aquaculture industry kept its current annual growth rate of roughly 7.1%. Wetlands Supplement (IPCC, 2013) for aquaculture emission adopted the literature from Hu et al. (2012 & 2013), and mentioned the need for in-situ experiments on the measurement of GHGs emissions from aquaculture systems primarily for developing emission factors of different wetland systems (Hu et al., 2013). Furthermore, some of the emission factors of Wetlands Supplement Guidelines were based on changes in soil C stocks or lab experiments, rather than on the limited field measurements available in the literature (Zheng et al., 2014). Given the limited available data to estimate GHGs emissions from constructed wetland system (IPCC, 2013), more scientific data from in-situ GHGs emissions measurement could support aquatic wetland emission estimation and management.

In addition, the reclamation of paddy fields for aquaculture was also an important mode of aquatic wetland system in China, which caused more GHGs emissions. Liu et al. (2016) demonstrated that CH<sub>4</sub> and N<sub>2</sub>O emissions reduced following conversion of rice paddies to inland crab-fish aquaculture. Particularly, N<sub>2</sub>O and CH<sub>4</sub> emission from aquaculture had become violent and vital because a vast input of feeding forage enriched substrates for biological activities in water and sediment (Hu et al., 2013); and Fang et al. (2022) confirmed that ebullition dominated the pathways of CH<sub>4</sub> emissions from inland aquaculture, which could be mainly influenced by water temperature and dissolved oxygen (Yang et al., 2023). Hence, the impact of natural and constructed wetlands for intensive aquatic culture on GHGs emissions should vary greatly due to differences in environment factors (water and sediment quality) and humanistic management practices (feed types, stocking density). Given this, the main purposes of this research were to (i) quantify the diurnal CHGs (including CO<sub>2</sub>, CH<sub>4</sub> and N<sub>2</sub>O) fluxes and seasonal variations in different types of managed wetlands; (ii) ascertain the effect of human-based management and environmental factors on CHGs emissions; and (iii) determine the influence of wetland utilization change on CHGs emissions.

## Materials and methods

### Site description

There was an abundance of aquaculture resources in Anhui province in China, with total water resources of 68 billion m<sup>3</sup>, 580,000 ha of inland water aquaculture, and an annual output of 2.3 Tg of aquatic products. Chizhou city was known for its aquatic products, with an annual output of 111 Gg, accounting for 11% of water resource in Anhui province. There were various types of aquaculture land with lakes accounting for 63% of the aquaculture area in inland waters, ponds for 20%, and paddy fields for 15%. Thus, Chizhou city (30°41' N, 117°34' E) was selected for this case study to measure GHGs emissions from different managed wetlands with a static floating gas-sampling chamber. The study area is close to the Yangtze River, and has a subtropical monsoon climate. Chizhou city has an annual mean temperature of 16.5 °C, an annual mean precipitation of 1,800 mm, a flat topography, dense network of rivers, and a suitable climate.

### Experiment design

Four wetlands, including natural wetland (NW), enclosure wetland for intensive aquaculture (EW<sub>IA</sub>), constructed wetland

for intensive aquaculture (CW<sub>IA</sub>) and constructed wetland for extensive aquaculture (CW<sub>EA</sub>), were selected for this study and they differ in terms of production and management. Of the four wetlands, one was natural wetland, while the other three were constructed. The information of the four wetlands are shown in Table 1.

### GHGs sampling and analysis

The static floating gas-sampling chamber was used for the experiments at four sites (Gao et al., 2014). The chambers were made of PVC and covered by a layer of foam and aluminum foil in order to insulating and reducing heat transmission and reflect light. Every chamber (30 cm inner diameter × 18 cm net height above the surface of the water after being dipped 5 cm into the water) had two holes on top, one for a sampling pipe with a three-way airtight valve and the other for a thermometer to monitor the temperature inside the chamber. All connections were keeping "air tight".

### Diurnal dynamics monitoring

For monitoring the diurnal dynamics of GHGs under different wetland management, the campaigns were conducted for 3 d at each site during the months of September to December 2014. Each site contained three replicate points, each of which was a tripod constructed in advance from three bamboo sticks. On each sampling day, five gas samples were taken every 2 h from 8:00 AM to 4:00 PM. For each chamber run, gas samples were collected at 0, 10, 20 and 30 min following the chamber installation (with a wire attached to a tripod to hang over the water). A syringe was used to extract a 40 mL gas sample, which was then injected into a vacuum gas sampling bottle (27 mL). At the end of each gas sample collection, the three-way airtight valve should be closed and the water, air and inside-chamber temperatures were noted. After four gas samples, floating chambers were raised and transported to the following sampling location. After collection, all samples were delivered to the lab to be measured over the course of two days.

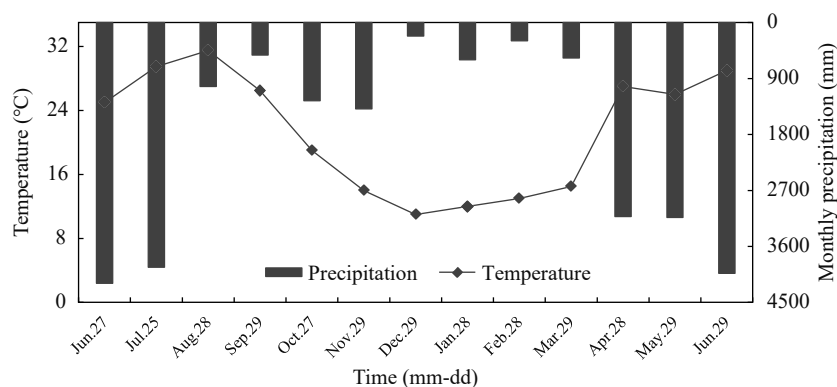
### Seasonal dynamics monitoring

The seasonal dynamic monitoring lasted 13 months, from June 2015 to June 2016 (Fig. 1) showed the temperature and precipitation during the experiments. The four sites of CW<sub>IA</sub>, CW<sub>EA</sub>, EW<sub>IA</sub> and NW had five, five, six and six replicate sampling points, respectively. On each sampling day, gas samples were taken between 9:00 to 12:00 AM. The same procedure as outlined above was applied to each chamber run. The seasonal monitoring frequency was once a month. Nevertheless, due to freezing temperatures, no data on GHG fluxes were available in January 2016.

