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Genetic improvement of barnyard millet (*Echinochlo aesculenta***): prospective and challenges**

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Abstract

Barnyard millet is a vital yet underutilized cereal crop due to limited genetic research and improvement efforts. This review explores promising avenues like QTL mapping and Marker-Assisted Selection for targeted breeding, along with the exciting possibilities of genomic selection and speed breeding for faster variety development in barnyard millet. Even gene editing with CRISPR-Cas holds promise, but challenges like incomplete genome knowledge need to be addressed. Additionally, bioinformatics and systems biology approaches offer powerful tools to understand the complex interaction of genes and traits in barnyard millet. While limitations exist, actively pursuing these advancements can unlock barnyard millet's full potential, significantly impacting global food security and sustainable agriculture.

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Introduction

Barnyard millet (*Echinochloa esculenta*) is garnering attention for its potential to contribute significantly to food and nutritional security, particularly in Asia^[1]. It ranks as the fourth most produced minor millet, and India leads globally in barnyard millet production, covering the largest area (0.146 mha) and yielding the highest output (0.147 mha) with an average productivity of 1034 kgha−1 over the past three years[2] . *E. esculenta*, known as Japanese barnyard millet, is valued for its fast growth, adaptability to various soil and water conditions, and nutritional grain profile, making it an excellent choice for forage and food[3] . Moreover, *E. frumentacea*, or Indian barnyard millet, shares similar traits but is often preferred in traditional farming systems in South Asia due to its hardiness and ease of cultivation^{[[4\]](#page-6-3)}. In crop improvement, E. esculenta's rapid growth and resilience are leveraged to enhance stress toler-ance and yield in other cereal crops^{[\[5,](#page-6-4)[6](#page-6-5)]}. Barnyard millet emerges as a beacon of hope for regions grappling with unpredictable weather patterns, and its secret lies in its remarkably short life cycle, maturing in just 60−80 days[7] . This swiftness allows farmers to double-crop, squeezing in two harvests within a single season^[8]. Technology interspective and **computer interspective and the effective of the energy interspective and the Propositive and Challengens**
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Even more crucially, the short cycle enables them to dodge droughts by planting just before the rains and harvesting before potential dry spells^[9]. This adaptability to erratic rainfall patterns is further bolstered by barnyard millet's inherent resilience^[1]. Additionally, the nutritional benefits of barnyard millet, including*rich in protein, carbohydrate and dietary fiber, low glycemic index, gluten-free, abundance of micronutrients like iron and zinc good source of vitamin B, and rich in antioxidants*, position it as a valuable crop for addressing nutritional deficiencies, particularly in hilly and tribal communities^[10].

Moreover, [Fig 1](#page-1-0) categorizes different biological data types and resources, along with the number of records available for

each in NCBI (National Center for Biotechnology Information) database as of 20th July 2024 for Japanese barnyard millet (*E. esculenta*) (*Taxonomy ID: 121770*) [11] . For Literature, sources include *Bookshelf, PubMed*, and *PubMed Central*, totalling 168 records[11] . Genes are covered by the *Gene* and *PopSet* databases with 142 records, while Proteins are split into *Identical Protein Groups* and *Protein databases* with 91 and 203 records respectively[11] . Genomes are sourced from *BioProject* and Taxonomy with 5 records, *BioSample* with 165 records, *Nucleotide* with 69 records, and the *Sequence Read Archive* with 164 records. Lastly, the *PubChem* database includes 21 BioAssays records^[11].

Moreover, the genetic improvement of barnyard millet holds significant potential but requires concerted research efforts to overcome existing challenges^[12]. However, genome research in barnyard millet is still in its early stages and lags significantly behind other minor millets, and this is primarily due to the complex nature of its genome (2n = $6x = 54$, hexaploid)^{[[6\]](#page-6-5)}. Enhanced genetic characterization, effective utilization of germplasm resources, and advanced breeding techniques, including interspecific hybridization, are essential for unlocking the full potential of this underutilized crop^[5]. The limited genetic diversity observed within cultivated varieties, compared to their wild relatives, further complicates breeding efforts aimed at enhancing desirable traits like yield, disease resistance, and abiotic stress tolerance^[13].

On the positive side, recent advancements in genomics have opened new avenues for the genetic enhancement of barnyard millet^[5]. The release of genome and transcriptome sequences for both wild and cultivated *Echinochloa* sp. has facilitated a better understanding of the genetic basis of important agronomic traits[9] . The identification of *quantitative trait loci* (QTL) and specific genes associated with these traits is crucial for the development of improved varieties^{[\[9\]](#page-6-8)}. Further-

Fig. 1 Distribution and abundance of different types of biological data of Japanese barnyard millet (*E. esculenta*) across various databases as of 20th July 2024^[11]

frumentacea has shown promise, despite challenges related to seed sterility in hybrid plants^{[\[14\]](#page-6-13)}. This hybridization can potentially introduce desirable traits from one species to the other, thereby broadening the genetic base and enhancing the crop's overall performance^[15].

Despite nutritional advantages,barnyard millet remains underutilized^[9]. The genetic improvement of barnyard millet faces several challenges and presents numerous prospects. One of the main challenges is the limited genetic and genomic resources available for this crop^[7]. Although recent efforts have led to the development of core collections and the identification of *single-nucleotide polymorphisms* (SNPs) within these collections, there is still a significant gap in understanding the full genetic architecture of barnyard millet^[16]. Moreover, integrating advanced molecular and biotechnological tools with conventional breeding approaches can significantly enhance the agronomic traits of *E. esculenta*[17] .

Techniques such as genome editing with CRISPR (Clustered *Regularly Interspaced Short Palindromic Repeats*) - Cas9 (*CRISPRassociated protein 9*) can precisely target and modify genes associ[ate](#page-6-17)d with important agronomic traits in barnyard millet^{[\[18\]](#page-6-17)}. *Marker-assisted selection* (MAS) allows for the efficient

identification and incorporation of desirable traits by using molecular markers linked to key agronomic characteristics in barnyard millet^[19]. Combining these molecular tools with traditional breeding ensures the development of superior *E. esculenta* varieties with enhanced performance and resilience^[20−22].

Therefore, the main focus of this review is to explore how these advanced tools can be effectively integrated with conventional methods to improve the agronomic traits of *E. esculenta*, emphasising on the potential for developing highyield, stress-resistant barnyard millet varieties to enhance food security and agricultural sustainability.

