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Effects of habitat fragmentation on the coastal *Vatica mangachapoi* forest (Dipterocarpaceae) in Shimei Bay, Hainan Island, China

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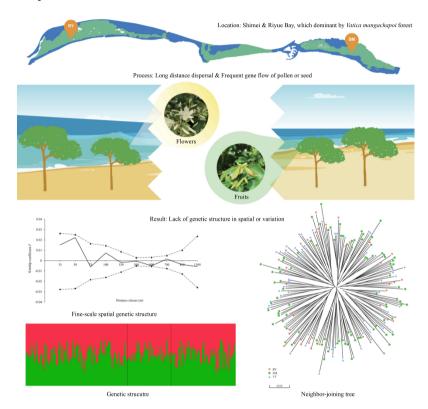
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In Brief

The coastal *Vatica mangachapoi*dominated forest growing on barren sand substrate in Shimei Bay, Hainan Island is quite a special tropical forest ecosystem, which plays important role in defense of coast. However, it has become fragmented due to logging, expansion of farmland and villages, and construction of tourism facilities. Conservation genetic study can shed light on the protection and sustainable management of this unique tropical coastal *V. mangachapoi*-dominated forest.

Graphical abstract



Highlights

- There is no significant difference in genetic diversity between the coastal and lowland *V. mangachapoi* populations.
- No fine-scale spatial genetic structure was detected in the coastal SM population.
- Strong gene flow between populations accounts for the homogenous genetic structure of the coastal and lowland populations.
- There is not enough time to accumulate differentiation between fragmented populations in the coastal forest.

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Effects of habitat fragmentation on the coastal Vatica mangachapoi forest (Dipterocarpaceae) in Shimei Bay, Hainan Island, China

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Abstract

Habitat fragmentation can cause isolation and decline of a formerly continuously distributed population, which leads to loss of genetic variation and increased risk of extinction. Vatica mangachapoi Blanco is a dominant tree species growing in the lowland rainforests of Hainan Island, China. Remarkably, this species dominates a coastal forest in Shimei Bay, Wanning City of Hainan Province (China). Due to logging, expansion of farmland and villages, and construction of tourism facilities, the coastal V. mangachapoi-dominated forest has become fragmented, threatening its future. To evaluate the effects of habitat fragmentation on this unique coastal forest, two V. mangachapoi populations (SM and RY) along the coast and one population in the lowland rainforest near the coast were selected, and their genetic diversity was assessed based on 12 SSR markers. In addition, the genetic structure of the three populations and gene flow among them, and the fine-scale spatial genetic structure (FSGS) of the SM population were also studied. The results show that the three V. mangachapoi populations had comparable levels of genetic variation, and differentiation among them is negligible ($F_{st} = 0.008 \sim 0.013$). Model-based clustering, Principal co-ordinate analysis and the Neighborjoining (NJ) methods consistently support a homogeneous genetic structure of the three populations, and strong gene flow was detected among them by MIGRATE analyses. Moreover, there is no significant FSGS in the SM population. A relatively short time since habitat fragmentation and gene flow mediated by seed dispersal might be the likely reasons for the high levels of genetic variation and an absence of genetic structure of the coastal V. mangachapoi populations. In conclusion, even though there are no significant effects of fragmentation on the coastal V. mangachapoi forest, strict protection is required to prevent further deforestation and fragmentation. Besides, saplings of V. mangachapoi should be planted in forest gaps to reconnect fragments of the coastal forest, which would be of benefit for the long-term survival of the tropical coastal V. mangachapoi-dominated forest.

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Introduction

Trees from family Dipterocarpaceae serve an important ecosystem function in the rainforest community of Asian tropical forests, where 20%–50% of the canopy layer belong to this family^[1]. With high-quality timber that has a high economic value, dipterocarp forests also form a major pillar of the global tropical timber trade^[2]. Due to long-term over-harvesting and land use change, tropical rainforests have become severely fragmented, and a large number of dipterocarps are today listed as endangered species and are at risk of extinction^[3]. As a result, ecosystem services of Asian tropical rainforests in which dipterocarps are the dominant species have been seriously impaired^[4]. Therefore, conservation genetics studies focusing on Dipterocarpaceae are urgently needed^[1].

The tropical forests in Hainan Island, China are located at the northern edge of tropical Asia and are distinct from the typical tropical rainforests of Southeast Asia in terms of species composition, community structure and appearance as a consequence of the influence of the Asian monsoon^[5]. Only three species of Dipterocarpaceae, *Hopea hainanensis* Merrill & Chun, *H. reticulata* Tardieu and *Vatica mangachapoi* Blanco can be found on this island. Although the species diversity of Dipterocarpaceae in Hainan Island has been greatly reduced as

assembly and ecosystem functioning of the lowland rainforests of this island^[5-7]. It is remarkable that V. mangachapoi has developed into a continuously distributed coastal forest growing on sand substrate with 25 kilometers-long and 400 to 500 meters-wide at Shimei Bay, Wanning City (China), which is estimated to be at least 4000 years old^[8,9]. A study showed that soil moisture and organic matters of the sand substrate are much lower than those of normal tropical soils^[10]. The formation of the coastal V. mangachapoi-dominated forest on barren and harsh sandy beach is unique and rare in itself, which could serve as an example to study the underlying physiological and genetical adaptation of V. mangachapoi to arid and poor substrate. In recent years, due to such factors as coastal development and village expansion, the area of the coastal V. mangachapoi-dominated forest in Shimei Bay has been reduced, and the formerly intact population has fragmented into several isolated patches. Coupled with the presence of forest gaps and fungal disease caused by human interference, the survival of the coastal V. mangachapoi is seriously threatened^[9,11,12]. Conservation management is thus needed to protect this unique coastal forest dominated by mangachapoi.