**Table 1.** Basic information of the four experimental sites.

Sites	Origin	Area (hm <sup>2</sup> )	Water table m	Location		Feed	Forage content			Yield t ha <sup>-1</sup> year <sup>-1</sup>
				Longitude	Latitude		Total C (g kg <sup>-1</sup> )	Total N (g kg <sup>-1</sup> )	C/N*	
NW	A natural lake	1,000	3–9	30.67°	117.52°	NA	NA	NA	NA	/
EW <sub>IA</sub>	Wetland shifted to aquatic production	53	2–4	30.64°	117.46°	Fed on forage from March to October	127.12 ± 7.41	77.85 ± 2.03	1.6:1	11.32
CW <sub>EA</sub>	Paddy converted constructed wetland with	2	1	30.73°	117.58°	Fed on wheat grain or grass or fertilizer irregularly				5.00
CW <sub>IA</sub>	Paddy converted constructed wetlands with intensive aquaculture	2	1–2	30.66°	117.50°	Fed on forage from April to October	151.31 ± 2.80	72.39 ± 1.29	2.1:1	7.50

\* C/N represented the carbon-nitrogen ratio.



**Fig. 1** Temporal variation of environmental temperature and precipitation at the experimental sites.

### Gases flux calculation

The gas samples were analyzed by a gas chromatograph (Agilent 7890A GC, USA), and its measurement accuracy (reproducibility), which was the average relative standard deviation, was less than 1%. The detector used for CH<sub>4</sub> and CO<sub>2</sub> analysis was the FID with detection sensitivity of 0.5 and 1 ppm for CO<sub>2</sub> and CH<sub>4</sub>, and detection limit  $\leq 5.1 \times 10^{-12}$  g CO<sub>2</sub> s<sup>-1</sup> and  $1.9 \times 10^{-12}$  g CH<sub>4</sub> s<sup>-1</sup>, the column temperature was 80 °C, and the detector temperature was 200 °C; N<sub>2</sub> as the carrier gas with the flow rate of 40 mL min<sup>-1</sup>; H<sub>2</sub> as the fuel with the flow rate of 35 mL min<sup>-1</sup>; air as auxiliary fuel with the flow rate of 350 mL min<sup>-1</sup>. The detector for N<sub>2</sub>O was the ECD with detection sensitivity of 0.5 ppm for N<sub>2</sub>O and detection limit  $\leq 4.4 \times 10^{-15}$  g mL<sup>-1</sup>, the column temperature was 65 °C, and the detector temperature was 320 °C, argon methane gas as the carrier gas with a flow rate of 30 mL min<sup>-1</sup>. Gases flux rates were calculated (Healy et al., 1996; Altor & Mitsch, 2006) according to the following equation:

$$F_G = kV/A \times (dC/dT) \times \rho \times \left[ \frac{T_0}{(T_0 + T_C)} \right] \times \frac{P}{P_0} \times 60 \quad (1)$$

Where,  $F_G$  represented the flux rate (mg m<sup>-2</sup> h<sup>-1</sup> for CO<sub>2</sub> and NH<sub>4</sub>;  $\mu$ g m<sup>-2</sup> h<sup>-1</sup> for N<sub>2</sub>O);  $V$  was the chamber volume (m<sup>3</sup>);  $A$  was the sample surface area of the chamber (m<sup>2</sup>);  $dC/dT$  denoted the ratio of GHGs concentration change over time in a static chamber (ppm min<sup>-1</sup>);  $\rho$  was the gas density (g m<sup>-3</sup>);  $T_C$  was the temperature (°C) of the chamber at the time of each sample;  $T_0$  and  $P_0$  were the standard temperature (273 K) and atmospheric pressure (assume 1 atm), respectively;  $k$  was the gas dimensional conversion constant; 60 was conversion efficiency of per minute to per hour emission. As N<sub>2</sub>O was involved, the equation should be multiplied by 10<sup>3</sup> for converting mg to  $\mu$ g. CO<sub>2</sub>, CH<sub>4</sub> and N<sub>2</sub>O were accounted with global warming potentials (GWP) of 1, 28 and 265, respectively (IPCC, 2013).

In general, the flux rate by different time intervals of 4 gas sample concentration by linear regression analysis (Zou et al., 2005). The daily and seasonal cumulative GHGs emissions could be obtained by average value and observation interval times.

### Physiochemical analysis of sediment and water properties

At the beginning of the season's monitoring in June 2015, sediment samples at 0–15 cm depth were collected for four points by soil sample collectors and kept in plastic bags. From June 2015 to February 2016, water samples under the water's surface at a depth of 30 cm were taken monthly during the

seasonal monitoring, and stored in 250 mL plastic bottles. Accordingly, no samples were collected in January 2016 due to the freezing temperatures. All the samples were stored in an ice-filled cooler box, and were delivered to the lab being stored in a 4 °C refrigerator, then indicator analysis was completed within 2 d.

In the laboratory, the soil samples were air-dried, and finely ground to pass a 2 mm sieve prior to analysis. A portion of soil was ground to pass a 0.15 mm sieve for physical and chemical analysis of pH, soil organic matter (SOM), ammonium-N (NH<sub>4</sub><sup>+</sup>-N), nitrate-N (NO<sub>3</sub><sup>-</sup>-N), and total nitrogen (TN) using the protocol described by Lu (2000). For the water samples, properties analysis of pH, NH<sub>4</sub><sup>+</sup>-N, NO<sub>3</sub><sup>-</sup>-N, nitrite-N (NO<sub>2</sub><sup>-</sup>-N), TN, dissolved organic carbon (DOC) were carried out after filtration by 0.45  $\mu$ m filter membrane using the method reported by Lu (2000).

### Data analysis

Data processing was performed using Microsoft Office Excel 2013 and Origin 2016, and all statistical analyses were conducted using one-way ANOVA of SPSS and the least significant difference test were used to check the differences among the GHGs emissions of different aquaculture wetlands types.