Origin, classification, and genomic relationship

Barnyard millet, despite its name, is a true powerhouse grain belonging to the grass family Poaceae^[22]. While its exact origin remains unclear, it's believed to have been domesticated in India or Southeast Asia. Classified within the genus *Echinochloa*, it shares some relatives [w](#page-6-21)ith other millets but possesses a unique genetic makeup^{[\[23\]](#page-6-21)}. Moreover, barnyard millet is a significant crop primarily cultivated in semi-arid tropical

regions of Asia and Africa, appreciated for its drought toler-ance, rapid maturation, and nutritional benefits^{[[24](#page-6-22)]}. The genus *Echinochloa* comprises multiple species, with the two major cultivated types being *E. frumentacea* (Indian barnyard millet) and *E.esculenta* (Japanese barnyard millet)^{[\[25\]](#page-6-23)}. The origins, classification, and genomic relationships of barnyard millet can be understood through a synthesis of genetic and genomic research^[24].

Barnyard millet has a complex origin involving multiple species and subspecies^[13,22]. The genomic relationship of barnyard millet species has been explored using various molecular markers[16] . A study utilizing whole-genome genotyping-bysequencing on a core collection of 89 barnyard millet accessions identified several thousand SNPs segregating within the collection^[16]. This study revealed four population clusters within *E. colona* and three within *E. crus-galli*, suggesting that the genetic diversity within these species is substantial and likely influenced by geographic factors^{[\[16\]](#page-6-15)}. Additionally, the use of RAPD (Random Amplified Polymorphic DNA) primers confirmed the genetic diversity among 21 accessions of the *Echinochloa* spp. complex, classifying them into two morphological races, *E. frumentacea* and *E. colona*, with a high degree of molecular diversity^[26].

Moreover, interspecific hybridization between *E. esculenta* and *E. frumentacea* offers potential for genetic enhancement[26] . A successful hybrid between *E. esculenta* cultivar PRJ 1 and *E. frumentacea* cultivar ER 72 is also reported, confirmed through rice SSR markers^[15]. Although the hybrid plants exhibited vigor and disease resistance, they were sterile, highlighting challenges in utilizing interspecific hybrids for immediate breeding programs^[15]. Therefore, the genetic characterization of barnyard millet is crucial for breeding and improvement programs. For instance, association analysis in germplasm and $F₂$ populations has shown positive direct effects of traits like stem girth, ear head weight, and plant height on grain yield^[27]. Moreover, understanding its genomic relationships with other *Echinochloa* species is crucial for breeding programs to develop improved barnyard millet varieties with enhanced yields and desirable traits^[23].

The phylogeny of barnyard millet has been the subject of multiple studies focusing on genetic diversity, population structure, and evolutionary relationships among different species and accessions^[16]. However, these studies identify thousands of SNPs and revealed distinct population clusters, and interestingly, these genetic clusters correlated more with geographic origin rather than morphological classification, highlighting the influence of geographic isolation on genetic diversity^[28].Another significant contribution to understanding the phylogeny of *Echinochloa* species comes from the analysis of chloroplast DNA (cpDNA) sequences from non-coding regions (trnT-L-F)^[29], and the findings suggest that domestication and adaptation to different environments occurred after the species diverged in Asia^[28].

Further phylogenetic insights are provided by the complete chloroplast genome sequencing of Indian barnyard millet (*E. frumentacea*), and this study revealed that *E. frumentacea* diverged from its close relatives *E. oryzicola* and *E. crus-galli* approximately 1.9-2.7 million years ago^[30]. However, morphoagronomic trait analysis has contributed to our understanding of barnyard millet phylogeny^{[[31](#page-7-3)]}. Lastly, the assessment of genetic diversity using molecular markers further elucidates the phylogenetic relationships within and between *E. crus-galli* and *E. frumentacea*[\[32\]](#page-7-4) . However, the phylogeny of barnyard millet is complex and influenced by geographic, genetic, and morpho-logical factors^{[\[33\]](#page-7-5)}. Moreover, the phylogeny of barnyard millet is characterized by significant genetic diversity and complex evolutionary relationships, and studies utilizing whole-genome sequencing, interspecific hybridization, and molecular markers have provided a deeper understanding of these relationships, offering valuable insights for crop improvement^[34].

Germplasm resources

Germplasm resources in barnyard millet hold significant potential for crop improvement, particularly in terms of enhancing traits such as drought tolerance, nutritional quality, salinity tolerance, and disease resistance^[35]. Several studies have characterized the genetic variability and diversity within barnyard millet germplasm, providing valuable insights for breeders and researchers^[35]. This genetic diversity is crucial for the development of improved cultivars with desirable traits^{[\[26\]](#page-6-24)}. The formation of a core set in barnyard millet germplasm using data on 24 morpho-agronomic traits has been instrumental in managing and utilizing the vast genetic resources available[\[16\]](#page-6-15). This core set captures most of the available diversity and serves as a representative sample for detailed evaluation and breeding programs^[31].

Similarly, another study characterized 494 barnyard millet germplasm for quantitative traits, revealing significant variation for most traits and highlighting traits like days to 50% flowering, plant height, and days to maturity as key contributors to total variance^[36]. Moreover, barnyard millet germplasm has been screened for resistance to grain smut with accessions showing varying levels of susceptibility, and the study suggests that smut resistance and grain yield can be improved simultaneously through proper breeding strategies^[37]. Additionally, the crop's tolerance to salinity has been investigated, with certain genotypes demonstrating the ability to withstand up to 200 mM salt concentration during germination, and it is found that, this tolerance is associated with traits such as higher relative water content and enzyme activity under salt stress^[38]. The genetic variability and association studies under sodic soil conditions have identified traits like days to fifty percent flowering, days to maturity, ear width, and thousand grain weight as important selection indices for enhancing grain yield^[39]. example. The metric of the metric of the spheric of the

The characterization of genetic diversity and the formation of core collections facilitate the efficient utilization of barnyard millet germplasm in breeding programs aimed at developing high-yielding, stress-tolerant, and disease-resistant cultivars^{[\[40\]](#page-7-12)}. While the exact number of barnyard millet germplasm accessions globally remains around 8,000 conserved in various gene banks, such as *Vivekananda Parvatiya Krishi Anusandhan Sansthan (VPKAS), India, ndian Institute of Millets Research (IIMR), India, National Institute of Agrobiological Sciences (NIAS), Japan, and Consultative Group on International Agricultural Research (CGIAR), including the International Crops Research Institute for the Semi-Arid Tropics (ICRISAT), India*, recent efforts focus on optimizing their useand maintenance^[9]. The extensive genetic resources available in barnyard millet germplasm provide a solid foundation for crop improvement^[4,41].