compared to that in Southeast Asia, the three species, espe-

cially V. mangachapoi (Fig. 1), play a key role in community



Fig. 1 Morphology of Vatica mangachapoi. (a) An individual tree, (b) flowers, (c) fruits.

Habitat fragmentation can cause an intact population into small, isolated patches, with reduced gene flow between patches and increased genetic drift and inbreeding within patches^[13–16]. If seed dispersal is limited and restricted within patches, genotypes are likely to be spatially clustered, producing a strong fine-scale spatial genetic structure (FSGS)^[17,18]. Vatica magachapoi has winged fruits, which may promote seed dispersal by wind. Studies indeed showed that the dispersal distance of winged fruits are generally further than non-winged fruits in dipterocarps^[19,20]. On the other hand, the strength of FSGS was also affected by pollen dispersal. Dipterocarps that are pollinated by large insects, such as bees, can achieve longer distance of pollen flow than those pollinated by small insects, such as thrips, because large pollinators can move further than small ones^[21,22]. The limited seed and pollen dispersal were confirmed as the main reason for significant FSGS of most dipterocarps in fragmented habitats^[23–26]. Is there a significant FSGS in the coastal forest of V. mangachapoi? Is genetic diversity lower in the coastal V. mangachapoi populations than the undisturbed rainforest populations nearby? Does significant genetic differentiation occur between patches of the coastal V. mangachapoi forest? These questions are important for the conservation and management of the unique coastal dipterocarp forest but remain to be resolved.

To answer the above questions, two coastal populations of *V.* mangachapoi (SM and RY) were sampled, and one population in the lowland rainforest near the coast (TT) were further collected for comparison. Genetic diversity, structure and population differentiation were assessed for the three *V.* mangachapoi populations using 12 SSR markers. Gene flow was estimated to test whether habitat fragmentation interrupted genetic exchange between them or not. Finally, FSGS were analyzed using the SM population to show whether significant spatial genetic structure has occurred within patches. Answering these questions could shed light on the conservation and continued survival of this unique coastal *V.* mangachapoi-dominated forest.

Materials and methods

Sample collection

The study area is located in the Provincial Nature Reserve of *V. mangachapoi*, Wanning City, Hainan Province (China). Two *V. mangachapoi* populations (SM and RY) in the coastal forest separated by villages, roads and human facilities, and one population (TT) in the lowland rainforest near the coast were selected (Table 1, Fig. 2). In total, 188 *V. mangachapoi* trees individually spaced out more than 25 m apart and with DBH > 5 cm were sampled. Mature leaves lacking disease spots were selected, dried by silica gel and then were stored in a -20 °C refrigerator. The voucher specimens of *V. mangachapoi* were kept in Hainan University (Hainan, China).

Genomic DNA extraction, PCR amplification and SSR genotyping

A modified CTAB method^[27] was used to extract the genomic DNA of *V. mangachapoi*. Twelve pairs of polymorphic SSR primers developed by Guo et al.^[28] were used in this study. PCR amplification were performed in a total volume of 10 μ L, containing 1.0 μ L of genomic DNA (around 50 ng), 5.0 μ L of Taq PCR Master Mix (GeneTech), 0.5 μ L of forward and reverse primers, and 3.0 μ L of ddH₂O. Amplification was carried out as follows: pre-denaturation at 95 °C for 5 min, followed by 35 cycles of denaturation at 95 °C for 20 s, annealing at 52–62 °C for 15 s, extension at 72 °C for 30 s, and finally extension at 72 °C for 7 min. PCR products were separated by capillary electrophoresis using ABI3730xl (Applied Biosystem) and SSR genotypes were analyzed by the GeneMarker software.

Data analysis

Population genetic parameters, including number of alleles (N_a) , effective number of alleles (N_e) , observed heterozygosity (H_o) , expected heterozygosity (H_e) , inbreeding coefficient (F_{is}) and genetic differentiation among populations (F_{st}) wereestimated using GenAlex 6.51^[29]. Polymorphism information content (*PIC*) and Nei & Chessers^[30] genetic distances were

Genetic divesrity and FSGS of Vatica mangachapoi

Table 1. Genetic diversity indices of the three V. mangachapoi populations based on 12 SSR markers.