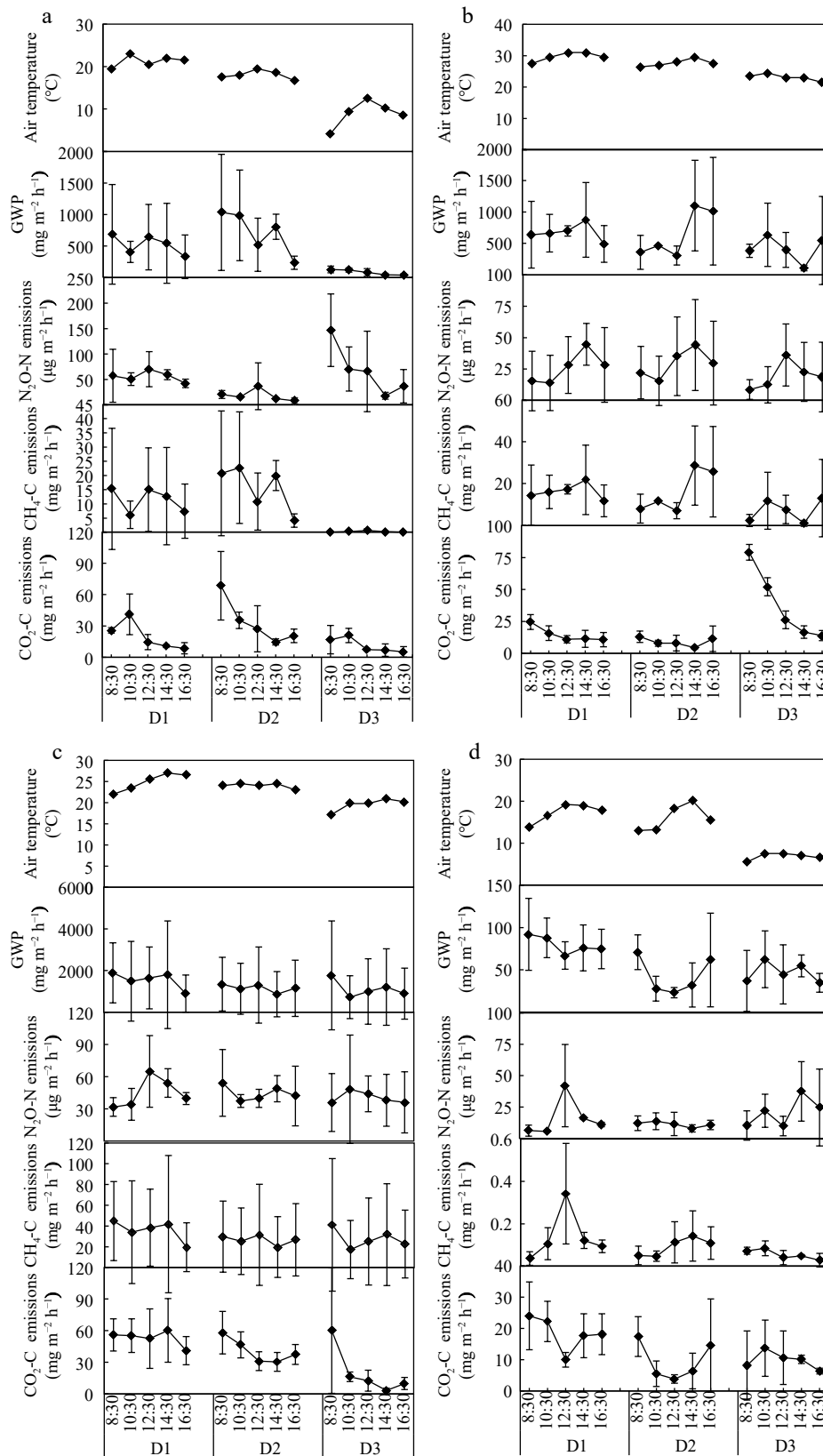
## Results

### Diurnal variation of GHGs fluxes

The diurnal variations of GHGs fluxes for four different aquaculture wetlands varied greatly at both time and spatial scales, of which were measured for three days with different measurement dates and temperature. As shown in Fig. 2, there was no consistency in the daily variation dynamics of N<sub>2</sub>O-N, CH<sub>4</sub>-C and CO<sub>2</sub>-C emission fluxes among the four sites. For the three human-managed wetland systems of CW<sub>IA</sub>, CW<sub>EA</sub> and EW<sub>IA</sub>, the GWP values were mainly determined by the CH<sub>4</sub> fluxes based on the similar trends. Particularly when the temperature dropped below 10 °C, the CH<sub>4</sub> emissions and GWP values at the CW<sub>IA</sub> site significantly decreased (Fig. 2a). However, this was not the case at the NW site, where the GHG fluxes remained relatively stable despite the temperature dropping below 10 °C, and the GWP values largely following the same dynamics as CO<sub>2</sub>-C fluxes (Fig. 2d).

### Seasonal variation of GHGs fluxes

As shown in Fig. 3, the emissions of N<sub>2</sub>O-N, CH<sub>4</sub>-C and CO<sub>2</sub>-C exhibited pronounced seasonal variations, with the highest emission rates occurring in the summer months. Firstly, the



**Fig. 2** Diurnal variation of CO<sub>2</sub>, CH<sub>4</sub> and N<sub>2</sub>O fluxes for (a) CW<sub>I</sub><sub>AV</sub>, (b) CW<sub>E</sub><sub>AV</sub>, (c) EW<sub>I</sub><sub>AV</sub>, (d) NW. D1, D2 and D3 for CW<sub>I</sub><sub>AV</sub> were October 16, November 3 and December 9 with daily mean temperature ranging from 21.3 to 9.0 °C; D1, D2 and D3 for CW<sub>E</sub><sub>AV</sub> were September 4, September 7, and September 22 with temperatures ranging from 29.7 to 23.1 °C; D1, D2 and D3 for EW<sub>I</sub><sub>AV</sub> were September 21, September 22, and October 14 with temperatures ranging from 24.9 to 19.6 °C; D1, D2 and D3 for NW were November 4, November 20, and December 11 with temperatures ranging from 17.6 to 8.4 °C. GWP, the global warming potential.