Genome sequencing

Genome sequencing in barnyard millet has made significant strides in recent years, providing valuable insights into the genetic makeup and potential for crop improvement in this under-researched species^{[\[15,](#page-6-14)[16](#page-6-15)]}. A groundbreaking study on the genetic characterization of a global barnyard millet germplasm collection utilized whole-genome genotyping-by-sequencing to identify several thousand SNPs in 89 accessions, providing a robust framework for breeders to enhance barnyard millet for smallholder farmers^[16]. However, researchers in China successfully released and annotated the whole genome sequence of weedy *E. crus-galli*, and the genome, sequenced at a depth of 171x, was estimated to be 1.27 Gb, covering approximately 90.7% of the predicted size^{[\[9\]](#page-6-8)}. Moreover, the study identified 4,534 contigs ranging from 1 kb to 11.7 Mb, with gene annotations revealing 108,771 protein-coding genes, 785 miRNAs, 514 Mb of repetitive elements, and various non-coding RNAs^[9]. NCBI resources for *Echinochloa* species included 1,246 nucleotide sequences, 822 gene sequences, 2,468 protein sequences, and other genomic data, with *E. crus-galli* leading in sequence availability^[9]. Despite this, cultivated barnyard millet species *E. frumentacea* and *E. esculenta* had significantly fewer sequences, representing only 4% of the total^[9].

Further insights were gained from the complete chloroplast genome sequence of Indian barnyard millet, *E. frumentacea*[30] . The chloroplast genome, which is 139,593 bp in length with a typical quadripartite structure, includes 112 individual genes, and these genes consist of 77 protein-coding genes, 30 tRNA genes, four rRNA genes, and one conserved open reading frame, with an overall GC content of 38.6%^[30]. Phylogenetic analysis indicated that *E. frumentacea* diverged from its close relatives, *E. oryzicola* and *E. crus-galli*, approximately 1.9–2.7 million years ago, highlighting the evolutionary history and genetic uniqueness of this species^[30]. Moreover, genomics and genetic studies in barnyard millet are rapidly advancing, providing a wealth of data and opportunities for crop improvement^[14]. However, in a recent study, the transcriptome profiling of barnyard millet was performed during grain development to reveal the genomic insights into iron accumulation, which has an insight towards genome sequencing^[5,8].

Availability of molecular markers

Molecular markers, specifically nucleotide sequences, play a crucial role in genetic diversity studies, linkage map construction, and marker-assisted selection in crop plants^[9]. Early research utilized RAPD markers to study genetic diversity and phylogeny among *Echinochloa* species, effectively distinguishing between cultivated and wild progenitors^[42,43]. Although RAPD markers provided insights into genetic diversity, their low polymorphism levels were noted in various studies^[44]. Moreover, *Amplified Fragment Length Polymorphism* (AFLP) markers later showed greater efficacy in revealing genetic diversity due to their higher allele production per primer^[5]. Advancements in sequencing technologies introduced sequence-based markers like SRAP *(Sequence-Related Amplified Polymorphism);* SSRs *(Simple Sequence Repeats);* EST-SSRs: *(Expressed Sequence Tag-Simple Sequence Repeats)*; and SNPs, which are more desirable due to co-dominant, reproducible, and highly polymorphic nature[\[18\]](#page-6-17) . Studies utilizing SSR markers in *Echinochloa* species have shown the [fo](#page-7-17)r[m](#page-7-18)ation of distinct clusters, highlighting the genetic diversity^{[[45](#page-7-17)−[48](#page-7-18)]}.

Genetic Improvement of Barnyard Millet

EST markers have proven effective in analyzing genetic diversity, with in silico mining and validation of EST-SSR primers identifying frequent repeat motifs in barnyard millet^{[49–[51\]](#page-7-20)}. Additionally, the RAD *(Restriction-site associated)* approach combined with Illumina DNA sequencing in *E. phyllopogon* facilitated rapid discovery of SSR and SNP markers, demonstrating the usefulness of these markers in studying genetic diversity and aiding molecular breeding^[52]. Further developments in whole-genome *genotyping-by-sequencing* (GBS) have identified a significant number of SNPs across various biotypes of *Echinochloa* species[53] . Moreover, population structure analysis using these SNPs has provided clear genetic differentiation among species, forming distinct clusters^[35]. This detailed SNP data enhances our understanding of genetic diversity, origins, and distribution, particularly in herbicide-resistant populations, and supports molecular breeding efforts^[54]. Recent studies have continued to use EST-SSR markers to analyze genetic diversity in Indian barnyard millet germplasm, reinforcing the importance of advanced molecular markers in crop genetic research and breeding programs^[55]. However, Table 1 summarizes the availability of molecular resources in barnyard millet.

Genetic improvement of barnyard millet

The genetic improvement of barnyard millet is essential for enhancing its productivity, resilience, and nutritional value, making it a key player in sustainable agriculture^[4]. This process involves utilizing advanced breeding techniques such as genomic selection, and biotechnological tools to develop superior cultivars with improved traitsincluding higher yield, enhanced resistance to biotic and abiotic stresses, and better nutritional profiles^[56]. Despite its potential, the genetic improvement of barnyard millet faces challenges like limited genetic diversity, insufficient genomic resources, and inadequate funding for research^[5,8]. Addressing these issues through comprehensive research initiatives and international collaboration can pave the way for the development of robust barnyard millet varieties^[15]. However, Fig 2 illustrates the process of improving Barnyard Millet cultivars by integrating genetic and omics resources. Genetic resources such as core germplasm, recombinant inbred lines, and mutants are utilized alongside genome sequencing to facilitate the identification of molecular markers and genetic improvement techniques like genomic selection and genome editing. Omics resources, including genomics, transcriptomics, and proteomics, play a crucial role in phenotyping, genotyping, and ultimately developing enhanced Barnyard Millet varieties. List consists in the same of the same interest in the same interest intere colour formestic to tracest on enhancement an[d](#page-6-4) a system delay meloc[it](#page-7-23)y b[e](#page-7-7)yond the system of eigenvector and a system delay melocity because the melocity because the system of the system of eigenvector in the system of the