Population	Location	Ν	Na	N _e	H _o	H _e	F _{is}
SM	110.26691° E, 18.66671° N	91	8	3.647	0.547	0.690	0.207
RY	110.17952° E, 18.59768° N	39	7	3.704	0.605	0.700	0.142
Π	110.24941° E, 18.67744° N	58	7.5	3.694	0.566	0.692	0.167
Average			7.5	3.682	0.572	0.694	0.172

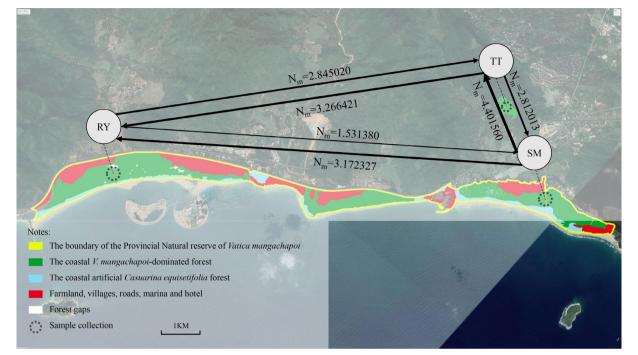


Fig. 2 Geographic distribution of the coastal *V. mangachapoi*-dominated forest and the locations of the three sampled *V. mangachapoi* populations. Gene flow among them was estimated, with the width of lines being proportional to the intensity of gene flow.

calculated by PowerMarker 3.25^[31]. Analysis of molecular variance (AMOVA) of the *V. mangachapoi* population was performed using Arlequin 3.5^[32]. Potential population bottleneck was examined by Wilcoxon sign-rank test and model shift using the Stepwise Mutation Model and Two-phased Mutation Model implemented in Bottleneck 1.3.2^[33].

Bayesian clustering analysis was performed using Structure 2.3.4^[34]. The values of *K* were set from 1 to 10, and for each value of *K*, 10 independent replicates were run with 100,000 burn-in iterations followed by 200,000 MCMC (Markov chain Monte Carlo) iterations. The best *K* was determined according to the delta *K* of STRUCTURE Harvester^[35]. The results of the 10 replicates were combined by the Greedy algorithm implemented in Clumpp1.1.2^[36], and the result of individual clustering were drawn using Distruct1.1^[37]. NJ trees were constructed using MEGA 11.0^[38] based on Nei & Chessers^[30] genetic distances. Principal co-ordinate analysis (PCoA) was performed with GenAlex 6.51.

The effective population size (θ) of the three *V. mangachapoi* populations and the migration rate (*M*) between them were calculated using MIGRATE^[39], a software based on the coalescent theory and Bayesian inference to estimate values of parameters of a user-specified population model. MIGRATE analyses were run under a Brownian motion model, and four heat chains with different temperatures of 1.0, 1.5, 3.0 and 1.0×10^5 were simulated. Gene flow was calculated according

to the equation $Nm = \theta^*M/x$. For SSR markers, x was set to 4. Three independent replications were run to ensure the convergence of the Markov chain Monte Carlo methods implemented in MIGRATE.

The fine-scale spatial genetic structure (FSGS) within-population was assessed using SPAGeDi $1.5^{[40]}$. The kinship coefficients (F_{ij} , kinship coefficients) between any two individuals were calculated and regressed against the natural logarithm of the spatial distance to obtain the regression slope $b_F^{[41]}$. We divided the distance between any pair of individuals sampled from the SM population into 10 distance classes (35, 50, 75, 100, 150, 300, 500, 700, 850, 1,100 m), with at least 30 pairs of individuals per distance class^[42,43].

The 95% confidence intervals of F_{ij} were calculated from 9,999 permutations of spatial distance among pairs of adults for 10 distance classes. If the F_{ij} was higher than the upper bound of the 95% confidence interval, there is significant spatial genetic structure in population and a high level of genetic similarity among individuals; if the F_{ij} fell within the 95% confidence interval, there is no spatial genetic structure in population and individuals were considered to be spatially randomly distributed; if the F_{ij} was less than the lower bound of the 95% confidence interval, individuals were considered to be uniformly distributed in space without spatial genetic structure. The value of the Sp statistic reflects the strength of FSGS and is defined as $Sp = -b_F / (1-F_{(1)})$, where b_F is the regression slope,

and $F_{(1)}$ is the mean pairwise kinship coefficient of the first distance class.

Results

Genetic diversity, differentiation and bottleneck test of the three *V. mangachapoi* populations

There is no significant difference in the level of genetic diversity between the coastal (SM and RY) and the rainforest (TT) populations (Table 1). The inbreeding coefficient was greater than 0, indicating inbreeding and an excess of homozygotes in the three *V. mangachapoi* populations. Totally 90 alleles were detected from the 12 SSR loci, and the number of alleles at a single locus ranged from 4.667 to 11.000, with an average of 7.500 alleles per locus. The observed and expected heterozygosity ranged from 0.303 to 0.769 and from 0.415 to 0.808, respectively. The primer sequences, range of allele sizes and genetic diversity indices of the 12 SSR loci are shown in Table 2. Genetic differentiation was weak among the three *V*. *mangachapoi* populations ($F_{st} = 0.008 \sim 0.013$). The result of AMOVA showed that 99% of genetic variation was partitioned within population, in line with little divergence among populations (Supplemental Table S1).