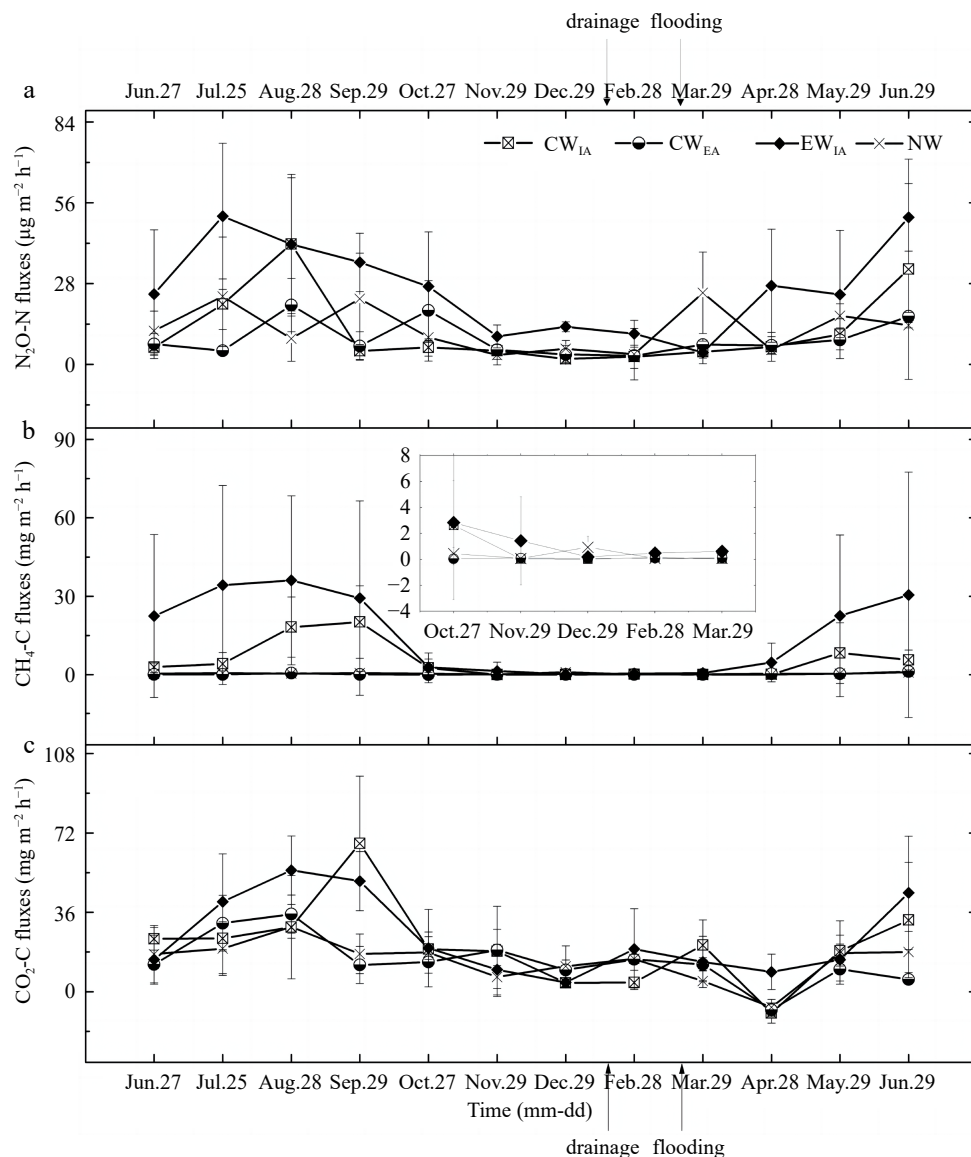
$\text{N}_2\text{O}$ -N fluxes ranged from 1.90 to 41.74  $\mu\text{g m}^{-2} \text{h}^{-1}$ , 3.03 to 20.59  $\mu\text{g m}^{-2} \text{h}^{-1}$ , 4.16 to 51.33  $\mu\text{g m}^{-2} \text{h}^{-1}$  and 3.25 to 24.82  $\mu\text{g m}^{-2} \text{h}^{-1}$  for  $\text{CW}_{\text{IA}}$ ,  $\text{CW}_{\text{EA}}$ ,  $\text{EW}_{\text{IA}}$  and NW, respectively (Fig. 3a). Secondly, the  $\text{CH}_4$ -C fluxes from  $\text{EW}_{\text{IA}}$  and  $\text{CW}_{\text{IA}}$  were higher compared to  $\text{CW}_{\text{EA}}$  and NW during the periods of June to September 2016 and May to June 2017. However, all sites showed low emission values ranging from 0.03 to 3.76  $\text{mg m}^{-2} \text{h}^{-1}$  during October 2016 to March 2017 (Fig. 3b). The  $\text{CO}_2$ -C fluxes varied across the sites, with values ranging from  $-9.71$  to 67.24  $\text{mg m}^{-2} \text{h}^{-1}$ ,  $-8.30$  to 35.20  $\text{mg m}^{-2} \text{h}^{-1}$ , 4.14 to 55.08  $\text{mg m}^{-2} \text{h}^{-1}$  and  $-7.05$  to 29.40  $\text{mg m}^{-2} \text{h}^{-1}$  for  $\text{CW}_{\text{IA}}$ ,  $\text{CW}_{\text{EA}}$ ,  $\text{EW}_{\text{IA}}$  and NW, respectively (Fig. 3c). In summary, the GHGs fluxes were consistently positive, except for the negatives values observed in April 2017, indicating that the four aquaculture systems were net emitters of GHGs.

Regarding cumulative GHG emissions,  $\text{EW}_{\text{IA}}$  had the highest cumulative  $\text{N}_2\text{O}$ ,  $\text{CH}_4$  and  $\text{CO}_2$  emissions with values of 2.43  $\text{kg N}_2\text{O-N ha}^{-1}$ , 1,360.27  $\text{kg CH}_4\text{-C ha}^{-1}$ , and 2,246.79  $\text{kg CH}_4\text{-C ha}^{-1}$  (Table 2). Additionally, the GHGs emission intensities per unit

product at the yield-scaled ranged from 1.21 to 5.30  $\text{kg CO}_2\text{-eq kg}^{-1}$ , and the highest  $\text{N}_2\text{O}$  and  $\text{CH}_4$  emission factors of aquaculture production were both from  $\text{EW}_{\text{IA}}$ , with the values of 0.34  $\text{g N}_2\text{O kg}^{-1}$  and 0.19  $\text{g CH}_4 \text{kg}^{-1}$ , respectively (Table 2). In Table 3, the GWP of  $\text{EW}_{\text{IA}}$ , with a value of 60.02  $\text{t CO}_2\text{-eq ha}^{-1}$  (the highest), was about 10 times higher than that of NW being 6.04  $\text{t CO}_2\text{-eq ha}^{-1}$ . For intensive aquaculture wetlands, the biggest GWP contributions came from  $\text{CH}_4$  emission, with proportions of 85% and 70% for  $\text{EW}_{\text{IA}}$  and  $\text{CW}_{\text{IA}}$ , respectively. In contrast, for NW and  $\text{CW}_{\text{EA}}$ , respectively,  $\text{CO}_2$  emissions made up the greatest quantities, at 72% and 80%.