Identification of QTLs and Marker Assisted Selection (MAS)

The identification of QTLs and the application of MAS in barnyard millet have been facilitated by the development of molecular markers such as SSRs^{[\[45,](#page-7-17)[46](#page-7-26)]} and SNPs^{[[52\]](#page-7-21)}. Although many SSR and SNP markers have been created to aid in linkage map construction and QTL mapping, progress has been slower compared to other millets like foxtail and finger millet^{[\[55\]](#page-7-24)}. Notable studies which identified SNP markers for waxy traits controlled by three loci (*EeWx1, EeWx2, EeWx3*) [57] , and linked the SSR marker *BMESSR 39* with anthocyanin pigments using bulk segregant analysis using F_2 individuals of ACM 331 \times MA 10^{[\[58\]](#page-7-28)}, provide opportunities to facilitate the construction of

Table 1. Availability of molecular markers in barnyard millet.

Fig. 2 Process of improving Barnyard Millet cultivars by integrating genetic and omics resources

MAS^[5]. Despite these advancements, barnyard millet genome mapping remains in its initial stages, requiring further inve[st](#page-6-7)i-gation to apply these findings to MAS for improved breeding^{[[8\]](#page-6-7)}.

Genomic selection and speed breeding

Barnyard millet's breeding is poised for a breakthrough with the combined power of genomic selection and speed breeding^{[\[45\]](#page-7-17)}. Genomic selection, unlike traditional methods,

analyzes a plant's entire genetic code to predict valuable traits like yield and disease resistance^{[\[46\]](#page-7-26)}. This allows breeders to identify promising candidates^{[\[55\]](#page-7-24)}. Speed breeding, on the other hand, accelerates the breeding cycle by manipulating growing conditions to produce multiple generations per year^{[\[5](#page-6-4),[8\]](#page-6-7)}. By identifying superior genetics early and rapidly testing them through multiple generations, this combined approach can significantly speed up the development of improved barnyard millet varieties with enhanced yields, disease resistance, and adaptation to local environments^[59,60].

Genome editing

Barnyard millet, despite its potential as a nutritious and climate-smart crop, faces limitations in breeding due to the lack of a complete genome sequence^[61]. While traditional breeding methods are ongoing, genome editing offers a promising but nascent approach^[62]. CRISPR-Cas, a revolutionary gene editing technique, could be a game-changer^[63]. However, its application in barnyard millet is still in its early stages^[61].Moreover, chloroplast genomic data can offer significant advantages when implementing the CRISPR/Cas9-based strategy for genetic improvement in barnyard millet. Firstly, chloroplasts possess unique genetic characteristics that can be exploited for genetic engineering^[64]. Unlike nuclear DNA, chloroplast DNA is maternally inherited in most plants, which can help contain the spread of transgenes via pollen^[63]. This characteristic is advantageous for minimizing gene flow to non-target species and ensuring biosafety in genetically modified crops^[65]. Secondly, the efficiency of chloroplast transformation has traditionally been lower than that of nuclear transformation due to the low rate of homologous recombination in chloroplasts^[63]. Technology in the entire operative contribution in the controller state and the entire of the properties of the properties of the entire o significantly speed to the consisten[ce](#page-8-1) of i[n](#page-8-6)tervals in a plane of the proposition is a powerful associate[d](#page-8-9) of [t](#page-8-8)he consistence of the consistence

However, recent advancements in CRISPR/Cas9 technology have shown that this can be improved^[65]. For instance, using CRISPR/Cas9 to introduce double-strand breaks at specific sites within the chloroplast genome can activate the DNA damage repair mechanism, thereby enhancing the efficiency of homologous recombination. Studies have demonstrated that incorporating CRISPR/Cas9 into the chloroplast transformation system can increase transformation efficiency by 6-10 times, which is a substantial improvement^[65]. Moreover, CRISPR/Cas9 technology allows for precise genetic modifications, including insertions, deletions, and substitutions, which are crucial for targeted genetic improvement^[46].

In the context of barnyard millet, precise editing of the chloroplast genome could be used to introduce traits that enhance photosynthetic efficiency, nutritional value, or stress tolerance, directly impacting crop yield and resilience^[64,66]. Additionally, the use of chloroplast genomic data can facilitate multiplex genome engineering, where multiple genes can be targeted simultaneously^[63]. This is particularly beneficial for complex traits that are controlled by multiple genes^[46]. For example, in cotton, CRISPR/Cas9 has been used to generate mutations in multiple genes concurrently, demonstrating the potential for similar approaches in barnyard millet^[46]. Furthermore, chloroplast-targeted CRISPR/Cas9 applications can improve the nutritional quality of barnyard millet by modifying pathways involved in the synthesis of essential nutrients^[65]. Given that chloroplasts are the site of important biosynthetic pathways, such as those for amino acids and fatty acids, editing the chloroplast genome provides a direct route to enhanc-ing the nutritional profile of the crop^{[[66](#page-8-4)]}. By successfully utilizing CRISPR-Cas, scientists could directly target genes responsible for desirable traits like higher yield, improved drought tolerance, or resistance to specific diseases, and this would significantly accelerate the development of enhanced barnyard millet varieties, but further research is needed to unlock the full potential of genome editing for this promising crop^{[[61](#page-7-33)]}.

Functional genomics, bioinformatics and systems biology approach

Functional genomics is a powerful approach for understanding the roles of genes in complex biological systems and can be highly beneficial for improving crops such as barnyard millet^[67,68]. Applying RNAi techniques to barnyard millet could similarly facilitate the identification of key genes involved in stress responses, growth, and yield^[69,70]. Another critical tool in functional genomics is insertional mutagenesis, such as transfer DNA (T-DNA) tagging, and adapting T-DNA tagging to barnyard millet would enable the systematic characterization of genes, potentially leading to the discovery of traits beneficial for crop improvement^[71]. Functional genomics databases creation for barnyard millet could provide comprehensive resources for storing, querying, and analyzing large-scale data sets, and these databases integrate various types of data, including metabolite profiles and small RNA (sRNA) data, and offer tools for identifying co-expressed genes and significant biological processes^{[\[68](#page-8-6)[,72\]](#page-8-10)}.