The Wilcoxon sign rank test found that the *p*-values were not significant under either the S.M.M or the T.P.M model for the three *V. mangachapoi* populations, and their allele frequency distributions were generally *L*-shaped (Fig. 3), indicating that the three populations have not experienced genetic bottle-necks recently.

Population genetic structure

STRUCTURE analysis found that delta *K* was maximized at K = 2, indicating two genetic clusters of the studied *V*. *mangachapoi* populations. The distribution of the two clusters did not differ significantly between the three populations, and this is also true for K = 3 or 5 (Fig. 4). Consistent with the results of STRUCTURE analyses, NJ tree (Supplemental Fig. S1) and PCoA analysis (Fig. 5) also suggest a homogeneous genetic structure of the three *V. mangachapoi* populations.

Table 2. Primer sequences, allele size and genetic diversity indices of the 12 SSR markers.

Loci	Primer sequences (5'-3')	Repeat motif	Allele size	GenAlex				PowerMarker
				Na	N _e	H _o	H _e	PIC
VM1	F:GAACCCTTATTGGCCTGCCTAC	(AT) ₁₁	166–184	7.333	4.231	0.740	0.763	0.7430
	R:GGGACCAAATGACTTGAGTAATCT							
VM2	F:ACCCTAACAATTCTCTTTGTTTCCT	(TAA) ₁₁	152–195	9.667	4.120	0.513	0.755	0.7364
	R:CCCCAATCTCAGTAAGGACTCA							
VM3	F:CTTGTGTCGAGCATGCATGTAT	(AT) ₁₁	175–191	8.333	4.857	0.761	0.793	0.7659
	R:TGCTGGCCTTTTATGTTAGGGT							
VM4	F:ATAGCAGGCACTTCGGAAGTAC	(TA) ₈	261-277	8.667	4.613	0.370	0.781	0.7533
	R:CCTGAGAAACAAAGCAACGCAT							
VM5	F:GCACTAGCACTAGCACTAGCTT	(CT) ₁₁	218-226	4.667	2.908	0.629	0.651	0.6026
	R:GGCTTTTCCAATTTCCATGGCT							
VM6	F:AGTTAAGGGACCAAATTTAGCGT	(TA) ₇	259–269	5.000	2.794	0.593	0.636	0.5902
	R:GTGTTTGTCAACTGGGCTTCAA							
VM7	F:CCCATGTGCTAGGCTAATGCTA	(AT) ₆	229–239	5.000	2.394	0.303	0.582	0.5409
	R:AAATCAGCATGAAACTTCTCCATT							
VM8	F:CACCACCACAGGCTTGAGTATA	(TA) ₇	168–182	5.667	1.722	0.374	0.415	0.4044
	R:GAAGGCCAACTAATCAAGCTGC							
VM9	F:TCATTTCTGTCTCACTCGACCC	(TTC) ₁₀	148–168	5.667	3.010	0.639	0.666	0.6097
	R:TCATCGACGAATCACTGTTCGA							
VM10	F:ACGGATAAGTTAACGGACTAGACA	(TA) ₁₀	215-227	9.333	4.713	0.568	0.776	0.7997
	R:AGATTTTCCCCCAGTCATCGAC							
VM11	F:GCTGGCACTTAGGATGCCTTAA	(ATT) ₁₁	138–150	11.000	3.564	0.610	0.702	0.6657
	R:AGCAACCAATTAGCTCAAATCAA							
VM12	F:GGGCAGCCTCGTAAATCAATTAC	(ATT) ₁₃	225-249	9.667	5.253	0.769	0.808	0.7958
	R:ATTACCTGGCACAACCTTAGCC							

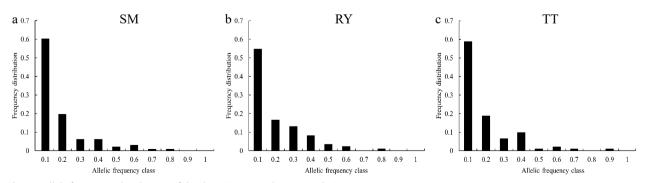


Fig. 3 Allele frequency distribution of the three V. mangachapoi populations.

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Genetic divesrity and FSGS of Vatica mangachapoi

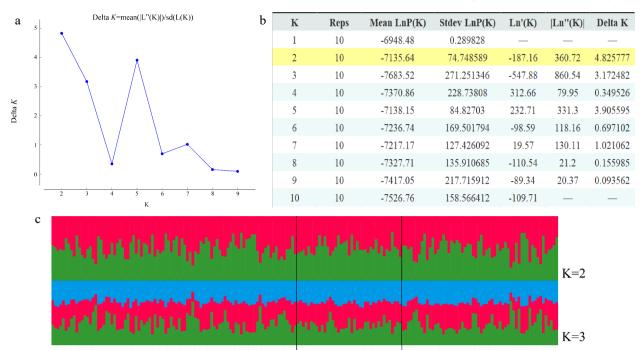
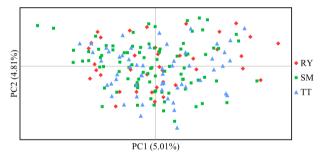


Fig. 4 Results of STRUCTURE analysis. (a) Best K determined using the delta K method. (b) Log probabilities and delta K values for K from two to ten. (c) The results of individual assignment at K = 2, 3 and 5. Each vertical bar represents an individual, and the proportion of the colors corresponds to the posterior probability of genetic clusters assigned to each individual.