### Environmental properties

Table 4 shows the analysis of environmental indicators at four different sites. For water quality, the pH values were slightly alkaline with an average of 7.48, ranging from 7.39 to 7.57. The concentrations of  $\text{NH}_4^+\text{-N}$  ranged between 0.25 to 0.66  $\text{mg L}^{-1}$ , and there was no significant difference among the four sites. The concentrations of  $\text{NO}_3^-$  were higher at, the  $\text{EW}_{\text{IA}}$



**Fig. 3** Seasonal GHG emission dynamics of (a)  $\text{N}_2\text{O}$ , (b)  $\text{CH}_4$  and (c)  $\text{CO}_2$  fluxes for different wetlands. The management practices of drainage and flooding occurred only to the sites of  $\text{CW}_{\text{IA}}$ ,  $\text{CW}_{\text{EA}}$  and  $\text{EW}_{\text{IA}}$ .

**Table 2.** Cumulative CO<sub>2</sub>, CH<sub>4</sub> and N<sub>2</sub>O fluxes and yield-scaled GHG emission intensity for different wetland systems.

Sites	Cumulative emission			GHG emission intensity per kg product		
	N <sub>2</sub> O-N (kg ha <sup>-1</sup> )	CH <sub>4</sub> -C (kg ha <sup>-1</sup> )	CO <sub>2</sub> -C (kg ha <sup>-1</sup> )	N <sub>2</sub> O (g kg <sup>-1</sup> )	CH <sub>4</sub> (kg kg <sup>-1</sup> )	Total (kg CO <sub>2</sub> -eq kg <sup>-1</sup> )
CW <sub>IA</sub>	1.06 ± 0.20	457.08 ± 161.20	1,877.04 ± 527.95	0.22	0.10	3.25
CW <sub>EA</sub>	0.81 ± 0.12	23.83 ± 7.80	1321.32 ± 91.11	0.25	0.01	1.21
EW <sub>IA</sub>	2.43 ± 0.20	1,360.27 ± 1552.98	2,246.79 ± 488.23	0.34	0.19	5.30
NW	1.12 ± 0.42	38.29 ± 11.99	1,305.48 ± 216.63	/	/	/

/: represented no data. The values were presented as the mean ± standard deviation.

site with a value of 1.36 mg L<sup>-1</sup> compared to the other three aquatic-wetland systems. For TN concentrations, NW had a lower value than CW<sub>IA</sub> and EW<sub>IA</sub>. The DOC content varied between 20.01 mg L<sup>-1</sup> at NW and 29.92 mg L<sup>-1</sup> at EW<sub>IA</sub>.

The sediment properties at all sites were weakly acidic, with pH values ranging from 6.0 to 6.9. Significantly, the pH value of CW<sub>EA</sub> was lower than that of the other three sites. The highest TN content was found at EW<sub>IA</sub> with a value of 3.34 g kg<sup>-1</sup>. The SOM contents showed a significant difference between NW and EW<sub>IA</sub>. The NH<sub>4</sub><sup>+</sup>-N concentrations was significantly higher at EW<sub>IA</sub> with a value of 203.22 mg kg<sup>-1</sup> compared to other three systems. The concentrations of NO<sub>3</sub><sup>-</sup>-N varied between 21.23 to 273.70 mg kg<sup>-1</sup>. The C/N ratio was between 7.00 to 8.58.

**Table 3.** Global warming potentials and their contributions for different wetland systems.

Sites	GWP	Contributions (%)		
	t CO <sub>2</sub> -eq ha <sup>-1</sup> y <sup>-1</sup>	N <sub>2</sub> O	CH <sub>4</sub>	CO <sub>2</sub>
CW <sub>IA</sub>	24.38	2%	70%	28%
CW <sub>EA</sub>	6.04	6%	14%	80%
EW <sub>IA</sub>	60.02	2%	85%	14%
NW	6.67	7%	21%	72%

**Table 4.** Water and sediment qualities at the four experimental sites.

Sites	pH	NH <sub>4</sub> <sup>+</sup> -N (mg L <sup>-1</sup> )	NO <sub>3</sub> <sup>-</sup> -N (mg L <sup>-1</sup> )	NO <sub>2</sub> <sup>-</sup> -N (mg L <sup>-1</sup> )	TN (g L <sup>-1</sup> )	DOC (mg L <sup>-1</sup> )	
							Water quality
	CW <sub>EA</sub>	7.39 ± 0.48 <sup>a</sup>	0.46 ± 0.33 <sup>a</sup>	0.36 ± 0.12 <sup>b</sup>	0.10 ± 0.16 <sup>ab</sup>	1.30 ± 0.52 <sup>bc</sup>	21.83 ± 7.67 <sup>ab</sup>
	EW <sub>IA</sub>	7.50 ± 0.28 <sup>a</sup>	0.66 ± 0.61 <sup>a</sup>	1.36 ± 0.56 <sup>a</sup>	0.36 ± 0.49 <sup>ab</sup>	2.07 ± 0.88 <sup>a</sup>	29.93 ± 8.92 <sup>a</sup>
	NW	7.57 ± 0.29 <sup>a</sup>	0.25 ± 0.20 <sup>a</sup>	0.28 ± 0.08 <sup>b</sup>	0.03 ± 0.03 <sup>b</sup>	0.66 ± 0.23 <sup>c</sup>	20.01 ± 7.71 <sup>b</sup>
Sites	pH	NH <sub>4</sub> <sup>+</sup> -N (mg kg <sup>-1</sup> )	NO <sub>3</sub> <sup>-</sup> -N (mg kg <sup>-1</sup> )	SOM (g kg <sup>-1</sup> )	TN (g kg <sup>-1</sup> )	C/N	
Sediment quality	CW <sub>IA</sub>	6.81 ± 0.13 <sup>a</sup>	36.67 ± 4.32 <sup>b</sup>	273.70 ± 185.08 <sup>a</sup>	39.69 ± 2.75 <sup>ab</sup>	2.95 ± 0.18 <sup>ab</sup>	7.81 ± 0.51 <sup>ab</sup>
	CW <sub>EA</sub>	6.22 ± 0.18 <sup>b</sup>	33.21 ± 6.39 <sup>b</sup>	54.57 ± 20.31 <sup>a</sup>	39.66 ± 4.16 <sup>ab</sup>	2.68 ± 0.19 <sup>b</sup>	8.58 ± 0.42 <sup>a</sup>
	EW <sub>IA</sub>	6.59 ± 0.29 <sup>a</sup>	203.22 ± 16.08 <sup>a</sup>	251.03 ± 263.67 <sup>a</sup>	44.00 ± 1.15 <sup>a</sup>	3.34 ± 0.06 <sup>a</sup>	7.46 ± 0.19 <sup>b</sup>
	NW	6.58 ± 0.14 <sup>a</sup>	27.86 ± 5.07 <sup>b</sup>	21.23 ± 5.10 <sup>a</sup>	35.78 ± 4.09 <sup>b</sup>	2.91 ± 0.47 <sup>ab</sup>	7.00 ± 0.73 <sup>b</sup>

TN, DOC, SOM and C/N represented total nitrogen, dissolved organic carbon, soil organic matter and carbon : nitrogen ratio, respectively.