In addition to these specific techniques, by comparing the functional genomics data of barnyard millet across different species and model organisms, researchers can gain insights into conserved genetic mechanisms and evolutionary adaptations^[73,74]. Moreover, the integration of bioinformatics and systems biology in barnyard millet research could advancethe understanding of its genetic and molecular underpinnings, facilitating the development of improved crop varieties^{[59[,75\]](#page-8-13)}. Bioinformatics tools could be used to analyze the barnyard millet genome, identify genes, and uncover genetic variations, while systems biology approaches map out complex interactions between genes, proteins, and metabolites^[60,76]. This integration allows for comprehensive analysis of gene expression, regulatory networks, and metabolic pathways^[61,77]. By combining these approaches, researchers can better understand the biological processes underlying key traits, such as stress tolerance and nutrient utilization, and apply this knowledge to optimize breeding strategies and develop resilient, high-performing varieties^[8,8,60,74,78].

Conclusions

Barnyard millet's potential as a nutritious and climate-adaptable crop is undeniable, but genetic improvement hinges on advancements in breeding techniques. QTL identification and Marker-Assisted Selection offer promising avenues to target desirable traits like yield and disease resistance. Additionally, CRISPR-Cas gene editing holds immense promise for precise improvement, although challenges like incomplete genome knowledge and transformation protocols need to be addressed. By overcoming these limitations and actively pursuing these genetic improvement strategies, researchers can unlock barnyard millet's full potential, significantly contributing to global food security and sustainable agriculture.

The authors confirm contribution to the paper as follows: study conception and design: *Jyoti Prakash Sahoo*; data collection: *Pratyush Priyadarshan Pradhan, Priti Bhuyan, Goutam Nag*; analysis and interpretation of results: *Jyoti Prakash Sahoo*; draft manuscript preparation: *Jyoti Prakash Sahoo*; review & editing: *Pratyush Priyadarshan Pradhan, Priti Bhuyan, Goutam Nag*; *Jyoti Prakash Sahoo*. All authors reviewed and approved the final version of the manuscript.

Data availability

Data sharing not applicable to this article as no datasets were generated or analyzed during the current study

Conflict of interest

The authors declare that they have no conflict of interest.

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References

- Gomashe SS. 2017. Barnyard millet: present status and future 1. thrust areas. *Millets and sorghum: biology and genetic improvement* 184−198
- IIMR. 2018. *Annual Report 2017-18. Hyderabad: Indian Institute of* 2. *Millets Research*. https://www.millets.res.in/annual_report/ar17- 18.pdf
- Durairajan MB, Sundararajan VV, Kannan G, Paul BM, Muniyandi K, 3. et al. 2024. Elicitation of nutritional, antioxidant, and antidiabetic potential of barnyard millet (*Echinochloa esculenta* (A. Braun) H. Scholz) sprouts and microgreens through in vitro bio-accessibility assessment. *Food Chemistry* 441:138282 version of the manuscri[p](https://doi.org/10.1111/jfpe.14611)t.

Sectio[n](https://doi.org/10.1007/s42535-022-00420-4) 2.18 (and a mod[e](https://doi.org/https://doi.org/10.1007/978-3-030-41866-3_15)l an[d](https://doi.org/10.1270/jsbbs.56.335) a model and a model
	- 4. Rajesh M, Shivaraj G, Ambethgar V, Vanniarajan C. 2024. Breeding Barnyard Millet for Biotic Stress Resistance. In *Genetic improvement of Small Millets*. pp. 513-528. Singapore: Springer Nature Singapore
	- 5. Padhiyar SM, Kheni J, Bhatt SB, Tomar RS. 2024a. Genetic Improvement of Barnyard Millet Through Advanced Biotechnological Methods. In *Genetic improvement of Small Millets*. pp. 529-553. Singapore: Springer Nature Singapore. https://doi.org/10.1007/97[8-981-99-7232-6_26](https://doi.org/https://doi.org/10.1007/978-981-99-7232-6_26)
	- 6. Mittal V, Kumar S, Gupta R, Panghal A, Kumar N. 2024. Review on millets dehulling and pearling technologies and machinery. *Journal of Food Process Engineering* 47(5):e14611
	- Sood S, Khulbe R, Gupta A, Agrawal PK, Upadhyaya H, et al. 2015. Barnyard millet – a potential food and feed crop of future. *Plant Breeding* 134:135−147 7.
	- Padhiyar SM, Kheni J, Bhatt SB, Desai H, Tomar RS. 2024b. Tran-8. scriptome profiling of barnyard millet (*Echinochloa frumentacea* L.) during grain development to reveal the genomic insights into iron accumulation. *Heliyon* 10
	- 9. Renganathan VG, Vanniarajan C, Senthil N, Nirmalakumari A, Karthikeyan A, et al. 2021. Genetics and molecular markers for anthocyanin pigmentation in barnyard millet (*Echinochloa frumentacea* (Roxb.) Link). *Plant Breeding* 140(2):246−253
	- Malathi VM, Jacob J, Venkateswarlu R, Kannababu N, Ratnavathi 10. CV. 2024. Nutritional Aspects, Phytochemical Composition and Potential Health Benefits of Small Millets. In *Genetic improvement of Small Millets*. pp. 129-152. Singapore: Springer Nature Singapore. [https://doi.org/10.1007/978-981-99-7232-6_7](https://doi.org/https://doi.org/10.1007/978-981-99-7232-6_7)
	- 11. NCBI, 2024. *Echinochloa esculenta*.

www.ncbi.nlm.nih.gov/search/all/?term=Echinochloa+esculenta (Accessed August 08, 2024)