RY



SM

Fig. 5 Principal co-ordinate analysis (PCoA) based on Nei & Chessers^[30] genetic distance among individual samples of V. mangachapoi.

Table 3. Mutation-scaled migration rate, effective population size and gene flow estimated by program MIGRATE.

Direction of gene flow	Migration rate (<i>M</i>)	Effective population size (θ)	Gene flow (N _m)
SM→RY	129.615	$\theta_{SM} = 0.09790$	3.172327
SM→TT	179.839		4.401560
RY→SM	63.241	$\theta_{\rm RY} = 0.09686$	1.531380
RY→TT	117.490		2.845020
TT→SM	115.412	$\theta_{\rm TT} = 0.09746$	2.812013
TT→RY	134.062		3.266421

Gene flow and effective population size

The effective population sizes of the three *V. mangachapoi* populations estimated by MIGRATE were similar, but the intensity of gene flow varied among them (Fig. 2, Table 3). The gene

flows from RY to the other two populations were less than their reverse gene flows, however, the gene flows from SM to the other two populations were greater than their reverse gene flows. These results suggested that gene flows between the *V. mangachapoi* populations were asymmetric.

TT

Fine-scale spatial genetic structure

No spatial genetic structure was detected at any of the 10 distance classes in the SM population (Fig. 6). The values of F_{ij} were less than zero over multiple distance classes, indicating that individual trees of *V. mangachapoi* were spatially uniformly distributed. Based on the mean affinity ($F_{(1)}$) for the first distance class (0.0151) and the regression slope b_F (-0.004605), the strength of FSGS (*Sp*) was derived as 0.004675 for the *V. mangachapoi* population in Shimei Bay.

Discussion

Genetic diversity, population structure and gene flow

Due to rapid coastal development and village expansion, the coastal population of *V. mangachapoi* declined and fragmented into several small and discontinuous patches^[12] (Fig. 2). However, there was no significant difference in genetic diversity between the coastal (SM and RY) and the rainforest (TT) populations (Table 1), and the effective sizes of the three populations are quite close (Table 3). Besides, no recent bottleneck could be detected in these populations (Fig. 3). The results indicated that the sizes of the two coastal populations are still large to maintain a comparative level of genetic diversity relative to

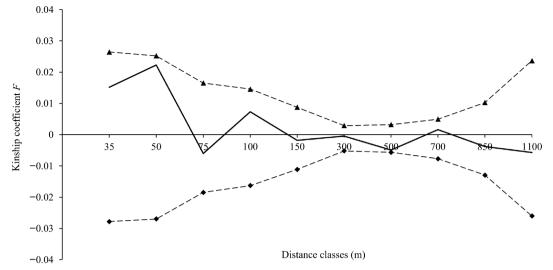


Fig. 6 Fine-scale genetic structure of *V. mangachapoi* in Shimei Bay. The solid line represents the mean Kinship coefficient *F* (Loiselle et al.^[41]), and the dashed lines represent the 95% confidence intervals of the mean Kinship coefficient *F*.

that of the rainforest population nearby^[44]. In addition, the three *V. mangachapoi* populations were demonstrated to share a homogenous genetic structure and there is little differentiation between them (Figs 4 & 5). In summary, patterns of SSR variation observed in *V. mangachapoi* suggested either genetic connection through gene flow or not enough time to accumulate divergence after fragmentation of the coastal forest dominated by *V. mangachapoi*.

Frequent gene flow can prevent rapid loss of genetic variation and differentiation between patches in a fragmented population^[45,46]. Vateriopsis seychellarum is endemic in the Seychelles, after a long period of logging, only a few hundred adult trees remained. Nevertheless, a relatively high level of genetic diversity was found in this species. Long-distance gene flow between isolated patches of Va. seychellarum was considered as the main reason to maintain genetic variation in this species^[25]. If pollinators can travel across the gaps created by fragmentation, pollen-mediated gene flow could be maintained between patches, as a result, genetic drift happened within patches would be mitigated in the short term^[26,47,48]. In this study, frequent gene flow (Nm > 1) was detected between the three V. mangachapoi populations. Besides, the time of fragmentation of the coastal V. mangachapoi forest is relatively short comparing with the generation time of this species. Differentiation between patches is probably impeded by gene flow or there is not enough time to accumulate significant divergence among them, and genetic variation could be largely maintained within the coastal V. mangachapoi forest.