**Table 5.** Correlation coefficients between GHGs emission and water parameters in all four wetlands.

Items	pH	DOC	NH <sub>4</sub> <sup>+</sup> -N	NO <sub>3</sub> <sup>-</sup> -N	NO <sub>2</sub> <sup>-</sup> -N	TN	Water temperature
Correlation	CH <sub>4</sub>	0.11	0.09	0.60**	0.80**	-0.17	0.13
	N <sub>2</sub> O	0.31	0.20	0.37*	0.60**	-0.22	0.10
	CO <sub>2</sub>	0.28	-0.11	0.70**	0.54**	-0.23	0.11
P value	CH <sub>4</sub>	0.574	0.633	0.002	0.000	0.399	0.517
	N <sub>2</sub> O	0.109	0.299	0.049	0.001	0.249	0.605
	CO <sub>2</sub>	0.144	0.576	0.000	0.003	0.246	0.575

\* Significant at P < 0.05; \*\* Significant at P < 0.01.

### Correlation analysis between environmental and management factors and GHGs emissions

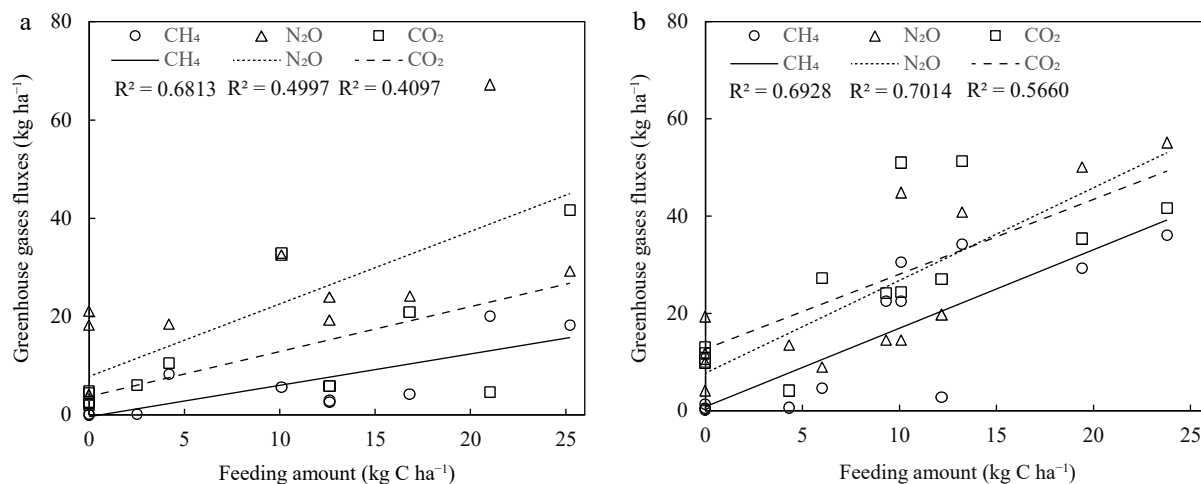
As shown in Table 5, there were strong positive correlations between NH<sub>4</sub><sup>+</sup>-N concentrations in water and the fluxes of CH<sub>4</sub> (R<sup>2</sup> = 0.60, P < 0.01), N<sub>2</sub>O (R<sup>2</sup> = 0.37, P < 0.05), and CO<sub>2</sub> (R<sup>2</sup> = 0.70, P < 0.01). Additionally, NO<sub>3</sub><sup>-</sup>-N concentrations in water were also strongly and positively correlated with CH<sub>4</sub> (R<sup>2</sup> = 0.80, P < 0.01), N<sub>2</sub>O (R<sup>2</sup> = 0.60, P < 0.01), and CO<sub>2</sub> fluxes (R<sup>2</sup> = 0.54, P < 0.01). In summary, the form and concentration of nitrogen in water had a significant impact on GHG emissions.

Furthermore, upon exploring forage-fed impact of CW<sub>IA</sub> and EW<sub>IA</sub> (Fig. 4), it was found that there were positive correlations between forage feeding and the CH<sub>4</sub> fluxes (R<sup>2</sup> = 0.68), N<sub>2</sub>O fluxes (R<sup>2</sup> = 0.50) and CO<sub>2</sub> fluxes (R<sup>2</sup> = 0.41) at the EW<sub>IA</sub> site; and there were positive correlations between forage feeding and the CH<sub>4</sub> fluxes (R<sup>2</sup> = 0.69), N<sub>2</sub>O fluxes (R<sup>2</sup> = 0.70) and CO<sub>2</sub> fluxes (R<sup>2</sup> = 0.57) at the CW<sub>IA</sub> site.

### Discussion

#### GHGs emissions in natural and constructed wetland

This study tracked GHGs emissions for four different aquatic-wetland systems, partially reflecting the variations in how human management practices affect wetland GHG emissions.



**Fig. 4** Dependence of  $\text{CO}_2$ ,  $\text{CH}_4$  and  $\text{N}_2\text{O}$  fluxes on feeding amount at the sites of (a)  $\text{CW}_{\text{IA}}$  and (b)  $\text{EW}_{\text{IA}}$ .