- 12. Nehru G, Reddy AT, Reddy CCM, Sreenivasulu KN. 2021. Assessment of genetic variability in Indian barnyard millet genetic 134 resources [*Echinochloa frumentacea* (L.)]. *[The Journal of Research](https://doi.org/10.5555/20220564567)* 49(4):134−137
- 13. Nozawa S, Takahashi M, Nakai H, Sato Y. 2006. Difference in SSR Variations Between Japanese Barnyard Millet (*Echinochloa esculenta*) and its Wild Relative *E. crus-galli*. *Breeding Science* 56:335−340
- 14. Sood S, Joshi DC, Pattanayak A. 2020. Breeding advancements in barnyard millet. In *Accelerated Plant Breeding, Volume 1: Cereal Crops*, 391-409. https://doi.org/10.1007/978-3-030-41866-3_15
- 15. Sood S, Khulbe R, Saini N, Gupta A, Agrawal P. 2014. Interspecific Hybrid between *Echinochloa esculenta* (Japanese barnyard millet) and *E. frumentacea* (Indian barnyard millet) – A New Avenue for Genetic Enhancement of Barnyard Millet. *Electronic Journal of Plant Breeding* 5:248−253
- 16. Wallace JG, Upadhyaya HD, Vetriventhan M, Buckler ES, Hash C, et al. 2015. The genetic makeup of a global barnyard millet germplasm collection. *The Plant Genome* 8:0067
- 17. Hu X, Liu R, Mao H, Xu Y, Chen B, et al. 2023. Inter-Species Investigation of Biological Traits among Eight *Echinochloa* Species. *[Plants](https://doi.org/10.3390/plants12173085)* 12(17):3085
- 18. Maithani D, Sharma A, Gangola S, Bhatt P, Bhandari G, et al. 2023. Barnyard millet (*Echinochloa* spp.): a climate resilient multipurpose crop. *Vegetos* 36(2):294−308
- 19. Rao AN. 2021. Echinochloa colona and Echinochloa crus-galli. In *Biology and Management of Problematic Crop Weed Species*. pp. 197-239. Academic Press. [https://doi.org/10.1016/B978-0-12-](https://doi.org/https://doi.org/10.1016/B978-0-12-822917-0.00013-6) [822917-0.00013-6](https://doi.org/https://doi.org/10.1016/B978-0-12-822917-0.00013-6)
- Matloob A, Chauhan BS. 2021. Utilization of the neighborhood 20. design to evaluate suitable cover crops and their density for *Echinochloa colona* management. *PLoS One* 16(7):e0254584
- 21. Bhatt P, Kumar V, Rastogi H, Malik MK, Dixit R, et al. 2023. Functional and tableting properties of alkali-isolated and phosphorylated barnyard millet (*Echinochloa esculenta*) starch. *[ACS omega](https://doi.org/10.1021/acsomega.3c03158)* 8(33):30294−30305
- 22. Bhinda MS, Hasan N, Joshi DC. 2023a. Barnyard Millet Improvement: From Pre-genomics to Post-Genomics Era. In *Smart Plant Breeding for Field Crops in Post-Genomics Era*. pp. 255-270. Singapore: Springer Nature Singapore. [https://doi.org/10.1007/978-981-](https://doi.org/https://doi.org/10.1007/978-981-19-8218-7_8) [19-8218-7_8](https://doi.org/https://doi.org/10.1007/978-981-19-8218-7_8)
- 23. Bhinda MS, Joshi DC, Parihar M, Meena RP, Joshi P, et al. 2023b. Genetics, breeding, and genomics of Indian barnyard millet (*Echinochloa frumentacea*): status and perspectives. In *Neglected and Underutilized Crops*. pp. 115-135. Academic Press. https://doi.org/10.1016/B978-0-323-90537-4.00017-X
- 24. Khan R, Bhanu AN, Aneesha N, Sirisha H, Gupta ARSSH, et al. 2024. Floral Biology, Pollination, Genetics, Origin, and Diversity in Barnyard Millet. In *Genetic improvement of Small Millets*. pp. 479-491. Singapore: Springer Nature Singapore. [https://doi.org/10.1007/978-981-99-7232-6_23](https://doi.org/https://doi.org/10.1007/978-981-99-7232-6_23)
- 25. Monteiro P, Sudharshana L, Ramachandra G. 1988. Japanese Barnyard Millet (*Echinochloa frumentacea*): protein content, quality and SDS-PAGE of protein fractions. *Journal of the Science of Food and Agriculture* 43:17−25
- 26. Moharil M, Gawai D, Dikshit N, Dudhare M, Jadhav P. 2016. Assessment of genetic diversity in Indian Barnyard millet (*Echinochloa* spp. complex) using morphological and molecular markers. *[Jour](https://doi.org/10.31018/jans.v8i3.1016)nal of Applied and Natural Science* 8:1643−1648
- 27. Renganathan VG, Vanniarajan C, Nirmalakumari A, Raveendran M, Eshwari T, et al. 2017. Association Analysis in Germplasm and F_2 Segregating Population of Barnyard Millet (*Echinochloa frumentacea* Roxb. Link) for Biometrical and Nutritional Traits. *[Interna](https://doi.org/10.20546/ijcmas.2017.607.406)[tional Journal of Current Microbiology and Applied](https://doi.org/10.20546/ijcmas.2017.607.406) Sciences* 6:3394−3400