Fine-scale spatial genetic structure

No significant FSGS was detected in the SM population of *V. mangachapoi*, which may result from long distance dispersal of seeds and/or pollens within population^[24,49]. Two of the five sepals of *V. mangachapoi* flowers keep growing and develop into functional wings of the fruits that would promote seed dispersal by wind^[19,50]. Hainan Island will experience frequent Pacific typhoon from June to October each year, and the time of fruit ripening of *V. mangachapoi* (mid to late July-August) coincides well with the activities of the Pacific typhoon^[51]. Moreover, Wanning City is one of the locations where typhoons frequently make landfall on Hainan Island. The coastal *V.*

mangachapoi forest at Shimei Bay would be highly likely to encounter a Pacific typhoon during its fruit ripening period. Strong convection currents can carry winged fruits of *V*. *mangachapoi* into the upper air and achieve a long-distance horizontal dispersal with hundreds of meters, which may account for the lack of significant FSGS in the SM population^[52,53].

Pollen-mediated gene flow can also influence the strength of FSGS, and restricted gene flow generally results in significant FSGS^[22,24,54]. Kettle et al. studied the FSGS of three dipterocarp species from the tropical rainforests at Borneo^[54]. No clear signal of FSGS was detected in Dipterocarpus grandiflorus, a species pollinated by large pollinators with strong mobility, which may facilitate long-distance pollen flow. On the contrary, S. xanthophylla and Parashorea tomentella had significant FSGS, probably because they were pollinated by small pollinators and consequently short distances of pollen flow. Lee et al. studied the FSGS of *S. parvifolia* populations in different habitats^[22]. They found that populations in montane rainforests, which were mainly pollinated by large pollinators, had no FSGS, whereas populations in lowland rainforests, which were pollinated by small pollinators, had significant FSGS. Lee et al. suggested that it is the restricted pollen flow that leads to strong FSGS in the lowland dipterocarp rainforests^[22]. However, as the distance of pollen-mediated gene flow of V. mangachapoi is unclear at present, the relative contribution of seed and pollen dispersal to the random distribution of genotypes in space deserve further studies.

Implications for conservation

In this study, we demonstrated that there was no significant difference in genetic diversity among the two coastal *V. mangachapoi* populations and one rainforest population near the coast. Moreover, little differentiation and frequent gene flow were detected between the three populations. Even though the coastal populations maintain a relatively high level of genetic diversity, the extremely simple community structure and poor species richness of the coastal *V. mangachapoi*-dominated forests indicate that this unique community is much more fragile than other dipterocarp communities^[55,56]. In addition, the coastal *V. mangachapoi*-dominated forest is likely to

degrade into a sandy scrub community or bare sandy beach in the near future if intense anthropogenic disturbance persists. Therefore, further logging and invasion of the coastal forest must be strictly prohibited, and saplings of *V. mangachapoi* should be planted in forest gaps to promote the restoration of the coastal *V. mangachapoi*-dominated forest landscape.

Conclusion

The fragmented coastal V. mangachapoi-dominated forest in Shimei Bay have not yet exhibited significant genetic differentiation and diversity loss. The winged fruits of V. mangachapoi may promote seed dispersal and maintain gene flow between populations, which could mitigate genetic drift and lead to random distribution of genotypes within population. In addition, comparing with the generation time of V. mangachapoi, the time of fragmentation of the coastal V. mangachapoi forest is relatively short, so there is not enough time to accumulate differentiation for populations from the fragmented forest. Based on the above findings, we suggest to strengthen the protection of the coastal V. mangachapoi forest to prevent further deforestation. Besides, saplings of V. mangachapoi should be planted to connect isolated populations and facilitate the restoration of the unique coastal V. mangachapoi forest.

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Conflict of interest

The authors declare that they have no conflict of interest.

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References

- Ghazoul J. 2016. Dipterocarp Biology, Ecology, and Conservation. 1st Edition. UK: Oxford University Press. https://doi.org/10.1093/ac prof:oso/9780199639656.001.0001
- Ashton PS. 1988. Dipterocarp biology as a window to the understanding of tropical forest structure. *Annual Review of Ecology and Systematics* 19:347–70
- Sodhi NS, Posa MRC, Lee TM, Bickford D, Koh LP, et al. 2010. The state and conservation of Southeast Asian biodiversity. *Biodiver*sity and Conservation 19:317–28
- Edwards DP, Tobias JA, Sheil D, Meijaard E, Laurance WF. 2014. Maintaining ecosystem function and services in logged tropical forests. *Trends in Ecology and Evolution* 29:511–20