In the current study, the average  $\text{CO}_2$ ,  $\text{CH}_4$  and  $\text{N}_2\text{O}$  emissions from nature wetland (NW) were  $51.09 \text{ mg m}^{-2} \text{ h}^{-1}$ ,  $0.55 \text{ mg m}^{-2} \text{ h}^{-1}$  and  $19.50 \text{ } \mu\text{g m}^{-2} \text{ h}^{-1}$ , respectively, which were similar to those values ranging from  $15.40$  to  $1082.00 \text{ mg CO}_2 \text{ m}^{-2} \text{ h}^{-1}$ ,  $0.04$  to  $175.94 \text{ mg CH}_4 \text{ m}^{-2} \text{ h}^{-1}$  and  $13.3$  to  $747 \text{ } \mu\text{g N}_2\text{O m}^{-2} \text{ h}^{-1}$  shown in Table 6. The high fluxes of  $\text{CH}_4$  and  $\text{CO}_2$  at the canal were mostly caused by domestic waste pollution (Han et al., 2013). The  $\text{N}_2\text{O}$  fluxes from aquatic-wetlands converted by agricultural land in the study were  $14.08 \text{ } \mu\text{g m}^{-2} \text{ h}^{-1}$  ( $\text{CW}_{\text{EA}}$ ) and  $18.65 \text{ } \mu\text{g m}^{-2} \text{ h}^{-1}$  ( $\text{CW}_{\text{IA}}$ ), which were higher than the values of  $10.74$  and  $11.8 \text{ } \mu\text{g m}^{-2} \text{ h}^{-1}$  for the aquatic wetlands systems of shrimp and mixed shrimp-fish pond (Yang et al., 2015), but lower than that of the crab-fish aquatic system with a value of  $48.1 \text{ } \mu\text{g m}^{-2} \text{ h}^{-1}$  (Liu et al., 2016). The  $\text{CH}_4$  fluxes of  $0.35 \text{ mg m}^{-2} \text{ h}^{-1}$  at the  $\text{CW}_{\text{EA}}$  site was the lowest among these inland aquaculture systems possibly due to the absence of amount forage feed.

The estimates of global wetland emissions were still subject to severe data limitations and distributional biases, especially for the emission under human intervention (Selvam et al., 2014). The 2006 IPCC Guidelines, which was limited to peatlands that had been drained and managed for peat extraction, provided methods for estimating national anthropogenic emissions (IPCC, 2006). The 2013 Supplement to the 2006 IPCC Guidelines for National Greenhouse Gas Inventories was published to reduce uncertainties of wetland emissions estimates. It included information on inland organic soils, wetlands on mineral soils, coastal wetlands, as well as constructed wetlands for wastewater treatment (IPCC, 2013). However, the evaluation methods for artificially managed aquatic wetlands still needed to be improved. Hence, based on more reliable observational data, scientists can simulate GHGs emissions through modeling and analyze human impacts on the GHG balance of wetland ecosystems (Petrescu et al., 2015).

### Effect of human management and environmental factors on GHG emissions

For aquatic wetland system, human factors such as C and N inputs, water level control, fish type and fish intensity could affect the intensity of GHGs emission (Yang et al., 2013; Zheng et al., 2014; Yang et al., 2023). Forage feeding rate directly affected ammonia concentrations in the aqueous phase, which then affected the fate or release rate of  $\text{CH}_4$  and  $\text{N}_2\text{O}$  from the

sediment-water interface to the atmosphere (Liu et al., 2016). Beaulieu et al. (2011) reported that approximately 1.00% of N input may be transformed to  $\text{N}_2\text{O}$  for rivers and estuaries, and Seitzinger & Kroeze (1998) discovered that roughly 0.75 % of N for streams was released to the atmosphere as  $\text{N}_2\text{O}$ . Hu et al. (2013) also revealed that about 1.3% of the nitrogen input was emitted as  $\text{N}_2\text{O}$  gas in laboratory-scaled aquaculture system. While this study found that approximately 0.2%–0.3% of N input became  $\text{N}_2\text{O}$  emissions. In addition, changes in water levels (such as drainage or irrigation) were common management practices in aquaculture, which could significantly affect  $\text{CH}_4$  emissions (Friborg et al., 2000; Jackowicz - Koczyński et al., 2010).

Environmental factors were closely related to the input of C and N, directly affecting GHGs emissions. Numerous research had demonstrated that the  $\text{CH}_4$  and  $\text{N}_2\text{O}$  fluxes were associated with environmental factors, such as air and soil temperature, dissolved oxygen and DOC in water, and the availability of C and N substrate in sediment (Bhattacharyya et al., 2013; Hu et al., 2016). According to this study, both the  $\text{NH}_4^+\text{-N}$  and  $\text{NO}_3^-\text{-N}$  concentrations showed strong positive correlations with  $\text{CH}_4$ ,  $\text{N}_2\text{O}$  and  $\text{CO}_2$  fluxes (Table 5). Liu et al. (2016) studied that  $\text{CH}_4$  and  $\text{N}_2\text{O}$  emissions were negatively related to the water DO concentration. Tsuneda et al. (2005) found that  $\text{N}_2\text{O}$  conversion increased when the salinity increased from 10 to 20  $\text{mg L}^{-1}$  during single nitrification. Whereas Barnes et al. (1999) and Hashimoto et al. (1999) revealed a negative relationship between salinity and dissolved  $\text{N}_2\text{O}$  concentration. Moreover, GHG emissions in aquaculture systems were linked to aquatic animal species due to variations in their digestion processes, such as filter- and deposit-feeding, as observed in studies by Stief et al. (2009) and Bhattacharyya et al. (2013). Therefore, managing the entire range of aquatic organisms in aquaculture can play a crucial role in mitigating GHG emissions.

Among various forms of aquatic management, GHGs emission factors on the scale of area and yield varied significantly. This study found a positive correlation between cumulative GHG emissions intensity and yield in aquatic wetlands. Generally speaking, the production of  $\text{CW}_{\text{EA}}$ ,  $\text{CW}_{\text{IA}}$  and  $\text{EW}_{\text{IA}}$  increased sequentially, reaching 5.00, 7.50 and 11.32  $\text{t ha}^{-1}$ , respectively. The corresponding annual cumulative emissions also increased sequentially, reaching 0.81, 1.06 and 2.43  $\text{kg N}_2\text{O-N ha}^{-1}$ , 23.83, 1,877.04 and 2,246.79  $\text{kg CH}_4\text{-C ha}^{-1}$ ,

**Table 6.** Comparison of greenhouse gases fluxes from different wetland types with previous studies in China and other regions.