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- 28. Wu D, Shen E, Jiang B, Feng Y, Tang W, et al. 2022. Genomic insights into the evolution of *Echinochloa* species as weed and orphan crop. *[Nature communications](https://doi.org/10.1038/s41467-022-28359-9)* 13(1):689
- 29. Yamaguchi H, Utano A, Yasuda K, Yano A, Soejima A. 2005. A molecular phylogeny of wild and cultivated Echinochloa in East Asia inferred from non-coding region sequences of trnT-L-F. *[Weed](https://doi.org/10.1111/j.1445-6664.2005.00185.x) [Biology and Management](https://doi.org/10.1111/j.1445-6664.2005.00185.x)* 5:210−218
- 30. Perumal S, Jayakodi M, Kim D, Yang T, Natesan SA. 2016. The complete chloroplast genome sequence of Indian barnyard millet, *Echinochloa frumentacea* (Poaceae). *Mitochondrial DNA*. *Part B, Resource* 1:79−80
- 31. Gowda J, Bharathi S, Somu G, Krishnappa M, Rekha D. 2009. Formation of core set in barnyard millet [*Echinochloa frumentacea* (Roxb.) Link] germplasm using data on twenty-four morpho-agronomic traits. *International Journal of Plant Sciences Muzaffarnagar* 4: 1-5. http://researchjournal.co.in/online/IJPS/IJPS%204(1)/4_A-1- 5.pdf [A](https://krishikosh.egranth.ac.in/handle/1/5810191808)cc[ep](https://doi.org/https://doi.org/10.1007/978-981-99-7232-6_1)te[d](https://doi.org/10.48130/TIA-2023-0010) to the control of the control of
	- 32. Prabha D, Negi Y, Khanna V. 2012. Assessment of Genetic Diversity of Barnyard Millet Accessions using Molecular Markers. *Indian Journal of Plant Genetic Resources* 25:174−179
	- 33. Maharajan T, Krishna TPA, Krishnakumar NM, Vetriventhan M, Kudapa H, et al. 2024. Role of genome sequences of major and minor millets in strengthening food and nutritional security for future generations. *Agriculture* 14(5):670
	- 34. Mohanapriya B, Shanmugam A, Francis N, Indhu SM, Ravikesavan R. 2024. Breeding Barnyard Millet for Abiotic Stress Tolerance. In *Genetic improvement of Small Millets*. pp. 493-511. Singapore: Springer Nature Singapore. https://doi.org/10.1007/978-981-99- 7232[-](https://doi.org/https://doi.org/10.1007/978-981-99-7232-6_24)6_24
	- 35. Babu BK, Chauhan R. 2017. In-Silico Identification of EST Based Microsatellite Markers and SNPs, and Comparative Genomic Analysis of ESTs in Barnyard Millet for their Omics Applications. *Current Agriculture Research Journal* 5(3):279
	- 36. Nandini C, Bhat S, Saritha HS, Pandey CD, Sushil Pandey P, et al. 2020. Characterization of barnyard millet (*Echinocloa frumentaceae* (Roxb.) Link) germplasm for quantitative traits to enhance its utilization. *Electronic Journal of Plant Breeding* 11(04):1066−1072
	- 37. Gupta A, Joshi D, Mahajan V, Gupta HS. 2009. Screening barnyard millet germplasm against grain smut (*Ustilago panici-frumentacei Brefeld*). *Plant Genetic Resources* 8:52−54
	- 38. Williams G, Vanniarajan C, Vetriventhan M, Thiageshwari S, Anandhi K, et al. 2019. Genetic variability for seedling stage salinity tolerance in barnyard millet [*Echinochloa frumentaceae* (Roxb.) Link]. *Electronic Journal of Plant Breeding* 10(2):552−558
	- 39. Dhanalakshmi R, Subramanian A, Thirumurugan T, Elangovan M, Kalaimagal T. 2019. Genetic variability and association studies in barnyard millet (*Echinochloa frumentacea* (Roxb.) Link) germplasm under sodic soil condition. *Electronic Journal of Plant Breeding* 10(2):430−439
	- Elangovan M, Venkatesh K. 2024. Small Millets Genetic Resources 40. Management. In *Genetic improvement of Small Millets*. pp. 1-16. Singapore: Springer Nature Singapore. https://doi.org/10.1007/97[8-981-99-7232-6_1](https://doi.org/https://doi.org/10.1007/978-981-99-7232-6_1)
	- Arthi N, Rajagopal B, Geethanjali S, Nirmalakumari A, Senthil N. 41. 2019. Screening of barnyard millet (*Echinochloafrumentacea*) germplasm for salinity tolerance. *Electronic Journal of Plant Breeding* 10(2):659−666
	- Jethava KM. 2021. *Molecular marker based genetic diversity analysis* 42. *in barnyard millet (Echinochloa frumentacea L.) 3325* (Doctoral dissertation, JAU, Junagadh). https://krishikosh.egranth.ac.in/handle/1/5810191808
	- 43. Roy B, Tiwari S, Tripathi N, Thakur VV. 2024. Molecular Diversity Analysis Among Indian Barnyard Millet [*Echinochloa frumentacea* (L)] Genotypes Using RAPD Markers. *Journal of Global Agriculture [and Ecology](https://doi.org/10.56557/jogae/2024/v16i18506)* 16(1):1−7
	- 44. Desai A, Dakappa SS. 2020. Genetic Diversity Analysis of Millet Crop Species of *Panicum*Genus Using Rapd Markers. *World Journal*

of Pharmaceutical Research 9(5):1132−1141

- 45. Cusaro CM, Grazioli C, Zambuto F, Capelli E, Brusoni M. 2021. An improved method for assessing simple sequence repeat (SSR) variation in *Echinochloa crus-galli (*L. *)* P. Beauv (Barnyard grass). *[Diver](https://doi.org/10.3390/d14010003)[sity](https://doi.org/10.3390/d14010003)* 14(1):3
- 46. Gao Y, Shen G, Yuan G, Tian Z. 2022. Comparative analysis of whole chloroplast genomes of three common species of *Echinochloa* (Gramineae) in paddy fields. *[International Journal o](https://doi.org/10.3390/ijms232213864)f Molecular Sciences* 23(22):13864
- 47. Dineshbhai CN. 2023. Assesment of molecular diversity and proxi*mate composition in millets*. [https://krishikosh.egranth.ac.in/server/api/core/bitstreams/75bf0b](https://krishikosh.egranth.ac.in/server/api/core/bitstreams/75bf0b10-b2ce-43f9-aaf1-74f2a077aa2b/content) [10-b2ce-43f9-aaf1-7](https://krishikosh.egranth.ac.in/server/api/core/bitstreams/75bf0b10-b2ce-43f9-aaf1-74f2a077aa2b/content)4f2a077aa2b/content
- 48. Meniya VH, Padhiyar SM, Desai H, Kheni JK, Tomar RS. 2023. Development and validation of microsatellite markers for Barnyard Millet obtained by partial genome assembly. *[Annals of Arid Zone](https://doi.org/10.59512/aaz.2023.62.1.9)* 62(1):83−89
- 49. Sahoo JP, Mahapatra M. 2023. International year of millets-2023: Revitalisation of millets towards a sustainable nutritional security. *Technology in Agronomy* 3(1):10
- 50. Jayakodi M, Madheswaran M, Adhimoolam K, Perumal S, Manickam D, et al. 2019. Transcriptomes of Indian barnyard millet and barnyard grass reveal putative genes involved in drought adaptation and micronutrient accumulation. *Acta Physiologiae Plantarum* 41. https://doi.org/10.1007/s11738-019-2855-4
- 51. Renganathan VG, Vanniarajan C, Karthikeyan A, Ramalingam J. 2020. Barnyard Millet for Food and Nutritional Security: Current Status and Future Research Direction. *Frontiers in genetics* 11:500
- 52. Chen G, Zhang W, Fang J, Dong L. 2017. Identification of massive molecular markers in *Echinochloa phyllopogon* using a restrictionsite associated DNA approach. *Plant diversity* 39(5):287−293
- Mundada PS, Kadam SB, Pable AA, Barvkar VT. 2022. Recent 53. Advances and Applicability of GBS, GWAS, and GS in Millet Crops. *Genotyping by Sequencing for Crop Improvement* 270−294
- 54. Babu BK, Rashmi C, Sood S. 2018. Cross transferability of finger millet and maize genomic SSR markers for genetic diversity and population structure analysis of barnyard millet. *Indian Journal of Genetics and Plant Breeding* 78(3):364−372
- 55. Parveen R, Kumar M, Swapnil N, Singh D, Shahani M, et al. 2023. Understanding the genomic selection for crop improvement: current progress and future prospects. *Molecular Genetics and Genomics* 298(4):813−821
- 56. Sao, A, Pali V, Patil HE. 2024. Genetic Improvement for Yield, Quality, Biotic, and Abiotic Stresses in Little Millet (*Panicum sumatrense* Roth. ex Roem. and Schult.). In *Genetic improvement of Small Millets* (pp. 571-599). Singapore: Springer Nature Singapore. https://doi.org/10.1007/978-981-99-7232-6_28
- 57. Ishikawa G, Seimiya Y, Saito M, Nakamura T, Hoshino T. 2013. Molecular characterization of spontaneous and induced mutations in the three homoeologous waxy genes of Japanese barnyard millet [*Echinochloa esculenta* (A. Braun) H. Scholz]. *[Molecular](https://doi.org/10.1007/s11032-012-9769-9) breeding* 31:69−78
- 58. Pradhan J, Sahoo JP, Samal KC, Dash M. (Eds.). 2024. Millets ana *Other Potential Crops: Ensuring Climate Resilience and Nutritional Security (1st ed.)*. CRC Press. [https://doi.org/10.1201/9781003531937](https://doi.org/https://doi.org/10.1201/9781003531937)
- 59. Kayastha S, Sahoo JP, Mahapatra M, Panda N. 2023. Understanding the molecular breeding and omics approaches for finger millet (*Eleusine coracana* L.) improvement towards a global sustainable nutritional security. *Crop Design* 3(1):100049
- 60. Kayastha S, Sahoo JP, Mahapatra M, Sharma SS. 2024. Finger millet (*Eleusine coracana*) enhancement through genomic resources and breeding methods: current implications and potential future interventions. *Planta* 259(6):139
- 61. Sapara V, Khisti M, Yogendra K, Reddy PS. 2024. Gene editing tool kit in millets: present status and future directions. *[The Nucleus](https://doi.org/10.1007/s13237-024-00485-3)* 67:157−179