- 5. Hu YJ, Li YX. 1992. *The Tropical of Rain Forest of Hainan Island*. 1st Edition. Guangdong, China: Guangdong Higher Education Press
- 6. Hu YJ. 1983. The phytocoenological features and types of Dipterocap forest in Hainan island. *Ecological Science* 2:16–24
- Xu H, Li YD, Luo SS, Chen DX, Lin MX. 2007. Review of Vatica mangachapoi, a national key protected plant on Hainan Island. Tropical Forestry 35:8–11
- Liang SQ, Lin Y, Yang XB, Huang SM, Fu SX, et al. 1993. The Vatica hainanensis Forest of Li Ji, Wan Ning County, Hainan Island. Natural Science Journal of Hainan University 11:1–9
- Yang XB, Hu RG. 2000. The floral components and soil properties of forest on the tropical sandy beach. *Chinese Journal of Ecology* 19:6–11
- Hu RG, Liang SQ, Lin Y. 1995. A study on the soil properties of Vatica mangachapoi forest in Li Ji, Wan Ning County, Hainan Province. Natural Science Journal of Hainan University 13:203–10
- Wu YH, Huang QM, Liang SQ, Lin Y. 1996. Disease and pest control of *Vatica mangachapoi* forest in Liji Town, Wanning City, Hainan Province. *Tropical Forestry* 24:47–51
- Wang G, Zhao JM, Hao QY. 2012. Landscape fragmentation of coastal Vatica mangachapoi forest nature reserve in Shimei bay. *Guangdong Agricultural Sciences* 39:171–4
- Kwak MM, Velterop O, Van Andel J. 1998. Pollen and gene flow in fragmented habitats. *Applied Vegetation Science* 1:37–54
- Kolb A, Diekmann M. 2005. Effects of life-history traits on responses of plant species to forest fragmentation. *Conservation Biology* 19:929–38
- Lowe AJ, Boshier D, Ward M, Bacles CFE, Navarro C. 2005. Genetic resource impacts of habitat loss and degradation; reconciling empirical evidence and predicted theory for neotropical trees. *Heredity* 95:255–73
- Ward M, Dick CW, Gribel R, Lowe AJ. 2005. To self, or not to self... A review of outcrossing and pollen-mediated gene flow in neotropical trees. *Heredity* 95:246–54
- Vekemans X, Hardy OJ. 2004. New insights from fine-scale spatial genetic structure analyses in plant populations. *Molecular Ecology* 13:921–35
- Sebbenn AM, Carvalho ACM, Freitas MLM, Moraes SMB, Gaino APSC, et al. 2011. Low levels of realized seed and pollen gene flow and strong spatial genetic structure in a small, isolated and fragmented population of the tropical tree *Copaifera langsdorffii* Desf. *Heredity* 106:134–45
- Suzuki E, Ashton PS. 1996. Sepal and nut size ratio of fruits of Asian Dipterocarpaceae and its implications for dispersal. *Journal of Tropical Ecology* 12:853–70
- Smith JR, Bagchi R, Ellens J, Kettle CJ, Burslem DFRP, et al. 2015. Predicting dispersal of auto-gyrating fruit in tropical trees: a case study from the Dipterocarpaceae. *Ecology and Evolution* 5(9):1794–801
- Momose K, Yumoto T, Nagamitsu T, Kato M, Nagamasu H, et al. 1998. Pollination biology in a lowland dipterocarp forest in Sarawak, Malaysia. I. Characteristics of the plant-pollinator community in a lowland dipterocarp forest. *American Journal of Botany* 10:1477–501
- Lee SL, Ng KKS, Ng CH, Tnah LH, Lee CT, et al. 2016. Spatial studies of Shorea parvifolia ssp parvifolia (Dipterocarpaceae) in a lowland and hill dipterocarp forest. Journal of Tropical Forest Science 28:309–17
- Kettle CJ, Maycock CR, Ghazoul J, Hollingsworth PM, Khoo E, et al. 2011. Ecological implications of a flower size/number trade-off in tropical forest trees. *PLoS One* 6:e16111
- Harata T, Nanami S, Yamakura T, Matsuyama S, Chong L, et al. 2012. Fine-scale spatial genetic structure of ten Dipterocarp tree species in a Bornean rain forest. *Biotropica* 44:586–94
- 25. Finger A, Kettle CJ, Kaiser-Bunbury CN, Valentin T, Mougal J, et al. 2012. Forest fragmentation genetics in a formerly widespread

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island endemic tree: Vateriopsis seychellarum (Dipterocarpaceae). Molecular Ecology 21:2369–82