	Location	Sources	Time horizon	Wetland types	N <sub>2</sub> O fluxes	CH <sub>4</sub> fluxes	CO <sub>2</sub> fluxes			
					$\mu\text{g m}^{-2} \text{h}^{-1}$	$\text{mg m}^{-2} \text{h}^{-1}$	$\text{mg m}^{-2} \text{h}^{-1}$			
China	East China	Current study	13 months	Constructed wetland for intensive aquaculture	18.65 ± 20.48	6.92 ± 9.41	77.23 ± 68.95			
				Constructed wetland for extensive aquaculture	14.08 ± 9.58	0.35 ± 0.45	51.21 ± 40.97			
				Enclosure wetland for intensive aquaculture	41.89 ± 24.83	20.60 ± 19.90	90.34 ± 65.45			
	East China	Liu et al. (2016)	12 months	Natural wetland	19.50 ± 12.48	0.55 ± 0.41	51.09 ± 33.48			
				Rice paddies	110.5	0.71				
				Crab-fish aquaculture	48.1	0.37				
	Southeastern China	Yang et al. (2015)	5 months	Shrimp pond	10.74	19.95	20.78			
				Shrimp-fish pond	11.8	1.65	-60.46			
	Lab simulation of aquatic system	Hu et al. (2014)	2 months	Control	5,504 ± 1,221		2,401 ± 343			
				Treatment	911 ± 488		4,590 ± 546			
Overseas of Nature wetlands	Yangtze estuarine, China	Wang et al. (2009)	12 months	Marsh site		2.06				
				Bare tidal flat		0.04				
	East China	Han et al. (2013)	3 months	Open river	62.17 ± 2.13	16.41 ± 3.06	162.18 ± 10.55			
				Canal	111.74 ± 7.41	175.94 ± 18.42	1,082.00 ± 90.53			
				Reservoir	50.99 ± 2.68	1.70 ± 0.14	25.61 ± 4.08			
	Northeast China	Yang et al. (2013)	3 months	Freshwater marsh	13.3	8.92	394.5			
				India	Selvam et al. (2014)	24 h/1–8 h	Ponds		11.93 ± 12.33	123.02 ± 117.33
	Rivers		4.13 ± 8.27	36.85 ± 70.58						
	Reservoirs		2.13 ± 2.33	15.40 ± 35.75						
	Colombia	Dennis et al. (2014)	11 months	Restored mangrove		162 ± 48	3.47 ± 1.52			
					97 ± 46	11.78 ± 10.54				
					747 ± 299	6.12 ± 9.69				
Costa Rica	Nahlik et al. (2011)	29 months	Forested marsh			5.05				
				Rainforest swamp			33.39			
				Alluvial marsh			39.94			
Australia	Allen et al. (2011)	6 months	Mangrove		30.00	0.30				
				Orinoco River Floodplain, Venezuela	Smith et al. (2000)	17 months	Open river		1.33	
							Flooded forest		4.67	
			Macrophyte mats			1.33				

1,321.32, 1,877.04 and 2,246.79 CO<sub>2</sub>-C kg ha<sup>-1</sup>. Petrescu et al. (2015) reported that management intensity strongly influenced the net climate footprint of wetlands. The highest N<sub>2</sub>O emission factor of aquaculture production from the intensive aquaculture type of EW<sub>IA</sub> was 0.34 g kg<sup>-1</sup>, which was still much lower than the values of 2.65 and 2.57 g kg<sup>-1</sup> reported by Hu et al. (2012) and Liu et al. (2016) using lab simulation experiments. In terms of GWP, aquatic system with high forage feeding intensity had significantly higher annual GWP compared to the wetland systems with no or low forage feeding intensity. After the intensive aquaculture management of wetlands, the main GHG emissions of the system shifted from CO<sub>2</sub> to CH<sub>4</sub>, which greatly increased the GWP. For instance, the largest contributions to GWP for the intensive aquaculture of EW<sub>IA</sub> and CW<sub>IA</sub> was CH<sub>4</sub> emission, while for natural aquatic-wetland and exclusive aquaculture, the main contributor turned out to be CO<sub>2</sub> emission (Table 3). Therefore, scientific management of aquaculture density, forage feeding intensity, and water level in aquatic wetland systems would play a crucial role for the trade-off between CH<sub>4</sub> and CO<sub>2</sub> emission, as well as GHG mitigation (Petrescu et al., 2015).

### Uncertainties analysis

The GHGs emission fluxes found in this investigation were usually characterized by high standard deviations (Figs 2 and 3), which were in agreement with the results from Nahlik et al. (2011) and Dennis et al. (2014). The large variances indicated significant differences in emissions between parallel points at

the same site, which might be caused by changes in water level, temperature, ebullition, and environmental disturbance (Liu et al., 2016; Petrescu et al., 2015; Fang et al., 2022; Yang et al., 2022b). For example, the sediment and water quality in the areas near the feeding machines were complicated, and the breeding density in this region was frequently high, both of which greatly affected the GHGs emission intensity (Chen et al., 2013; Hou et al., 2013).

Besides, the sampling points were uniformly set at different locations of each site, and the process of sailing for sampling could also cause some interference to the water body, like creating some surface waves, thereby affecting the detection results and causing uncertainties to the results (Gao et al., 2014). More importantly, the bamboo poles used for fixation were more or less influenced by ships, causing disturbance to the sediment and generating bubbles, further affecting the fate or release rate of GHGs through the sediment-water interface system (Hirota et al., 2004; Bastviken et al., 2008). Therefore, future work should improve the equipment technology for monitoring GHGs emission in wetlands. Finally, diurnal dynamic measurements were carried out at the four sites for eight hours due to the safety issues, night time monitoring was excluded. This may consequently lead to the overestimation of the GHGs fluxes because of lower night time temperatures. So, it is necessary to monitor the emission characteristics throughout the whole day as much as possible.



## Conclusions

For the four aquatic-wetlands, the extensive aquaculture system from reclaimed paddy land ( $CW_{EA}$ ) had the lowest GHG emission intensity with  $0.81 \text{ kg N}_2\text{O-N ha}^{-1}$ ,  $23.83 \text{ kg CH}_4\text{-C ha}^{-1}$  and  $1,321.32 \text{ kg CO}_2\text{-C ha}^{-1}$ , while the intensive aquaculture system from wetland enclosure ( $EW_{IA}$ ) had the highest cumulative GHG emission with the values of  $2.43 \text{ kg N}_2\text{O-N ha}^{-1}$ ,  $1360.27 \text{ kg CH}_4\text{-C ha}^{-1}$ ,  $2,246.79 \text{ kg CO}_2\text{-C ha}^{-1}$ , respectively. And the GHG emission intensity per unit of production ranged from  $1.21 (CW_{EA})$ – $5.30 (EW_{IA}) \text{ kg CO}_2\text{-eq kg}^{-1}$ . The current study supported a valuable reference for  $\text{N}_2\text{O}$  and  $\text{CH}_4$  emission factors in constructed wetlands and demonstrated that the intensive aquaculture ultimately led to the high area-scaled and yield-scaled GHGs emission intensity. These measurement data served as a crucial support for the follow-up model simulation and GHGs emission prediction on a large scale.

## Acknowledgments

This work was financially supported by Natural Science Foundation of China under a grant number 41907073 and 41877546.

## Conflict of interest

The authors declare that they have no conflict of interest.

## Dates

Received 15 January 2023; Accepted 19 April 2023; Published online 11 May 2023

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