- 62. Ceasar A. 2022. Genome-editing in millets: current knowledge and future perspectives. *[Molecular Biology Reports,](https://doi.org/10.1007/s11033-021-06975-w)* 49(1):773−781
- 63. Tang N, Xia Y, Zhan Y, Dan J, Yu M, et al. 2020. Improvement of chloroplast transformation using CRISPR/Cas9. *[Journal of Biobased](https://doi.org/10.1166/jbmb.2020.1970) [Materials and Bioenergy](https://doi.org/10.1166/jbmb.2020.1970)* 14(3):401−407
- 64. Arora L, Narula A. 2017. Gene editing and crop improvement using CRISPR-Cas9 system. *[Frontiers in plant science](https://doi.org/10.3389/fpls.2017.01932)* 8:1932
- 65. Karkute SG, Singh AK, Gupta OP, Singh PM, Singh B. 2017. CRISPR/Cas9 mediated genome engineering for improvement of horticultural crops. *Frontiers in plant science* 8:1635
- El-Mounadi K, Morales-Floriano ML, Garcia-Ruiz H. 2020. Principles, 66. applications, and biosafety of plant genome editing using CRISPR-Cas9. *Frontiers in plant science* 11:56
- 67. Buza T, McCarthy F. 2013. Functional genomics: applications to production agriculture. *Cab Reviews: Perspectives in Agriculture, Veterinary Science, Nutrition and Natural Resources* 8:54
- 68. Ajeesh KTP, Maharajan T, Ceasar SA. 2022. Improvement of millets in the post-genomic era. *Physiology and molecular biology of plants* 28(3):669−685
- 69. Matthew L. 2004. RNAi for Plant Functional Genomics. Compara*tive and Functional Genomics* 5:240−244
- 70. Dhaka A, Singh RK, Muthamilarasan M, Prasad M. 2021. Genetics and genomics interventions for promoting millets as functional foods. *Current Genomics* 22(3):154
- 71. Lo S, Fan M, Hsing Y, Chen L, Che S, et al. 2016. Genetic resources offer efficient tools for rice functional genomics research. *Plant, cell & environment* 39(5):998−1013
- Fei Z, Joung J, Tang X, Zheng Y, Huang M, et al. 2010. Tomato 72. Functional Genomics Database: a comprehensive resource and

analysis package for tomato functional genomics. *[Nucleic Acids](https://doi.org/10.1093/nar/gkq991) [Research](https://doi.org/10.1093/nar/gkq991)* 39:D1156−D1163

- 73. Sato F. 2005. RNAi and functional genomics. [Plant Biotechnology](https://doi.org/10.5511/plantbiotechnology.22.431) 22:431−442
- 74. Powell F. 2003. Functional genomics and the comparative physiology of hypoxia. *[Annual review of physiology](https://doi.org/10.1146/annurev.physiol.65.092101.142711)* 65:203−230
- 75. Gao W, Long L, Tian X, Xu F, Liu J, et al. 2017. Genome editing in cotton with the CRISPR/Cas9 system. *[Frontiers in plant scien](https://doi.org/10.3389/fpls.2017.01364)ce* 8:1364
- 76. Rana S, Tiwari S, Gupta N, Tripathi MK., Tripathi N, et al. 2023. Validating the Nutraceutical Significance of Minor Millets by Employing Nutritional–Antinutritional Profiling. *Life* 13(9):1918
- 77. Sun M, Yan H, Zhang A, Jin Y, Lin C, et al. 2023. Milletdb: a multiomics database to accelerate the research of functional genomics and molecular breeding of millets. *Plant Biotechnology Journal* 21:2348−2357
- 78. Supritha Raj DS, Ragi S, Pattanashetti BM, Mendapera I. 2024. Breeding Proso Millet (*Panicummiliaceum* L.) for Abiotic Stress Resistance. In *Genetic improvement of Small Millets*. pp. 443-453. Singapore: Springer Nature Singapore. [https://doi.org/10.1007/978-981-99-7232-6_20](https://doi.org/https://doi.org/10.1007/978-981-99-7232-6_20)

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