- 26. Widiyatno, Indrioko S, Na'iem M, Purnomo S, Hosaka T, et al. 2017. Effects of logging rotation in a lowland dipterocarp forest on mating system and gene flow in *Shorea parvifolia*. *Tree Genetics and Genomes* 13:85
- 27. Doyle JJ. 1990. Isolation of Plant DNA from Fresh Tissue. *Focus* 12:13–15
- 28. Guo JJ, Shang SB, Wang CS, Zhao ZG, Zeng J. 2017. Twenty microsatellite markers for the endangered *Vatica mangachapoi* (Dipterocarpaceae). *Applications in Plant Sciences* 5:1600134
- Peakall R, Smouse PE. 2006. GENALEX 6: genetic analysis in Excel. Population genetic software for teaching and research. *Molecular Ecology Notes*, 6:288–95
- 30. Nei M, Chesser RK. 1983. Estimation of fixation indices and gene diversities. *Annals of Human Genetics* 47:253–59
- 31. Liu K, Muse SV. 2005. PowerMarker: an integrated analysis environment for genetic marker analysis. *Bioinformatics* 21:2128–29
- 32. Excoffier L, Lischer HEL. 2010. Arlequin suite ver 3.5: a new series of programs to perform population genetics analyses under Linux and Windows. *Molecular Ecology Resources* 10:564–7
- 33. Piry S, Luikart G, Cornuet JM. 1999. BOTTLENECK: a computer program for detecting recent reductions in the effective size using allele frequency data. *Journal of Heredity* 90:502–3
- Falush D, Stephens M, Pritchard JK. 2003. Inference of population structure using multilocus genotype data: linked loci and correlated allele frequencies. *Genetics* 164:1567–87
- Earl DA, VonHoldt BM. 2012. STRUCTURE HARVESTER: a website and program for visualizing STRUCTURE output and implementing the Evanno method. *Conservation Genetics Resources* 4:359–61
- Jakobsson M, Rosenberg NA. 2007. CLUMPP: a cluster matching and permutation program for dealing with label switching and multimodality in analysis of population structure. *Bioinformatics* 23:1801–6
- Rosenberg NA. 2004. DISTRUCT: a program for the graphical display of population structure. *Molecular Ecology Notes* 4:137–38
- Tamura K, Stecher G, Kumar S. 2021. MEGA11: Molecular Evolutionary Genetics Analysis Version 11. *Molecular Biology and Evolution* 38:3022–27
- Beerli P. 2006. Comparison of Bayesian and maximum-likelihood inference of population genetic parameters. *Bioinformatics* 22:341–5
- 40. Hardy OJ, Vekemans X. 2002. SPAGeDi: a versatile computer program to analyse spatial genetic structure at the individual or population levels. *Molecular Ecology Notes* 2:618–20
- Loiselle BA, Sork VL, Nason J, Graham C. 1995. Spatial genetic structure of a tropical understory shrub, *Psychotria officinalis* (Rubiaceae). *American Journal of Botany* 82:1420–25
- 42. Doligez A, Joly HI. 1997. Genetic diversity and spatial structure within a natural stand of a tropical forest tree species, *Carapa procera* (Meliaceae), in French Guiana. *Heredity* 79:72–82

- 43. Cavers S, Degen B, Caron H, Lemes MR, Margis R, et al. 2005. Optimal sampling strategy for estimation of spatial genetic structure in tree populations. *Heredity* 95:281–89
- 44. Chen W, Qi H, Fu R, Li D, Jiang D. 2021. Survey and population dynamics analysis of *Vatica mangachapoi* germplasm resource in Hainan Island. *Molecular Plant Breeding* 14:4846–54
- Hamrick JL, Godt MJW, Sherman-Broyles SL. 1992. Factors influencing levels of genetic diversity in woody plant species. *New Forest* 6:95–124
- 46. Ony MA, Nowicki M, Boggess SL, Klingeman WE, Zobel JM, et al. 2020. Habitat fragmentation influences genetic diversity and differentiation: Fine-scale population structure of *Cercis canadensis* (eastern redbud). *Ecology and Evolution* 10:3655–70
- Bacles CFE, Burczyk J, Lowe AJ, Ennos RA. 2005. Historical and contemporary mating patterns in remnant populations of the forest tree *Fraxinus excelsior* L. *Evolution* 59:979–90
- Kikuchi S, Shibata M, Tanaka H. 2015. Effects of forest fragmentation on the mating system of a cool-temperate heterodichogamous tree Acer mono. Global Ecology and Conservation 3:789–801
- 49. Tito de Morais C, Ghazoul J, Maycock CR, Bagchi R, Burslem DFRP, et al. 2015. Understanding local patterns of genetic diversity in dipterocarps using a multi-site, multi-species approach: Implications for forest management and restoration. *Forest Ecology and Management* 356:153–65
- Smith JR, Ghazoul J, Burslem DFRP, Itoh A, Khoo E, et al. 2018. Are patterns of fine-scale spatial genetic structure consistent between sites within tropical tree species? *PLoS One* 13(3):e0193501
- 51. Lü B, Wang N, Liu S, Hao Q. 2015. Reproductive phenological characteristics of hainan coastal *Vatica mangachapoi* forest. *Acta Ecologica Sinica* 35:416–23
- 52. Tackenberg O. 2003. Modeling long-distance dispersal of plant diaspores by wind. *Ecological Monographs* 73:173–89
- Maurer KD, Bohrer G, Medvigy D, Wright SJ. 2013. The timing of abscission affects dispersal distance in a wind-dispersed tropical tree. *Functional Ecology* 27:208–18
- 54. Kettle CJ, Hollingsworth PM, Burslem DFRP, Maycock CR, Khoo E, et al. 2011. Determinants of fine-scale spatial genetic structure in three co-occurring rain forest canopy trees in Borneo. *Perspectives in Plant Ecology, Evolution and Systematics* 13:47–56
- 55. Wang B. 1987. Approach to the horizontal zonation of monsoon forests. *Acta Phytoecologica et Geobotanica Sinica* 11:154–58
- Liang S, Lin Y, Yang X, Huang S, Fu S, et al. 1994. The Vatica mangachapoi Forest of Li Ji, Wan Ning County, Hainan Island. Natural Science Journal of Hainan University 12:14–19

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